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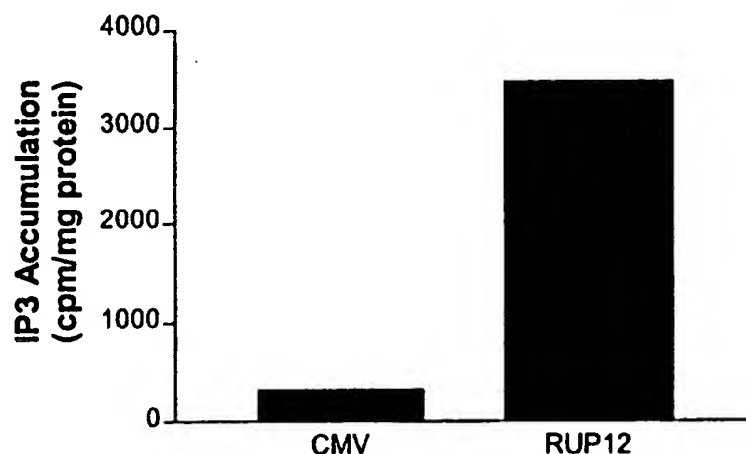
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(54) Title: ENDOGENOUS AND NON-ENDOGENOUS VERSIONS OF HUMAN G PROTEIN-COUPLED RECEPTORS

IP3 Assay in 293 Cells

(57) Abstract: The invention disclosed in this patent document relates to transmembrane receptors, more particularly to a human G protein-coupled receptor for which the endogenous ligand is unknown ("orphan GPCR receptors"), and most particularly to mutated (non-endogenous) versions of the human GPCRs for evidence of constitutive activity.

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ENDOGENOUS AND NON-ENDOGENOUS VERSIONS OF HUMAN G PROTEIN-COUPLED RECEPTORS

FIELD OF THE INVENTION

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The invention disclosed in this patent document relates to transmembrane receptors, and more particularly to human G protein-coupled receptors, and specifically to endogenous human GPCRs with particular emphasis on non-endogenous versions of the GPCRs that have been altered to establish or enhance

10 constitutive activity of the receptor. Preferably, the altered GPCRs are used for the direct identification of candidate compounds as receptor agonists, inverse agonists or partial agonists having potential applicability as therapeutic agents.

15

BACKGROUND OF THE INVENTION

Although a number of receptor classes exist in humans, by far the most abundant and therapeutically relevant is represented by the G protein-coupled receptor (GPCR or GPCRs) class. It is estimated that there are some 100,000 genes within the human

20 genome, and of these, approximately 2%, or 2,000 genes, are estimated to code for GPCRs. Receptors, including GPCRs, for which the endogenous ligand has been identified are referred to as "known" receptors, while receptors for which the endogenous ligand has not been identified are referred to as "orphan" receptors. GPCRs represent an important area for the development of pharmaceutical products: from

25 approximately 20 of the 100 known GPCRs, approximately 60% of all prescription pharmaceuticals have been developed.

GPCRs share a common structural motif. All these receptors have seven sequences of between 22 to 24 hydrophobic amino acids that form seven alpha helices, each of which spans the membrane (each span is identified by number, *i.e.*,

30 transmembrane-1 (TM-1), transmebrane-2 (TM-2), etc.). The transmembrane helices are joined by strands of amino acids between transmembrane-2 and transmembrane-3,

transmembrane-4 and transmembrane-5, and transmembrane-6 and transmembrane-7 on the exterior, or "extracellular" side, of the cell membrane (these are referred to as "extracellular" regions 1, 2 and 3 (EC-1, EC-2 and EC-3), respectively). The transmembrane helices are also joined by strands of amino acids between
5 transmembrane-1 and transmembrane-2, transmembrane-3 and transmembrane-4, and transmembrane-5 and transmembrane-6 on the interior, or "intracellular" side, of the cell membrane (these are referred to as "intracellular" regions 1, 2 and 3 (IC-1, IC-2 and IC-3), respectively). The "carboxy" ("C") terminus of the receptor lies in the intracellular space within the cell, and the "amino" ("N") terminus of the receptor lies in the
10 extracellular space outside of the cell.

Generally, when an endogenous ligand binds with the receptor (often referred to as "activation" of the receptor), there is a change in the conformation of the intracellular region that allows for coupling between the intracellular region and an intracellular "G-protein." It has been reported that GPCRs are "promiscuous" with respect to G proteins,
15 *i.e.*, that a GPCR can interact with more than one G protein. *See*, Kenakin, T., 43 *Life Sciences* 1095 (1988). Although other G proteins exist, currently, Gq, Gs, Gi, Gz and Go are G proteins that have been identified. Endogenous ligand-activated GPCR coupling with the G-protein begins a signaling cascade process (referred to as "signal transduction"). Under normal conditions, signal transduction ultimately results in
20 cellular activation or cellular inhibition. It is thought that the IC-3 loop as well as the carboxy terminus of the receptor interact with the G protein.

Under physiological conditions, GPCRs exist in the cell membrane in equilibrium between two different conformations: an "inactive" state and an "active" state. A receptor in an inactive state is unable to link to the intracellular signaling
25 transduction pathway to produce a biological response. Changing the receptor

conformation to the active state allows linkage to the transduction pathway (via the G-protein) and produces a biological response.

A receptor may be stabilized in an active state by an endogenous ligand or a compound such as a drug. Recent discoveries, including but not exclusively limited to
5 modifications to the amino acid sequence of the receptor, provide means other than endogenous ligands or drugs to promote and stabilize the receptor in the active state conformation. These means effectively stabilize the receptor in an active state by simulating the effect of an endogenous ligand binding to the receptor. Stabilization by such ligand-independent means is termed "constitutive receptor activation."

10

SUMMARY OF THE INVENTION

Disclosed herein are endogenous and non-endogenous versions of human GPCRs and uses thereof.

15

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 provides an illustration of second messenger IP_3 production from endogenous version RUP12 ("RUP12") as compared with the control ("CMV").

Figure 2 is a graphic representation of the results of a second messenger cell-based cyclic AMP assay providing comparative results for constitutive signaling of
20 endogenous RUP13 ("RUP13") and a control vector ("CMV").

Figure 3 is a diagrammatic representation of the signal measured comparing CMV, endogenous RUP13 ("RUP13 wt") and non-endogenous, constitutively activated RUP13 ("RUP13(A268K)"), utilizing 8XCRE-Luc reporter plasmid.

Figure 4 is a graphic representation of the results of a [³⁵S]GTPγS assay providing comparative results for constitutive signaling by RUP13:Gs Fusion Protein ("RUP13-Gs") and a control vector ("CMV").

Figure 5 is a diagrammatic representation of the signal measured comparing
5 CMV, endogenous RUP14 ("RUP14 wt") and non-endogenous, constitutively activated RUP13 ("RUP14(L246K)"), utilizing 8XCRE-Luc reporter plasmid.

Figure 6 is a diagrammatic representation of the signal measured comparing CMV, endogenous RUP15 ("RUP15 wt") and non-endogenous, constitutively activated RUP15 ("RUP15(A398K)"), utilizing 8XCRE-Luc reporter plasmid.

10 **Figure 7** is a graphic representation of the results of a second messenger cell-based cyclic AMP assay providing comparative results for constitutive signaling of endogenous RUP15 ("RUP15 wt"), non-endogenous, constitutively activated version of RUP15 ("RUP15(A398K)") and a control vector ("CMV").

Figure 8 is a graphic representation of the results of a [³⁵S]GTPγS assay
15 providing comparative results for constitutive signaling by RUP15:Gs Fusion Protein ("RUP15-Gs") and a control vector ("CMV").

Figure 9 provides an illustration of second messenger IP₃ production from endogenous version RUP17 ("RUP17") as compared with the control ("CMV").

Figure 10 provides an illustration of second messenger IP₃ production from
20 endogenous version RUP21 ("RUP21") as compared with the control ("CMV").

Figure 11 is a diagrammatic representation of the signal measured comparing CMV, endogenous RUP23 ("RUP23 wt") and non-endogenous, constitutively activated RUP23 ("RUP23(W275K)"), utilizing 8XCRE-Luc reporter plasmid.

Figure 12 is a graphic representation of results from a primary screen of several candidate compounds against RUP13; results for "Compound A" are provided in well A2 and "Compound B" are provided in well G9.

5

DETAILED DESCRIPTION

The scientific literature that has evolved around receptors has adopted a number of terms to refer to ligands having various effects on receptors. For clarity and consistency, the following definitions will be used throughout this patent document. To the extent that these definitions conflict with other definitions for these terms, the following definitions shall control:

10

AGONISTS shall mean materials (*e.g.*, ligands, candidate compounds) that activate the intracellular response when they bind to the receptor, or enhance GTP binding to membranes.

AMINO ACID ABBREVIATIONS used herein are set out in Table A:

TABLE A

ALANINE	ALA	A
ARGININE	ARG	R
ASPARAGINE	ASN	N
ASPARTIC ACID	ASP	D
CYSTEINE	CYS	C
GLUTAMIC ACID	GLU	E
GLUTAMINE	GLN	Q
GLYCINE	GLY	G
HISTIDINE	HIS	H
ISOLEUCINE	ILE	I
LEUCINE	LEU	L
LYSINE	LYS	K
METHIONINE	MET	M

PHENYLALANINE	PHE	F
PROLINE	PRO	P
SERINE	SER	S
THREONINE	THR	T
TRYPTOPHAN	TRP	W
TYROSINE	TYR	Y
VALINE	VAL	V

PARTIAL AGONISTS shall mean materials (*e.g.*, ligands, candidate compounds) that activate the intracellular response when they bind to the receptor to a lesser degree/extent than do agonists, or enhance GTP binding to membranes to a lesser degree/extent than do agonists.

ANTAGONIST shall mean materials (*e.g.*, ligands, candidate compounds) that competitively bind to the receptor at the same site as the agonists but which do not activate the intracellular response initiated by the active form of the receptor, and can thereby inhibit the intracellular responses by agonists or partial agonists.

10 **ANTAGONISTS** do not diminish the baseline intracellular response in the absence of an agonist or partial agonist.

CANDIDATE COMPOUND shall mean a molecule (for example, and not limitation, a chemical compound) that is amenable to a screening technique. Preferably, the phrase "candidate compound" does not include compounds which were publicly known to be compounds selected from the group consisting of inverse agonist, agonist or antagonist to a receptor, as previously determined by an indirect identification process ("indirectly identified compound"); more preferably, not including an indirectly identified compound which has previously been determined to have therapeutic efficacy in at least one mammal; and, most preferably, not including an indirectly identified compound which has previously been determined to have therapeutic utility in humans.

COMPOSITION means a material comprising at least one component; a "pharmaceutical composition" is an example of a composition.

COMPOUND EFFICACY shall mean a measurement of the ability of a compound to inhibit or stimulate receptor functionality, as opposed to receptor binding
5 affinity. Exemplary means of detecting compound efficacy are disclosed in the Example section of this patent document.

CODON shall mean a grouping of three nucleotides (or equivalents to nucleotides) which generally comprise a nucleoside (adenosine (A), guanosine (G), cytidine (C), uridine (U) and thymidine (T)) coupled to a phosphate group and which,
10 when translated, encodes an amino acid.

CONSTITUTIVELY ACTIVATED RECEPTOR shall mean a receptor subject to constitutive receptor activation. A constitutively activated receptor can be endogenous or non-endogenous.

CONSTITUTIVE RECEPTOR ACTIVATION shall mean stabilization of a
15 receptor in the active state by means other than binding of the receptor with its endogenous ligand or a chemical equivalent thereof.

CONTACT or **CONTACTING** shall mean bringing at least two moieties together, whether in an in vitro system or an in vivo system.

DIRECTLY IDENTIFYING or **DIRECTLY IDENTIFIED**, in relationship to
20 the phrase "candidate compound", shall mean the screening of a candidate compound against a constitutively activated receptor, preferably a constitutively activated orphan receptor, and most preferably against a constitutively activated G protein-coupled cell surface orphan receptor, and assessing the compound efficacy of such compound. This phrase is, under no circumstances, to be interpreted or understood to be encompassed by
25 or to encompass the phrase "indirectly identifying" or "indirectly identified."

ENDOGENOUS shall mean a material that a mammal naturally produces. **ENDOGENOUS** in reference to, for example and not limitation, the term "receptor," shall mean that which is naturally produced by a mammal (for example, and not limitation, a human) or a virus. By contrast, the term **NON-ENDOGENOUS** in this context shall mean that which is not naturally produced by a mammal (for example, and not limitation, a human) or a virus. For example, and not limitation, a receptor which is not constitutively active in its endogenous form, but when manipulated becomes constitutively active, is most preferably referred to herein as a "non-endogenous, constitutively activated receptor." Both terms can be utilized to describe both "in vivo" and "in vitro" systems. For example, and not limitation, in a screening approach, the endogenous or non-endogenous receptor may be in reference to an in vitro screening system. As a further example and not limitation, where the genome of a mammal has been manipulated to include a non-endogenous constitutively activated receptor, screening of a candidate compound by means of an in vivo system is viable.

G PROTEIN COUPLED RECEPTOR FUSION PROTEIN and **GPCR FUSION PROTEIN**, in the context of the invention disclosed herein, each mean a non-endogenous protein comprising an endogenous, constitutively activate GPCR or a non-endogenous, constitutively activated GPCR fused to at least one G protein, most preferably the alpha (α) subunit of such G protein (this being the subunit that binds GTP), with the G protein preferably being of the same type as the G protein that naturally couples with endogenous orphan GPCR. For example, and not limitation, in an endogenous state, if the G protein "G α " is the predominate G protein that couples with the GPCR, a GPCR Fusion Protein based upon the specific GPCR would be a non-endogenous protein comprising the GPCR fused to G α ; in some circumstances, as will be set forth below, a non-predominant G protein can be fused to the GPCR. The G

protein can be fused directly to the c-terminus of the constitutively active GPCR or there may be spacers between the two.

HOST CELL shall mean a cell capable of having a Plasmid and/or Vector incorporated therein. In the case of a prokaryotic Host Cell, a Plasmid is typically replicated as a autonomous molecule as the Host Cell replicates (generally, the Plasmid is thereafter isolated for introduction into a eukaryotic Host Cell); in the case of a eukaryotic Host Cell, a Plasmid is integrated into the cellular DNA of the Host Cell such that when the eukaryotic Host Cell replicates, the Plasmid replicates. Preferably, for the purposes of the invention disclosed herein, the Host Cell is eukaryotic, more preferably, mammalian, and most preferably selected from the group consisting of 293, 293T and COS-7 cells.

INDIRECTLY IDENTIFYING or **INDIRECTLY IDENTIFIED** means the traditional approach to the drug discovery process involving identification of an endogenous ligand specific for an endogenous receptor, screening of candidate compounds against the receptor for determination of those which interfere and/or compete with the ligand-receptor interaction, and assessing the efficacy of the compound for affecting at least one second messenger pathway associated with the activated receptor.

INHIBIT or **INHIBITING**, in relationship to the term "response" shall mean that a response is decreased or prevented in the presence of a compound as opposed to in the absence of the compound.

INVERSE AGONISTS shall mean materials (e.g., ligand, candidate compound) which bind to either the endogenous form of the receptor or to the constitutively activated form of the receptor, and which inhibit the baseline intracellular response initiated by the active form of the receptor below the normal base level of activity which

is observed in the absence of agonists or partial agonists, or decrease GTP binding to membranes. Preferably, the baseline intracellular response is inhibited in the presence of the inverse agonist by at least 30%, more preferably by at least 50%, and most preferably by at least 75%, as compared with the baseline response in the absence of the inverse
5 agonist.

KNOWN RECEPTOR shall mean an endogenous receptor for which the endogenous ligand specific for that receptor has been identified.

LIGAND shall mean an endogenous, naturally occurring molecule specific for an endogenous, naturally occurring receptor.

10 **MUTANT** or **MUTATION** in reference to an endogenous receptor's nucleic acid and/or amino acid sequence shall mean a specified change or changes to such endogenous sequences such that a mutated form of an endogenous, non-constitutively activated receptor evidences constitutive activation of the receptor. In terms of equivalents to specific sequences, a subsequent mutated form of a human receptor is
15 considered to be equivalent to a first mutation of the human receptor if (a) the level of constitutive activation of the subsequent mutated form of a human receptor is substantially the same as that evidenced by the first mutation of the receptor; and (b) the percent sequence (amino acid and/or nucleic acid) homology between the subsequent mutated form of the receptor and the first mutation of the receptor is at least about 80%,
20 more preferably at least about 90% and most preferably at least 95%. Ideally, and owing to the fact that the most preferred cassettes disclosed herein for achieving constitutive activation includes a single amino acid and/or codon change between the endogenous and the non-endogenous forms of the GPCR, the percent sequence homology should be at least 98%.

NON-ORPHAN RECEPTOR shall mean an endogenous naturally occurring molecule specific for an endogenous naturally occurring ligand wherein the binding of a ligand to a receptor activates an intracellular signaling pathway.

ORPHAN RECEPTOR shall mean an endogenous receptor for which the
5 endogenous ligand specific for that receptor has not been identified or is not known.

PHARMACEUTICAL COMPOSITION shall mean a composition comprising at least one active ingredient, whereby the composition is amenable to investigation for a specified, efficacious outcome in a mammal (for example, and not limitation, a human). Those of ordinary skill in the art will understand and appreciate the
10 techniques appropriate for determining whether an active ingredient has a desired efficacious outcome based upon the needs of the artisan.

PLASMID shall mean the combination of a Vector and cDNA. Generally, a Plasmid is introduced into a Host Cell for the purposes of replication and/or expression of the cDNA as a protein.

15 **SECOND MESSENGER** shall mean an intracellular response produced as a result of receptor activation. A second messenger can include, for example, inositol triphosphate (IP₃), diacylglycerol (DAG), cyclic AMP (cAMP), and cyclic GMP (cGMP). Second messenger response can be measured for a determination of receptor activation. In addition, second messenger response can be measured for the direct
20 identification of candidate compounds, including for example, inverse agonists, agonists, partial agonists and antagonists.

STIMULATE or **STIMULATING**, in relationship to the term "response" shall mean that a response is increased in the presence of a compound as opposed to in the absence of the compound.

VECTOR in reference to cDNA shall mean a circular DNA capable of incorporating at least one cDNA and capable of incorporation into a Host Cell.

The order of the following sections is set forth for presentational efficiency and is not intended, nor should be construed, as a limitation on the disclosure or the claims to follow.

A. Introduction

The traditional study of receptors has always proceeded from the a priori assumption (historically based) that the endogenous ligand must first be identified before discovery could proceed to find antagonists and other molecules that could affect the receptor. Even in cases where an antagonist might have been known first, the search immediately extended to looking for the endogenous ligand. This mode of thinking has persisted in receptor research even after the discovery of constitutively activated receptors. What has not been heretofore recognized is that it is the active state of the receptor that is most useful for discovering agonists, partial agonists, and inverse agonists of the receptor. For those diseases which result from an overly active receptor or an under-active receptor, what is desired in a therapeutic drug is a compound which acts to diminish the active state of a receptor or enhance the activity of the receptor, respectively, not necessarily a drug which is an antagonist to the endogenous ligand. This is because a compound that reduces or enhances the activity of the active receptor state need not bind at the same site as the endogenous ligand. Thus, as taught by a method of this invention, any search for therapeutic compounds should start by screening compounds against the ligand-independent active state.

B. Identification of Human GPCRs

The efforts of the Human Genome project has led to the identification of a plethora of information regarding nucleic acid sequences located within the human genome; it has been the case in this endeavor that genetic sequence information has been made available without an understanding or recognition as to whether or not any particular genomic sequence does or may contain open-reading frame information that translate human proteins. Several methods of identifying nucleic acid sequences within the human genome are within the purview of those having ordinary skill in the art. For example, and not limitation, a variety of human GPCRs, disclosed herein, were discovered by reviewing the GenBank™ database. Table B, below, lists several endogenous GPCRs that we have discovered, along with other GPCR's that are homologous to the disclosed GPCR.

TABLE B

Disclosed Human Orphan GPCRs	Accession Number Identified	Open Reading Frame (Base Pairs)	Reference To Homologous GPCR	Per Cent Homology To Designated GPCR
hRUP8	AL121755	1,152bp	NPY2R	27%
hRUP9	AC0113375	1,260bp	GAL2R	22%
hRUP10	AC008745	1,014bp	C5aR	40%
hRUP11	AC013396	1,272bp	HM74	36%
hRUP12	AP000808	966bp	Mas1	34%
hRUP13	AC011780	1,356bp	Fish GPRX-ORYLA	43%
hRUP14	AL137118	1,041bp	CysLT1R	35%
hRUP15	AL016468	1,527bp	RE2	30%
hRUP16	AL136106	1,068bp	GLR101	37%
hRUP17	AC023078	969bp	Mas1	37%
hRUP18	AC008547	1,305bp	Oxytocin	31%
hRUP19	AC026331	1,041bp	HM74	52%
hRUP20	AL161458	1,011bp	GPR34	25%
hRUP21	AC026756	1,014bp	P2Y1R	37%
hRUP22	AC027026	993bp	RUP17 Mas1	67% 37%

hRUP23	AC007104	1,092bp	Rat GPR26	31%
hRUP24	AL355388	1,125bp	SALPR	44%
hRUP25	AC026331	1,092bp	HM74	95%
hRUP26	AC023040	1,044bp	Rabbit 5HT1D	27%
hRUP27	AC027643	158,700	MCH	38%

Receptor homology is useful in terms of gaining an appreciation of a role of the receptors within the human body. As the patent document progresses, we will disclose techniques for mutating these receptors to establish non-endogenous, constitutively
5 activated versions of these receptors.

The techniques disclosed herein have also been applied to other human, orphan GPCRs known to the art, as will be apparent as the patent document progresses.

C. Receptor Screening

10 Screening candidate compounds against a non-endogenous, constitutively activated version of the human GPCRs disclosed herein allows for the direct identification of candidate compounds which act at this cell surface receptor, without requiring use of the receptor's endogenous ligand. Using routine, and often commercially available techniques, one can determine areas within the body where the
15 endogenous version of human GPCRs disclosed herein is expressed and/or over-expressed. It is also possible using these techniques to determine related disease/disorder states which are associated with the expression and/or over-expression of the receptor; such an approach is disclosed in this patent document.

With respect to creation of a mutation that may evidence constitutive activation
20 of the human GPCR disclosed herein is based upon the distance from the proline residue at which is presumed to be located within TM6 of the GPCR; this algorithmic technique is disclosed in co-pending and commonly assigned patent document PCT Application

Number PCT/US99/23938, published as WO 00/22129 on April 20, 2000, which, along with the other patent documents listed herein, is incorporated herein by reference. The algorithmic technique is not predicated upon traditional sequence "alignment" but rather a specified distance from the aforementioned TM6 proline residue (or, of course, endogenous constitutive substitution for such proline residue). By mutating the amino acid residue located 16 amino acid residues from this residue (presumably located in the IC3 region of the receptor) to, most preferably, a lysine residue, such activation may be obtained. Other amino acid residues may be useful in the mutation at this position to achieve this objective.

10

D. Disease/Disorder Identification and/or Selection

As will be set forth in greater detail below, most preferably inverse agonists and agonists to the non-endogenous, constitutively activated GPCR can be identified by the methodologies of this invention. Such inverse agonists and agonists are ideal candidates as lead compounds in drug discovery programs for treating diseases related to this receptor. Because of the ability to directly identify inverse agonists to the GPCR, thereby allowing for the development of pharmaceutical compositions, a search for diseases and disorders associated with the GPCR is relevant. For example, scanning both diseased and normal tissue samples for the presence of the GPCR now becomes more than an academic exercise or one which might be pursued along the path of identifying an endogenous ligand to the specific GPCR. Tissue scans can be conducted across a broad range of healthy and diseased tissues. Such tissue scans provide a preferred first step in associating a specific receptor with a disease and/or disorder.

Preferably, the DNA sequence of the human GPCR is used to make a probe for (a) dot-blot analysis against tissue-mRNA, and/or (b) RT-PCR identification of the expression of the receptor in tissue samples. The presence of a receptor in a tissue

source, or a diseased tissue, or the presence of the receptor at elevated concentrations in diseased tissue compared to a normal tissue, can be preferably utilized to identify a correlation with a treatment regimen, including but not limited to, a disease associated with that disease. Receptors can equally well be localized to regions of organs by this technique. Based on the known functions of the specific tissues to which the receptor is localized, the putative functional role of the receptor can be deduced.

E. Screening of Candidate Compounds

1. Generic GPCR screening assay techniques

When a G protein receptor becomes constitutively active, it binds to a G protein (*e.g.*, Gq, Gs, Gi, Gz, Go) and stimulates the binding of GTP to the G protein. The G protein then acts as a GTPase and slowly hydrolyzes the GTP to GDP, whereby the receptor, under normal conditions, becomes deactivated. However, constitutively activated receptors continue to exchange GDP to GTP. A non-hydrolyzable analog of GTP, [³⁵S]GTPγS, can be used to monitor enhanced binding to membranes which express constitutively activated receptors. It is reported that [³⁵S]GTPγS can be used to monitor G protein coupling to membranes in the absence and presence of ligand. An example of this monitoring, among other examples well-known and available to those in the art, was reported by Traynor and Nahorski in 1995. The preferred use of this assay system is for initial screening of candidate compounds because the system is generically applicable to all G protein-coupled receptors regardless of the particular G protein that interacts with the intracellular domain of the receptor.

2. Specific GPCR screening assay techniques

Once candidate compounds are identified using the “generic” G protein-coupled receptor assay (*i.e.*, an assay to select compounds that are agonists, partial agonists, or inverse agonists), further screening to confirm that the compounds have interacted at the

receptor site is preferred. For example, a compound identified by the “generic” assay may not bind to the receptor, but may instead merely “uncouple” the G protein from the intracellular domain.

a. Gs, Gz and Gi.

5 Gs stimulates the enzyme adenylyl cyclase. Gi (and Gz and Go), on the other hand, inhibit this enzyme. Adenylyl cyclase catalyzes the conversion of ATP to cAMP; thus, constitutively activated GPCRs that couple the Gs protein are associated with increased cellular levels of cAMP. On the other hand, constitutively activated GPCRs that couple Gi (or Gz, Go) protein are associated with decreased cellular levels of cAMP.

10 *See, generally, “Indirect Mechanisms of Synaptic Transmission,” Chpt. 8, From Neuron To Brain (3rd Ed.) Nichols, J.G. et al eds. Sinauer Associates, Inc. (1992).* Thus, assays that detect cAMP can be utilized to determine if a candidate compound is, *e.g.*, an inverse agonist to the receptor (*i.e.*, such a compound would decrease the levels of cAMP). A variety of approaches known in the art for measuring cAMP can be utilized;

15 a most preferred approach relies upon the use of anti-cAMP antibodies in an ELISA-based format. Another type of assay that can be utilized is a whole cell second messenger reporter system assay. Promoters on genes drive the expression of the proteins that a particular gene encodes. Cyclic AMP drives gene expression by promoting the binding of a cAMP-responsive DNA binding protein or transcription

20 factor (CREB) that then binds to the promoter at specific sites called cAMP response elements and drives the expression of the gene. Reporter systems can be constructed which have a promoter containing multiple cAMP response elements before the reporter gene, *e.g.*, β -galactosidase or luciferase. Thus, a constitutively activated Gs-linked receptor causes the accumulation of cAMP that then activates the gene and expression of

the reporter protein. The reporter protein such as β -galactosidase or luciferase can then be detected using standard biochemical assays (Chen et al. 1995).

b. Go and Gq.

5 Gq and Go are associated with activation of the enzyme phospholipase C, which in turn hydrolyzes the phospholipid PIP₂, releasing two intracellular messengers: diacylglycerol (DAG) and inistol 1,4,5-triphoisphate (IP₃). Increased accumulation of IP₃ is associated with activation of Gq- and Go-associated receptors. *See, generally,* "Indirect Mechanisms of Synaptic Transmission," Chpt. 8, From Neuron To Brain (3rd Ed.) Nichols, J.G. et al eds. Sinauer Associates, Inc. (1992). Assays that detect IP₃ accumulation can be utilized to determine if a candidate compound is, *e.g.*, an inverse agonist to a Gq- or Go-associated receptor (*i.e.*, such a compound would decrease the levels of IP₃). Gq-associated receptors can also been examined using an AP1 reporter assay in that Gq-dependent phospholipase C causes activation of genes containing AP1
10 elements; thus, activated Gq-associated receptors will evidence an increase in the expression of such genes, whereby inverse agonists thereto will evidence a decrease in such expression, and agonists will evidence an increase in such expression. Commercially available assays for such detection are available.

3. GPCR Fusion Protein

20 The use of an endogenous, constitutively activate orphan GPCR or a non-endogenous, constitutively activated orphan GPCR, for use in screening of candidate compounds for the direct identification of inverse agonists, agonists and partial agonists provide an interesting screening challenge in that, by definition, the receptor is active even in the absence of an endogenous ligand bound thereto. Thus, in order to
25 differentiate between, *e.g.*, the non-endogenous receptor in the presence of a candidate compound and the non-endogenous receptor in the absence of that compound, with an

aim of such a differentiation to allow for an understanding as to whether such compound may be an inverse agonist, agonist, partial agonist or have no affect on such a receptor, it is preferred that an approach be utilized that can enhance such differentiation. A preferred approach is the use of a GPCR Fusion Protein.

5 Generally, once it is determined that a non-endogenous orphan GPCR has been constitutively activated using the assay techniques set forth above (as well as others), it is possible to determine the predominant G protein that couples with the endogenous GPCR. Coupling of the G protein to the GPCR provides a signaling pathway that can be assessed. Because it is most preferred that screening take place by use of a mammalian
10 expression system, such a system will be expected to have endogenous G protein therein. Thus, by definition, in such a system, the non-endogenous, constitutively activated orphan GPCR will continuously signal. In this regard, it is preferred that this signal be enhanced such that in the presence of, *e.g.*, an inverse agonist to the receptor, it is more likely that it will be able to more readily differentiate, particularly in the context of
15 screening, between the receptor when it is contacted with the inverse agonist.

 The GPCR Fusion Protein is intended to enhance the efficacy of G protein coupling with the non-endogenous GPCR. The GPCR Fusion Protein is preferred for screening with a non-endogenous, constitutively activated GPCR because such an approach increases the signal that is most preferably utilized in such screening
20 techniques. This is important in facilitating a significant "signal to noise" ratio; such a significant ratio is import preferred for the screening of candidate compounds as disclosed herein.

 The construction of a construct useful for expression of a GPCR Fusion Protein is within the purview of those having ordinary skill in the art. Commercially available
25 expression vectors and systems offer a variety of approaches that can fit the particular

needs of an investigator. The criteria of importance for such a GPCR Fusion Protein construct is that the endogenous GPCR sequence and the G protein sequence both be in-frame (preferably, the sequence for the endogenous GPCR is upstream of the G protein sequence) and that the "stop" codon of the GPCR must be deleted or replaced such that upon expression of the GPCR, the G protein can also be expressed. The GPCR can be linked directly to the G protein, or there can be spacer residues between the two (preferably, no more than about 12, although this number can be readily ascertained by one of ordinary skill in the art). We have a preference (based upon convenience) of use of a spacer in that some restriction sites that are not used will, effectively, upon expression, become a spacer. Most preferably, the G protein that couples to the non-endogenous GPCR will have been identified prior to the creation of the GPCR Fusion Protein construct. Because there are only a few G proteins that have been identified, it is preferred that a construct comprising the sequence of the G protein (*i.e.*, a universal G protein construct) be available for insertion of an endogenous GPCR sequence therein; this provides for efficiency in the context of large-scale screening of a variety of different endogenous GPCRs having different sequences.

As noted above, constitutively activated GPCRs that couple to Gi, Gz and Go are expected to inhibit the formation of cAMP making assays based upon these types of GPCRs challenging (*i.e.*, the cAMP signal decreases upon activation thus making the direct identification of, *e.g.*, inverse agonists (which would further decrease this signal), interesting. As will be disclosed herein, we have ascertained that for these types of receptors, it is possible to create a GPCR Fusion Protein that is not based upon the endogenous GPCR's endogenous G protein, in an effort to establish a viable cyclase-based assay. Thus, for example, an endogenous Gi coupled receptor can be fused to a Gs protein – we believe that such a fusion construct, upon expression, "drives" or "forces"

the endogenous GPCR to couple with, *e.g.*, Gs rather than the “natural” Gi protein, such that a cyclase-based assay can be established. Thus, for Gi, Gz and Go coupled receptors, we prefer that when a GPCR Fusion Protein is used and the assay is based upon detection of adenylyl cyclase activity, that the fusion construct be established with Gs (or an equivalent G protein that stimulates the formation of the enzyme adenylyl cyclase).

Equally effective is a G Protein Fusion construct that utilizes a Gq Protein fused with a Gs, Gi, Gz or Go Protein. A most preferred fusion construct can be accomplished with a Gq Protein wherein the first six (6) amino acids of the G-protein α -subunit (“G α q”) is deleted and the last five (5) amino acids at the C-terminal end of G α q is replaced with the corresponding amino acids of the G α of the G protein of interest. For example, a fusion construct can have a Gq (6 amino acid deletion) fused with a Gi Protein, resulting in a “Gq/Gi Fusion Construct”. We believe that this fusion construct will force the endogenous Gi coupled receptor to couple to its non-endogenous G protein, Gq, such that the second messenger, for example, inositol triphosphate or diacylglycerol, can be measured in lieu of cAMP production.

4. Co-transfection of a Target Gi Coupled GPCR with a Signal-Enhancer Gs Coupled GPCR (cAMP Based Assays)

A Gi coupled receptor is known to inhibit adenylyl cyclase, and, therefore, decrease the level of cAMP production, which can make assessment of cAMP levels challenging. An effective technique in measuring the decrease in production of cAMP as an indication of constitutive activation of a receptor that predominantly couples Gi upon activation can be accomplished by co-transfecting a signal enhancer, *e.g.*, a non-endogenous, constitutively activated receptor that predominantly couples with Gs upon activation (*e.g.*, TSHR-A623I, disclosed below), with the Gi linked GPCR. As is

apparent, constitutive activation of a Gs coupled receptor can be determined based upon an increase in production of cAMP. Constitutive activation of a Gi coupled receptor leads to a decrease in production cAMP. Thus, the co-transfection approach is intended to advantageously exploit these "opposite" affects. For example, co-transfection of a non-endogenous, constitutively activated Gs coupled receptor (the "signal enhancer") with the endogenous Gi coupled receptor (the "target receptor") provides a baseline cAMP signal (*i.e.*, although the Gi coupled receptor will decrease cAMP levels, this "decrease" will be relative to the substantial increase in cAMP levels established by constitutively activated Gs coupled signal enhancer). By then co-transfecting the signal enhancer with a constitutively activated version of the target receptor, cAMP would be expected to further decrease (relative to base line) due to the increased functional activity of the Gi target (*i.e.*, which decreases cAMP).

Screening of candidate compounds using a cAMP based assay can then be accomplished, with two provisos: first, relative to the Gi coupled target receptor, "opposite" effects will result, *i.e.*, an inverse agonist of the Gi coupled target receptor will increase the measured cAMP signal, while an agonist of the Gi coupled target receptor will decrease this signal; second, as would be apparent, candidate compounds that are directly identified using this approach should be assessed independently to ensure that these do not target the signal enhancing receptor (this can be done prior to or after screening against the co-transfected receptors).

F. Medicinal Chemistry

Generally, but not always, direct identification of candidate compounds is preferably conducted in conjunction with compounds generated via combinatorial chemistry techniques, whereby thousands of compounds are randomly prepared for such analysis. Generally, the results of such screening will be compounds having

unique core structures; thereafter, these compounds are preferably subjected to additional chemical modification around a preferred core structure(s) to further enhance the medicinal properties thereof. Such techniques are known to those in the art and will not be addressed in detail in this patent document.

5

G. Pharmaceutical compositions

Candidate compounds selected for further development can be formulated into pharmaceutical compositions using techniques well known to those in the art. Suitable pharmaceutically-acceptable carriers are available to those in the art; for example, see
10 Remington's Pharmaceutical Sciences, 16th Edition, 1980, Mack Publishing Co., (Oslo et al., eds.).

H. Other Utility

Although a preferred use of the non-endogenous versions the human GPCRs
15 disclosed herein may be for the direct identification of candidate compounds as inverse agonists, agonists or partial agonists (preferably for use as pharmaceutical agents), these versions of human GPCRs can also be utilized in research settings. For example, *in vitro* and *in vivo* systems incorporating GPCRs can be utilized to further elucidate and understand the roles these receptors play in the human condition, both normal and
20 diseased, as well as understanding the role of constitutive activation as it applies to understanding the signaling cascade. The value in non-endogenous human GPCRs is that their utility as a research tool is enhanced in that, because of their unique features, non-endogenous human GPCRs can be used to understand the role of these receptors in the human body before the endogenous ligand therefore is identified. Other uses of the
25 disclosed receptors will become apparent to those in the art based upon, *inter alia*, a review of this patent document.

EXAMPLES

The following examples are presented for purposes of elucidation, and not limitation, of the present invention. While specific nucleic acid and amino acid sequences are disclosed herein, those of ordinary skill in the art are credited with the ability to make minor modifications to these sequences while achieving the same or substantially similar results reported below. The traditional approach to application or understanding of sequence cassettes from one sequence to another (*e.g.* from rat receptor to human receptor or from human receptor A to human receptor B) is generally predicated upon sequence alignment techniques whereby the sequences are aligned in an effort to determine areas of commonality. The mutational approach disclosed herein does not rely upon this approach but is instead based upon an algorithmic approach and a positional distance from a conserved proline residue located within the TM6 region of human GPCRs. Once this approach is secured, those in the art are credited with the ability to make minor modifications thereto to achieve substantially the same results (*i.e.*, constitutive activation) disclosed herein. Such modified approaches are considered within the purview of this disclosure.

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Example 1

ENDOGENOUS HUMAN GPCRS

1. Identification of Human GPCRs

The disclosed endogenous human GPCRs were identified based upon a review of the GenBank™ database information. While searching the database, the following cDNA clones were identified as evidenced below (Table C).

TABLE C

Disclosed Human Orphan GPCRs	Accession Number Identified	Complete DNA Sequence (Base Pairs)	Open Reading Frame (Base Pairs)	Nucleic Acid SEQ.ID. NO.	Amin Acid SEQ.ID. NO.
hRUP8	AL121755	147,566bp	1,152bp	1	2
hRUP9	AC0113375	143,181bp	1,260bp	3	4
hRUP10	AC008745	94,194bp	1,014bp	5	6
hRUP11	AC013396	155,086bp	1,272bp	7	8
hRUP12	AP000808	177,764bp	966bp	9	10
hRUP13	AC011780	167,819bp	1,356bp	11	12
hRUP14	AL137118	168,297bp	1,041bp	13	14
hRUP15	AL016468	138,828bp	1,527bp	15	16
hRUP16	AL136106	208,042bp	1,068bp	17	18
hRUP17	AC023078	161,735bp	969bp	19	20
hRUP18	AC008547	117,304bp	1,305bp	21	22
hRUP19	AC026331	145,183bp	1,041bp	23	24
hRUP20	AL161458	163,511bp	1,011bp	25	26
hRUP21	AC026756	156,534bp	1,014bp	27	28
hRUP22	AC027026	151,811bp	993bp	29	30
hRUP23	AC007104	200,000bp	1,092bp	31	32
hRUP24	AL355388	190,538bp	1,125bp	33	34
hRUP25	AC026331	145,183bp	1,092bp	35	36
hRUP26	AC023040	178,508bp	1,044bp	37	38
hRUP27	AC027643	158,700bp	1,020bp	39	40

5

2. Full Length Cloning

a. hRUP8 (Seq. Id. Nos. 1 & 2)

The disclosed human RUP8 was identified based upon the use of EST database (*dbEST*) information. While searching the *dbEST*, a cDNA clone with accession number

AL121755 was identified to encode a novel GPCR. The following PCR primers were used for RT-PCR with human testis Marathon-Ready cDNA (Clontech) as templates:

5'-CTTGCAGACATCACCATGGCAGCC-3' (SEQ.ID.NO.:41; sense) and

5'-GTGATGCTCTGAGTACTGGACTGG-3' (SEQ.ID.NO.: 42; antisense).

5 PCR was performed using Advantage cDNA polymerase (Clontech; manufacturing instructions will be followed) in 50ul reaction by the following cycles: 94°C for 30 sec; 94°C for 10 sec; 65°C for 20 sec, 72°C for 1.5 min, and 72°C for 7 min. Cycles 2 through 4 were repeated 35 times.

A 1.2kb PCR fragment was isolated and cloned into the pCRII-TOPO vector
10 (Invitrogen) and sequenced using the ABI Big Dye Terminator kit (P.E. Biosystem).
See, SEQ.ID.NO.:1. The putative amino acid sequence for RUP8 is set forth in
SEQ.ID.NO.:2.

b. hRUP9 (Seq. Id. Nos. 3 & 4)

The disclosed human RUP9 was identified based upon the use of GeneBank
15 database information. While searching the database, a cDNA clone with Accession
Number AC011375 was identified as a human genomic sequence from chromosome
5. The full length RUP9 was cloned by PCR using primers:

5'-GAAGCTGTGAAGAGTGATGC-3' (SEQ.ID.NO.:43; sense),

5'-GTCAGCAATATTGATAAGCAGCAG-3' (SEQ.ID.NO.:44; antisense)

20 and human genomic DNA (Promega) as a template. Taq Plus Precision polymerase
(Stratagene) was used for the amplification in a 100µl reaction with 5% DMSO by the
following cycle with step 2 to step 4 repeated 35 times: 94°C for 1 minute; 94°C for
30 seconds; 56°C for 30 seconds; 72°C for 2 minutes; 72°C for 5 minutes.

A 1.3 Kb PCR fragment was isolated and cloned into the pCRII-TOPO vector
25 (Invitrogen) from 1% agarose gel and completely sequenced using the ABI Big Dye

Terminator kit (P.E. Biosystem). See, SEQ.ID.NO.:3. The putative amino acid sequence for RUP8 is set forth in SEQ.ID.NO.:4. The sequence of RUP9 clones isolated from human genomic DNA matched with the sequence obtained from data base.

c. hRUP10 (Seq. Id. Nos. 5 & 6)

5 The disclosed human RUP10 was identified based upon the use of GenBank database information. While searching the database, a cDNA clone with accession number AC008754 was identified as a human genomic sequence from chromosome 19. The full length RUP10 was cloned by RT-PCR using primers:

5'-CCATGGGGAACGATTCTGTCAGCTACG-3' (SEQ.ID.NO.:45; sense) and

10 5'-GCTATGCCTGAAGCCAGTCTTGTG-3' (SEQ.ID.NO.:46; antisense)

and human leukocyte Marathon-Ready cDNA (Clontech) as a template. Advantage cDNA polymerase (Clontech) was used for the amplification in a 50µl reaction by the following cycle with step 2 to step 4 repeated 35 times: 94°C for 30 seconds; 94°C for 10 seconds; 62°C for 20 seconds; 72°C for 1.5 minutes; 72°C for 7 minutes. A 1.0

15 Kb PCR fragment was isolated and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye Terminator kit (P.E. Biosystem). The nucleic acid sequence of the novel human receptor RUP10 is set forth in SEQ.ID.NO.:5 and the putative amino acid sequence thereof is set forth in SEQ.ID.NO.:6.

20

d. hRUP11 (Seq. Id. Nos. 7 & 8)

The disclosed human RUP11 was identified based upon the use of GenBank database information. While searching the database, a cDNA clone with accession
25 number AC013396 was identified as a human genomic sequence from chromosome 2.

Terminator kit (P.E. Biosystem). See, SEQ.ID.NO.:3. The putative amino acid sequence for RUP8 is set forth in SEQ.ID.NO.:4. The sequence of RUP9 clones isolated from human genomic DNA matched with the sequence obtained from data base.

c. hRUP10 (Seq. Id. Nos. 5 & 6)

5 The disclosed human RUP10 was identified based upon the use of GenBank database information. While searching the database, a cDNA clone with accession number AC008754 was identified as a human genomic sequence from chromosome 19. The full length RUP10 was cloned by RT-PCR using primers:

5'-CCATGGGGAACGATTCTGTCAGCTACG-3' (SEQ.ID.NO.:45; sense) and

10 5'-GCTATGCCTGAAGCCAGTCTTGTG-3' (SEQ.ID.NO.:46; antisense)

and human leukocyte Marathon-Ready cDNA (Clontech) as a template. Advantage cDNA polymerase (Clontech) was used for the amplification in a 50µl reaction by the following cycle with step 2 to step 4 repeated 35 times: 94°C for 30 seconds; 94°C for 10 seconds; 62°C for 20 seconds; 72°C for 1.5 minutes; 72°C for 7 minutes. A 1.0

15 Kb PCR fragment was isolated and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye Terminator kit (P.E. Biosystem). The nucleic acid sequence of the novel human receptor RUP10 is set forth in SEQ.ID.NO.:5 and the putative amino acid sequence thereof is set forth in SEQ.ID.NO.:6.

20.

d. hRUP11 (Seq. Id. Nos. 7 & 8)

The disclosed human RUP11 was identified based upon the use of GenBank database information. While searching the database, a cDNA clone with accession
25 number AC013396 was identified as a human genomic sequence from chromosome 2.

The full length RUP11 was cloned by PCR using primers:

5'-CCAGGATGTTGTGTCACCGTGGTGGC-3' (SEQ.ID.NO.:47; sense),

5'-CACAGCGCTGCAGCCCTGCAGCTGGC-3' (SEQ.ID.NO.:48; antisense)

and human genomic DNA (Clontech) as a template. TaqPlus Precision DNA

5 polymerase (Stratagene) was used for the amplification in a 50 μ l reaction by the

following cycle with step 2 to step 4 repeated 35 times: 94°C for 3 minutes; 94°C for 20

seconds; 67°C for 20 seconds; 72°C for 1.5 minutes; 72°C for 7 minutes. A 1.3 Kb PCR

fragment was isolated and cloned into the pCRII-TOPO vector (Invitrogen) and

completely sequenced using the ABI Big Dye Terminator kit (P.E. Biosystem). The

10 nucleic acid sequence of the novel human receptor RUP11 is set forth in SEQ.ID.NO.:7

and the putative amino acid sequence thereof is set forth in SEQ.ID.NO.:8.

e. hRUP12 (Seq. Id. Nos. 9 & 10)

The disclosed human RUP12 was identified based upon the use of GenBank database. While searching the database, a cDNA clone with accession number

15 AP000808 was identified to encode a new GPCR, having significant homology with rat

RTA and human mas1 oncogene GPCRs. The full length RUP12 was cloned by PCR

using primers:

5'-CTTCCTCTCGTAGGGATGAACCAGAC-3' (SEQ.ID.NO.:49; sense)

5'-CTCGCACAGGTGGGAAGCACCTGTGG-3' (SEQ.ID.NO.:50; antisense)

20 and human genomic DNA (Clontech) as template. TaqPlus Precision DNA polymerase

(Stratagene) was used for the amplification by the following cycle with step 2 to step 4

repeated 35 times: 94°C for 3 min; 94°C for 20 sec; 65°C for 20sec; 72°C for 2 min and

72°C for 7 min. A 1.0kb PCR fragment was isolated and cloned into the pCRII-TOPO

vector (Invitrogen) and completely sequenced using the ABI Big Dye Terminator kit

(P.E. Biosystem) (*see*, SEQ.ID.NO.:9 for nucleic acid sequence and SEQ.ID.NO.:10 for deduced amino acid sequence).

f. hRUP13 (Seq. Id. Nos. 11 & 12)

The disclosed human RUP13 was identified based upon the use of
5 GenBank database. While searching the database, a cDNA clone with accession number
AC011780 was identified to encode a new GPCR, having significant homology with
GPCR fish GPRX-ORYLA. The full length RUP13 was cloned by PCR using primers:
5'-GCCTGTGACAGGAGGTACCCTGG-3' (SEQ.ID.NO.:51; sense)
5'-CATATCCCTCCGAGTGTCCAGCGGC-3' (SEQ.ID.NO.:52; antisense)
10 and human genomic DNA (Clontech) as template. TaqPlus Precision DNA polymerase
(Stratagene) was used for the amplification by the following cycle with step 2 to step 4
repeated 35 times: 94°C for 3 min; 94°C for 20 sec; 65°C for 20sec; 72°C for 2 min and
72°C for 7 min. A 1.35kb PCR fragment was isolated and cloned into the pCRII-TOPO
vector (Invitrogen) and completely sequenced using the ABI Big Dye Terminator kit
15 (P.E. Biosystem) (*see*, SEQ.ID.NO.:11 for nucleic acid sequence and SEQ.ID.NO.:12
for deduced amino acid sequence).

g. hRUP14 (Seq. Id. Nos. 13 & 14)

The disclosed human RUP14 was identified based upon the use of GeneBank
database information. While searching the database, a cDNA clone with Accession
20 Number AL137118 was identified as a human genomic sequence from chromosome
13. The full length RUP14 was cloned by PCR using primers:
5'-GCATGGAGAGAAAATTTATGTCCTTGCAACC-3' (SEQ.ID.NO.:53; sense)
5'-CAAGAACAGGTCTCATCTAAGAGCTCC-3' (SEQ.ID.NO.:54; antisense)
and human genomic DNA (Promega) as a template. Taq Plus Precision polymerase
25 (Stratagene) and 5% DMSO were used for the amplification by the following cycle

with step 2 and step 3 repeated 35 times: 94°C for 3 minute; 94°C for 20 seconds; 58°C for 2 minutes; 72°C for 10 minutes.

A 1.1 Kb PCR fragment was isolated and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye Terminator kit (P.E. Biosystem) (see, SEQ.ID.NO.:13 for nucleic acid sequence and SEQ.ID.NO.:14 for deduced amino acid sequence). The sequence of RUP14 clones isolated from human genomic DNA matched with the sequence obtained from database.

h. hRUP15 (Seq. Id. Nos. 15 & 16)

The disclosed human RUP15 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession Number AC016468 was identified as a human genomic sequence. The full-length RUP15 was cloned by PCR using primers:

5'-GCTGTTGCCATGACGTCCACCTGCAC-3' (SEQ.ID.NO.:55; sense)

5'-GGACAGTTCAAGGTTTGCCTTAGAAC-3' (SEQ.ID.NO.:56; antisense)

and human genomic DNA (Promega) as a template. Taq Plus Precision polymerase (Stratagene) was used for the amplification by the following cycle with step 2 to 4 repeated 35 times: 94°C for 3 minute; 94°C for 20 seconds; 65°C for 20 seconds; 72°C for 2 minutes and 72°C for 7 minutes.

A 1.5 Kb PCR fragment was isolated and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye Terminator kit (P.E. Biosystem). See, SEQ.ID.NO.:15 for nucleic acid sequence and SEQ.ID.NO.:16 for deduced amino acid sequence. The sequence of RUP15 clones isolated from human genomic DNA matched with the sequence obtained from database.

i. hRUP16 (Seq. Id. Nos. 17 & 18)

The disclosed human RUP16 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession Number AL136106 was identified as a human genomic sequence from chromosome 13. The full length RUP16 was cloned by PCR using primers:

- 5 5'-CTTTCGATACTGCTCCTATGCTC-3' (SEQ.ID.NO.:57; sense, 5' of initiation codon),
5'-GTAGTCCACTGAAAGTCCAGTGATCC-3' (SEQ.ID.NO.:58; antisense, 3' of stop codon)
and human skeletal muscle Marathon-Ready cDNA (Clontech) as template. Advantage
cDNA polymerase (Clontech) was used for the amplification in a 50ul reaction by the
following cycle with step 2 to 4 repeated 35 times: 94°C for 30 seconds; 94°C for 5
10 seconds; 69°C for 15 seconds; 72°C for 1 minute and 72°C for 5 minutes.

A 1.1 Kb PCR fragment was isolated and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the T7 sequenase kit (Amsham). See, SEQ.ID.NO.:17 for nucleic acid sequence and SEQ.ID.NO.:18 for deduced amino acid sequence. The sequence of RUP16 clones matched with four unordered segments of
15 AL136106, indicating that the RUP16 cDNA is composed of 4 exons.

j. hRUP17 (Seq. Id. Nos. 19 & 20)

- The disclosed human RUP17 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession Number AC023078 was identified as a human genomic sequence from chromosome
20 11. The full length RUP17 was cloned by PCR using primers:
5'-TTTCTGAGCATGGATCCAACCATCTC-3' (SEQ.ID.NO.:59; sense, containing initiation
codon)
5'-CTGTCTGACAGGGCAGAGGCTCTTC-3' (SEQ.ID.NO.:60; antisense, 3' of stop codon)
and human genomic DNA (Promega) as template. Advantage cDNA polymerase mix
25 (Clontech) was used for the amplification in a 100ul reaction with 5% DMSO by the

following cycle with step 2 to 4 repeated 30 times: 94°C for 1 min; 94°C for 15 sec;
67°C for 20 sec; 72°C for 1 min and 30 sec; and 72°C for 5 min.

A 970bp PCR fragment was isolated from 1% agarose gel and cloned into the
pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye
5 Termiantor Kit (P.E. Biosystem). See, SEQ.ID.NO.:19 for nucleic acid sequence and
SEQ.ID.NO.:20 for deduced amino acid sequence.

k. hRUP18 (Seq. Id. Nos. 21 & 22)

The disclosed human RUP18 was identified based upon the use of GeneBank
database information. While searching the database, a cDNA clone with Accession
10 Number AC008547 was identified as a human genomic sequence from chromosome
5. The full length RUP18 was cloned by PCR using primers:

5'-GGAAGTCGTATAGACCCAGCGTCGCTCC-3' (SEQ.ID.NO.:61; sense, 5' of the
initiation codon),

5'-GGAGGTTGCGCCTTAGCGACAGATGACC-3' (SEQ.ID.NO.:62; antisense, 3' of stop
15 codon)

and human genomic DNA (Promega) as template. TaqPlus precision DNA
polymerase (Stratagene) was used for the amplification in a 100ul reaction with 5%
DMSO by the following cycle with step 2 to 4 repeated 35 times: 95°C for 5 min;
95°C for 30 sec; 65°C for 30 sec; 72°C for 2 min; and 72°C for 5 min.

20 A 1.3kb PCR fragment was isolated from 1% agarose gel and cloned into the
pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye
Termiantor Kit (P.E. Biosystem). See, SEQ.ID.NO.:21 for nucleic acid sequence and
SEQ.ID.NO.:22 for deduced amino acid sequence.

l. hRUP19 (Seq. Id. Nos. 23 & 24)

The disclosed human RUP19 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession Number AC026331 was identified as a human genomic sequence from chromosome 12. The full length RUP19 was cloned by PCR using primers:

- 5 5'-CTGCACCCGGACACTTGCTCTG-3' (SEQ.ID.NO.:63; sense, 5' of initiation codon),
5'-GTCTGCTTGTTCAGTGCCACTCAAC-3' (SEQ.ID.NO.:64; antisense, containing the stop codon)

and human genomic DNA (Promega) as template. TaqPlus Precision DNA polymerase (Stratagene) was used for the amplification with 5% DMSO by the following cycle with step 2 to 4 repeated 35 times: 94°C for 1 min; 94°C for 15 sec; 70°C for 20 sec; 72°C for 1 min and 30 sec; and 72°C for 5 min.

A 1.1kp PCR fragment was isolated from 1% agarose gel and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye Terminator Kit (P.E. Biosystem). See, SEQ.ID.NO.:23 for nucleic acid sequence and
15 SEQ.ID.NO.:24 for deduced amino acid sequence.

m. hRUP20 (Seq. Id. Nos. 25 & 26)

The disclosed human RUP20 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession Number AL161458 was identified as a human genomic sequence from chromosome

- 20 1. The full length RUP20 was cloned by PCR using primers:
5'-TATCTGCAATTCTATTCTAGCTCCTG-3' (SEQ.ID.NO.:65; sense, 5' of initiation codon),
5'-TGTCCTAATAAAGTCACATGAATGC-3' (SEQ.ID.NO.:66; antisense, 3' of stop codon)
and human genomic DNA (Promega) as template. Advantage cDNA polymerase mix (Clontech) was used for the amplification with 5% DMSO by the following cycle with

step 2 to 4 repeated 35 times: 94°C for 1 min; 94°C for 15 sec; 60°C for 20 sec; 72°C for 1 min and 30 sec; and 72°C for 5 min.

A 1.0 kp PCR fragment was isolated from 1% agarose gel and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye
5 Termiantor Kit (P.E. Biosystem). See, SEQ.ID.NO.:25 for nucleic acid sequence and SEQ.ID.NO.:26 for deduced amino acid sequence.

n. hRUP21 (Seq. Id. Nos. 27 & 28)

The disclosed human RUP21 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession
10 Number AC026756 was identified as a human genomic sequence from chromosome 13. The full length RUP21 was cloned by PCR using primers:

5'– GGAGACAACCATGAATGAGCCAC –3' (SEQ.ID.NO.:67; sense)

5'– TATTTCAAGGGTTGTTTGAGTAAC –3' (SEQ.ID.NO.:68; antisense)

and human genomic DNA (Promega) as template. Taq Plus Precision polymerase
15 (Stratagene) was used for the amplification in a 100ul reaction with 5% DMSO by the following cycle with step 2 to 4 repeated 30 times: 94°C for 1 min; 94°C for 15 sec; 55°C for 20 sec; 72°C for 1 min and 30 sec; and 72°C for 5 min.

A 1,014 bp PCR fragment was isolated from 1% agarose gel and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye
20 Termiantor Kit (P.E. Biosystem). See, SEQ.ID.NO.:27 for nucleic acid sequence and SEQ.ID.NO.:28 for deduced amino acid sequence.

o. hRUP22 (Seq. Id. Nos. 29 & 30)

The disclosed human RUP22 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession

Number AC027026 was identified as a human genomic sequence from chromosome

11. The full length RUP22 was cloned by PCR using primers:

5'-GGCACCAAGTGGAGGTTTTCTGAGCATG-3' (SEQ.ID.NO.:69; sense, containing initiation codon)

5-CTGATGGAAGTAGAGGCTGTCCATCTC-3' (SEQ.ID.NO.:70; antisense, 3' of stop codon)

and human genomic DNA (Promega) as template. TaqPlus Precision DNA polymerase (Stratagene) was used for the amplification in a 100ul reaction with 5% DMSO by the following cycle with step 2 to 4 repeated 30 times: 94°C, 1 minutes 94°C, 15 seconds

10 55°C, 20 seconds 72°C, 1.5 minute 72°C, 5 minutes.

A 970bp PCR fragment was isolated from 1% agarose gel and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye Terminator Kit (P.E. Biosystem). See, SEQ.ID.NO.:29 for nucleic acid sequence and SEQ.ID.NO.:30 for deduced amino acid sequence.

15 **p. hRUP23 (Seq. Id. Nos. 31 & 32)**

The disclosed human RUP23 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession Number AC007104 was identified as a human genomic sequence from chromosome

4. The full length RUP23 was cloned by PCR using primers:

20 5'-CCTGGCGAGCCGCTAGCGCCATG-3' (SEQ.ID.NO.:71; sense, ATG as the initiation codon),

5'-ATGAGCCCTGCCAGGCCCTCAGT-3' (SEQ.ID.NO.:72; antisense, TCA as the stop codon)

and human placenta Marathon-Ready cDNA (Clontech) as template. Advantage cDNA

25 polymerase (Clontech) was used for the amplification in a 50ul reaction by the following

cycle with step 2 to 4 repeated 35 times: 95°C for 30 sec; 95°C for 15 sec; 66°C for 20 sec; 72°C for 1 min and 20 sec; and 72°C for 5 min.

A 1.0 kb PCR fragment was isolated and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye Terminator Kit (P.E. Biosystem). See, SEQ.ID.NO.:31 for nucleic acid sequence and SEQ.ID.NO.:32 for deduced amino acid sequence.

q. hRUP24 (Seq. Id. Nos. 33 & 34)

The disclosed human RUP25 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession Number AC026331 was identified as a human genomic sequence from chromosome 12. The full length RUP25 was cloned by PCR using primers:
5'-GCTGGAGCATTCACTAGGCGAG-3' (SEQ.ID.NO.:73; sense, 5' of initiation codon),
5'-AGATCCTGGTTCTTGGTGACAATG-3' (SEQ.ID.NO.:74; antisense, 3' of stop codon)
and human genomic DNA (Promega) as template. Advantage cDNA polymerase mix (Clontech) was used for the amplification with 5% DMSO by the following cycle with step 2 to 4 repeated 35 times: 94°C for 1 minute; 94°C for 15 seconds; 56°C for 20 seconds 72°C for 1 minute 30 seconds and 72°C for 5 minutes.

A 1.2kb PCR fragment was isolated from 1% agarose gel and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye Terminator Kit (P.E. Biosystem). See, SEQ.ID.NO.:33 for nucleic acid sequence and SEQ.ID.NO.:34 for deduced amino acid sequence.

r. hRUP25 (Seq. Id. Nos. 35 & 36)

The disclosed human RUP25 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession

Number AC026331 was identified as a human genomic sequence from chromosome 12. The full length RUP25 was cloned by PCR using primers:

5'-GCTGGAGCATTCACTAGGCGAG-3' (SEQ.ID.NO.:75; sense, 5' of initiation codon),

5'-AGATCCTGGTTCTTGGTGACAATG-3' (SEQ.ID.NO.:76; antisense, 3' of stop codon)

5 and human genomic DNA (Promega) as template. Advantage cDNA polymerase mix (Clontech) was used for the amplification with 5% DMSO by the following cycle with step 2 to 4 repeated 35 times: 94°C for 1 minute; 94°C for 15 seconds; 56°C for 20 seconds 72°C for 1 minute 30 seconds and 72°C for 5 minutes.

10 A 1.2kb PCR fragment was isolated from 1% agarose gel and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye Termiantor Kit (P.E. Biosystem). See, SEQ.ID.NO.:35 for nucleic acid sequence and SEQ.ID.NO.:36 for deduced amino acid sequence.

s. hRUP26 (Seq. Id. Nos. 37 & 38)

The disclosed human RUP26 was identified based upon the use of GeneBank 15 database information. While searching the database, a cDNA clone with Accession Number AC023040 was identified as a human genomic sequence from chromosome 2. The full length RUP26 was cloned by RT-PCR using RUP26 specific primers:

5'-AGCCATCCCTGCCAGGAAGCATGG-3' (SEQ.ID.NO.:77; sense, containing initiation codon)

20 5'-CCAGACTGTGGACTCAAGAACTCTAGG-3' (SEQ.ID.NO.:78; antisense, containing stop codon)

and human pancreas Marathon - Ready cDNA (Clontech) as template. Advantage cDNA polymerase mix (Clontech) was used for the amplification in a 100µl reaction with 5% DMSO by the following cycle with step 2 to 4 repeated 35 times: 94°C for 5 minute; 25 95°C for 30 seconds; 65°C for 30 seconds 72°C for 2 minute and 72°C for 5 minutes.

A 1.1kb PCR fragment was isolated from 1% agarose gel and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye Termiantor Kit (P.E. Biosystem). See, SEQ.ID.NO.:37 for nucleic acid sequence and SEQ.ID.NO.:38 for deduced amino acid sequence.

5 **t. hRUP27 (Seq. Id. Nos. 39 & 40)**

The disclosed human RUP27 was identified based upon the use of GeneBank database information. While searching the database, a cDNA clone with Accession Number AC027643 was identified as a human genomic sequence from chromosome 12. The full length RUP27 was cloned by PCR using RUP27 specific primers:

10 5'-AGTCCACGAACAAATGAATCCATTTCATG-3' (SEQ.ID.NO.:79; sense, containing initiation codon),
5'-ATCATGTCTAGACTCATGGTGATCC-3' (SEQ.ID.NO.:80; antisense, 3' of stop codon)
and the human adult brain Marathon-Ready cDNA (Clontech) as template. Advantage cDNA polymerase mix (Clontech) was used for the amplification in a 50µl reaction with 5%
15 DMSO by the following cycle with step 2 to 4 repeated 35 times: 94°C for 1 minute; 94°C for 10 seconds; 58°C for 20 seconds 72°C for 1 minute 30 seconds and 72°C for 5 minutes.

A 1.1kb PCR fragment was isolated from 1% agarose gel and cloned into the pCRII-TOPO vector (Invitrogen) and completely sequenced using the ABI Big Dye
20 Termiantor Kit (P.E. Biosystem). See, SEQ.ID.NO.:35 for nucleic acid sequence and SEQ.ID.NO.:36 for deduced amino acid sequence. The sequence of RUP27 cDNA clone isolated from human brain was determined to match with five unordered segments of AC027643, indicating that the RUP27 cDNA is composed of 5 exons.

Example 2

PREPARATION OF NON-ENDOGENOUS, CONSTITUTIVELY ACTIVATED GPCRS

Those skilled in the art are credited with the ability to select techniques for mutation of a nucleic acid sequence. Presented below are approaches utilized to create non-endogenous versions of several of the human GPCRs disclosed above. The mutations disclosed below are based upon an algorithmic approach whereby the 16th amino acid (located in the IC3 region of the GPCR) from a conserved proline (or an endogenous, conservative substitution therefore) residue (located in the TM6 region of the GPCR, near the TM6/IC3 interface) is mutated, preferably to an alanine, histidine, arginine or lysine amino acid residue, most preferably to a lysine amino acid residue.

1. Transformer Site-Directed™ Mutagenesis

Preparation of non-endogenous human GPCRs may be accomplished on human GPCRs using Transformer Site-Directed™ Mutagenesis Kit (Clontech) according to the manufacturer instructions. Two mutagenesis primers are utilized, most preferably a lysine mutagenesis oligonucleotide that creates the lysine mutation, and a selection marker oligonucleotide. For convenience, the codon mutation to be incorporated into the human GPCR is also noted, in standard form (Table D):

TABLE D

Receptor Identifier	Codon Mutation
hRUP8	V274K
hRUP9	T249K
hRUP10	R232K
hRUP11	M294K
hRUP12	F220K
hRUP16	A238K

hRUP17	Y215K
hRUP18	L294K
hRUP19	T219K
hRUP20	K248A K248H K248R
hRUP21	R240K
hRUP22	Y222K
hRUP24	A245K
hRUP25	I230K
hRUP26	V285K
hRUP27	T248K

2. QuikChange™ Site-Directed™ Mutagenesis

Preparation of non-endogenous human GPCRs can also be accomplished by using QuikChange™ Site-Directed™ Mutagenesis Kit (Stratagene, according to manufacturer's instructions). Endogenous GPCR is preferably used as a template and two mutagenesis primers utilized, as well as, most preferably, a lysine mutagenesis oligonucleotide and a selection marker oligonucleotide (included in kit). For convenience, the codon mutation incorporated into the novel human GPCR and the respective oligonucleotides are noted, in standard form (Table E):

TABLE E

Receptor Identifier	Codon Mutation	5'-3' orientation (sense), (SEQ.ID.NO.) mutation underlined	5'-3' orientation (antisense) (SEQ.ID.NO.)	Cycle Conditions Min ('), Sec (") Cycles 2-4 repeated 16 times
hRUP13	A268K	GGGGAGGGAAAGCAA <u>AGGTGGTCCTCCTGG</u> (81)	CCAGGAGAACCACCT <u>TTGCTTTCCTCCCC</u> (82)	98° for 2' 98° for 30" 56°C for 30" 72° for 11' 40" 72° for 5'
hRUP14	L246K	CAGGAAGGCAAAGAC CACCATCATCATC (85)	GATGATGATGGTGGT <u>CTTGCCTTCCTG</u> (86)	98° for 2' 98° for 30" 55°C for 30" 72° for 11' 40" 72° for 5'

hRUP15	A398K	CCAGTGCAAAGCTAAG AAAGTGATCTTC (89)	GAAGATCACITTTCTTA GCTTTGCACTGG (90)	98° for 2' 98° for 30" 55°C for 30" 72° for 11' 40" 72° for 5'
hRUP23	W275K	GCCGCCACCGCGCCAA GAGGAAGATTGGC (93)	GCCAATCTTCCTCTTG GCGCGGTGGCGGC (94)	98° for 2' 98° for 30" 56°C for 30" 72° for 11' 40" 72° for 5'

The non-endogenous human GPCRs were then sequenced and the derived and verified nucleic acid and amino acid sequences are listed in the accompanying "Sequence Listing" appendix to this patent document, as summarized in Table F below:

TABLE F

Non Endogenous Human GPCR	Nucleic Acid Sequence Listing	Amino Acid Sequence Listing
hRUP13	SEQ.ID.NO.:83	SEQ.ID.NO.:84
hRUP14	SEQ.ID.NO.:87	SEQ.ID.NO.:88
hRUP15	SEQ.ID.NO.:91	SEQ.ID.NO.:92
hRUP23	SEQ.ID.NO.:95	SEQ.ID.NO.:96

Example 3

RECEPTOR EXPRESSION

Although a variety of cells are available to the art for the expression of proteins, it is most preferred that mammalian cells be utilized. The primary reason for this is predicated upon practicalities, *i.e.*, utilization of, *e.g.*, yeast cells for the expression of a GPCR, while possible, introduces into the protocol a non-mammalian cell which may not (indeed, in the case of yeast, does not) include the receptor-coupling, genetic-mechanism and secretory pathways that have evolved for mammalian systems – thus, results obtained in non-mammalian cells, while of

potential use, are not as preferred as that obtained from mammalian cells. Of the mammalian cells, COS-7, 293 and 293T cells are particularly preferred, although the specific mammalian cell utilized can be predicated upon the particular needs of the artisan.

5 **a. Transient Transfection**

On day one, 6×10^6 10 cm dish of 293 cells well were plated out. On day two, two reaction tubes were prepared (the proportions to follow for each tube are per plate): tube A was prepared by mixing $4\mu\text{g}$ DNA (*e.g.*, pCMV vector; pCMV vector with receptor cDNA, etc.) in 0.5 ml serum free DMEM (Gibco BRL); tube B was prepared by
10 mixing $24\mu\text{l}$ lipofectamine (Gibco BRL) in 0.5ml serum free DMEM. Tubes A and B were admixed by inversions (several times), followed by incubation at room temperature for 30-45min. The admixture is referred to as the "transfection mixture". Plated 293 cells were washed with 1XPBS, followed by addition of 5 ml serum free DMEM. 1 ml of the transfection mixture were added to the cells, followed by incubation for 4hrs at
15 $37^\circ\text{C}/5\% \text{CO}_2$. The transfection mixture was removed by aspiration, followed by the addition of 10ml of DMEM/10% Fetal Bovine Serum. Cells were incubated at $37^\circ\text{C}/5\% \text{CO}_2$. After 48hr incubation, cells were harvested and utilized for analysis.

b. Stable Cell Lines: Gs Fusion Protein

Approximately 12×10^6 293 cells are plated on a 15cm tissue culture plate.
20 Grown in DME High Glucose Medium containing ten percent fetal bovine serum and one percent sodium pyruvate, L-glutamine, and anti-biotics. Twenty-four hours following plating of 293 cells to ~80% confluency, the cells are transfected using $12\mu\text{g}$ of DNA. The $12\mu\text{g}$ of DNA is combined with $60\mu\text{l}$ of lipofectamine and 2mL of DME High Glucose Medium without serum. The medium is aspirated from the plates and the
25 cells are washed once with medium without serum. The DNA, lipofectamine, and

medium mixture is added to the plate along with 10mL of medium without serum. Following incubation at 37 degrees Celsius for four to five hours, the medium is aspirated and 25ml of medium containing serum is added. Twenty-four hours following transfection, the medium is aspirated again, and fresh medium with serum is added.

5 Forty-eight hours following transfection, the medium is aspirated and medium with serum is added containing geneticin (G418 drug) at a final concentration of 500µg/mL. The transfected cells now undergo selection for positively transfected cells containing the G418 resistant gene. The medium is replaced every four to five days as selection occurs. During selection, cells are grown to create stable pools, or split for stable clonal

10 selection.

Example 4

ASSAYS FOR DETERMINATION OF CONSTITUTIVE ACTIVITY OF NON-ENDOGENOUS GPCRS

15 A variety of approaches are available for assessment of constitutive activity of the non-endogenous human GPCRs. The following are illustrative; those of ordinary skill in the art are credited with the ability to determine those techniques that are preferentially beneficial for the needs of the artisan.

1. Membrane Binding Assays: [³⁵S]GTPγS Assay

20 When a G protein-coupled receptor is in its active state, either as a result of ligand binding or constitutive activation, the receptor couples to a G protein and stimulates the release of GDP and subsequent binding of GTP to the G protein. The alpha subunit of the G protein-receptor complex acts as a GTPase and slowly hydrolyzes

25 the GTP to GDP, at which point the receptor normally is deactivated. Constitutively activated receptors continue to exchange GDP for GTP. The non-hydrolyzable GTP analog, [³⁵S]GTPγS, can be utilized to demonstrate enhanced binding of [³⁵S]GTPγS to membranes expressing constitutively activated receptors. The advantage of using

[³⁵S]GTPγS binding to measure constitutive activation is that: (a) it is generically applicable to all G protein-coupled receptors; (b) it is proximal at the membrane surface making it less likely to pick-up molecules which affect the intracellular cascade.

The assay utilizes the ability of G protein coupled receptors to stimulate
5 [³⁵S]GTPγS binding to membranes expressing the relevant receptors. The assay can, therefore, be used in the direct identification method to screen candidate compounds to known, orphan and constitutively activated G protein-coupled receptors. The assay is generic and has application to drug discovery at all G protein-coupled receptors.

The [³⁵S]GTPγS assay was incubated in 20 mM HEPES and between 1 and
10 about 20mM MgCl₂ (this amount can be adjusted for optimization of results, although 20mM is preferred) pH 7.4, binding buffer with between about 0.3 and about 1.2 nM [³⁵S]GTPγS (this amount can be adjusted for optimization of results, although 1.2 is preferred) and 12.5 to 75 μg membrane protein (*e.g.*, 293 cells expressing the Gs Fusion Protein; this amount can be adjusted for optimization) and 10 μM GDP (this amount can
15 be changed for optimization) for 1 hour. Wheatgerm agglutinin beads (25 μl; Amersham) were then added and the mixture incubated for another 30 minutes at room temperature. The tubes were then centrifuged at 1500 x g for 5 minutes at room temperature and then counted in a scintillation counter.

2. Adenylyl Cyclase

20 A Flash Plate™ Adenylyl Cyclase kit (New England Nuclear; Cat. No. SMP004A) designed for cell-based assays can be modified for use with crude plasma membranes. The Flash Plate wells can contain a scintillant coating which also contains a specific antibody recognizing cAMP. The cAMP generated in the wells can be quantitated by a direct competition for binding of radioactive cAMP tracer to the cAMP

antibody. The following serves as a brief protocol for the measurement of changes in cAMP levels in whole cells that express the receptors.

Transfected cells were harvested approximately twenty four hours after transient transfection. Media is carefully aspirated off and discarded. 10ml of PBS is gently
5 added to each dish of cells followed by careful aspiration. 1ml of Sigma cell dissociation buffer and 3ml of PBS are added to each plate. Cells were pipeted off the plate and the cell suspension was collected into a 50ml conical centrifuge tube. Cells were then centrifuged at room temperature at 1,100 rpm for 5 min. The cell pellet was carefully re-suspended into an appropriate volume of PBS (about 3ml/plate). The cells
10 were then counted using a hemocytometer and additional PBS was added to give the appropriate number of cells (with a final volume of about 50 μ l/well).

cAMP standards and Detection Buffer (comprising 1 μ Ci of tracer [125 I] cAMP (50 μ l] to 11 ml Detection Buffer) was prepared and maintained in accordance with the manufacturer's instructions. Assay Buffer was prepared fresh for screening and
15 contained 50 μ l of Stimulation Buffer, 3 μ l of test compound (12 μ M final assay concentration) and 50 μ l cells, Assay Buffer was stored on ice until utilized. The assay was initiated by addition of 50 μ l of cAMP standards to appropriate wells followed by addition of 50 μ l of PBSA to wells H-11 and H12. 50 μ l of Stimulation Buffer was added to all wells. DMSO (or selected candidate compounds) was added to appropriate wells
20 using a pin tool capable of dispensing 3 μ l of compound solution, with a final assay concentration of 12 μ M test compound and 100 μ l total assay volume. The cells were then added to the wells and incubated for 60 min at room temperature. 100 μ l of Detection Mix containing tracer cAMP was then added to the wells. Plates were then incubated additional 2 hours followed by counting in a Wallac MicroBeta scintillation

counter. Values of cAMP/well were then extrapolated from a standard cAMP curve which was contained within each assay plate.

3. Cell-Based cAMP for Gi Coupled Target GPCRs

5 TSHR is a Gs coupled GPCR that causes the accumulation of cAMP upon activation. TSHR will be constitutively activated by mutating amino acid residue 623 (*i.e.*, changing an alanine residue to an isoleucine residue). A Gi coupled receptor is expected to inhibit adenylyl cyclase, and, therefore, decrease the level of cAMP production, which can make assessment of cAMP levels challenging. An effective
10 technique for measuring the decrease in production of cAMP as an indication of constitutive activation of a Gi coupled receptor can be accomplished by co-transfecting, most preferably, non-endogenous, constitutively activated TSHR (TSHR-A623I) (or an endogenous, constitutively active Gs coupled receptor) as a “signal enhancer” with a Gi linked target GPCR to establish a baseline level of cAMP. Upon creating a non-
15 endogenous version of the Gi coupled receptor, this non-endogenous version of the target GPCR is then co-transfected with the signal enhancer, and it is this material that can be used for screening. We will utilize such approach to effectively generate a signal when a cAMP assay is used; this approach is preferably used in the direct identification of candidate compounds against Gi coupled receptors. It is noted that for a Gi coupled
20 GPCR, when this approach is used, an inverse agonist of the target GPCR will increase the cAMP signal and an agonist will decrease the cAMP signal.

On day one, 2×10^4 293 and 293 cells/well will be plated out. On day two, two reaction tubes will be prepared (the proportions to follow for each tube are per plate): tube A will be prepared by mixing 2 μ g DNA of each receptor transfected into the
25 mammalian cells, for a total of 4 μ g DNA (*e.g.*, pCMV vector; pCMV vector with mutated TSHR (TSHR-A623I); TSHR-A623I and GPCR, etc.) in 1.2ml serum free

DMEM (Irvine Scientific, Irvine, CA); tube B will be prepared by mixing 120µl lipofectamine (Gibco BRL) in 1.2ml serum free DMEM. Tubes A and B will then be admixed by inversions (several times), followed by incubation at room temperature for 30-45min. The admixture is referred to as the "transfection mixture". Plated 293 cells
5 will be washed with 1XPBS, followed by addition of 10ml serum free DMEM. 2.4ml of the transfection mixture will then be added to the cells, followed by incubation for 4hrs at 37°C/5% CO₂. The transfection mixture will then be removed by aspiration, followed by the addition of 25ml of DMEM/10% Fetal Bovine Serum. Cells will then be incubated at 37°C/5% CO₂. After 24hr incubation, cells will then be harvested and
10 utilized for analysis.

A Flash Plate™ Adenylyl Cyclase kit (New England Nuclear; Cat. No. SMP004A) is designed for cell-based assays, however, can be modified for use with crude plasma membranes depending on the need of the skilled artisan. The Flash Plate wells will contain a scintillant coating which also contains a specific antibody
15 recognizing cAMP. The cAMP generated in the wells can be quantitated by a direct competition for binding of radioactive cAMP tracer to the cAMP antibody. The following serves as a brief protocol for the measurement of changes in cAMP levels in whole cells that express the receptors.

Transfected cells will be harvested approximately twenty four hours after
20 transient transfection. Media will be carefully aspirated off and discarded. 10ml of PBS will be gently added to each dish of cells followed by careful aspiration. 1ml of Sigma cell dissociation buffer and 3ml of PBS will be added to each plate. Cells will be pipeted off the plate and the cell suspension will be collected into a 50ml conical centrifuge tube. Cells will then be centrifuged at room temperature at 1,100 rpm for 5 min. The cell
25 pellet will be carefully re-suspended into an appropriate volume of PBS (about

3ml/plate). The cells will then be counted using a hemocytometer and additional PBS is added to give the appropriate number of cells (with a final volume of about 50µl/well).

cAMP standards and Detection Buffer (comprising 1 µCi of tracer [¹²⁵I] cAMP (50 µl] to 11 ml Detection Buffer) will be prepared and maintained in accordance with the manufacturer's instructions. Assay Buffer should be prepared fresh for screening and contained 50µl of Stimulation Buffer, 3ul of test compound (12uM final assay concentration) and 50µl cells, Assay Buffer can be stored on ice until utilized. The assay can be initiated by addition of 50µl of cAMP standards to appropriate wells followed by addition of 50µl of PBSA to wells H-11 and H12. 50ul of Stimulation Buffer will be added to all wells. Selected compounds (*e.g.*, TSH) will be added to appropriate wells using a pin tool capable of dispensing 3µl of compound solution, with a final assay concentration of 12µM test compound and 100µl total assay volume. The cells will then be added to the wells and incubated for 60 min at room temperature. 100µl of Detection Mix containing tracer cAMP will then be added to the wells. Plates were then incubated additional 2 hours followed by counting in a Wallac MicroBeta scintillation counter. Values of cAMP/well will then be extrapolated from a standard cAMP curve which is contained within each assay plate.

4. Reporter-Based Assays

a. CRE-LUC Reporter Assay (Gs-associated receptors)

293 and 293T cells are plated-out on 96 well plates at a density of 2×10^4 cells per well and were transfected using Lipofectamine Reagent (BRL) the following day according to manufacturer instructions. A DNA/lipid mixture is prepared for each 6-well transfection as follows: 260ng of plasmid DNA in 100µl of DMEM were gently mixed with 2µl of lipid in 100µl of DMEM (the 260ng of plasmid DNA consisted of

200ng of a 8xCRE-Luc reporter plasmid, 50ng of pCMV comprising endogenous receptor or non-endogenous receptor or pCMV alone, and 10ng of a GPRS expression plasmid (GPRS in pcDNA3 (Invitrogen)). The 8XCRE-Luc reporter plasmid was prepared as follows: vector SRIF- β -gal was obtained by cloning the rat somatostatin promoter (-71/+51) at BglV-HindIII site in the p β gal-Basic Vector (Clontech). Eight (8) copies of cAMP response element were obtained by PCR from an adenovirus template AdpCF126CCRE8 (*see, 7 Human Gene Therapy* 1883 (1996)) and cloned into the SRIF- β -gal vector at the Kpn-BglV site, resulting in the 8xCRE- β -gal reporter vector. The 8xCRE-Luc reporter plasmid was generated by replacing the beta-galactosidase gene in the 8xCRE- β -gal reporter vector with the luciferase gene obtained from the pGL3-basic vector (Promega) at the HindIII-BamHI site. Following 30 min. incubation at room temperature, the DNA/lipid mixture was diluted with 400 μ l of DMEM and 100 μ l of the diluted mixture was added to each well. 100 μ l of DMEM with 10% FCS were added to each well after a 4hr incubation in a cell culture incubator. The following day the transfected cells were changed with 200 μ l/well of DMEM with 10% FCS. Eight (8) hours later, the wells were changed to 100 μ l /well of DMEM without phenol red, after one wash with PBS. Luciferase activity were measured the next day using the LucLite™ reporter gene assay kit (Packard) following manufacturer instructions and read on a 1450 MicroBeta™ scintillation and luminescence counter (Wallac).

b. AP1 reporter assay (Gq-associated receptors)

A method to detect Gq stimulation depends on the known property of Gq-dependent phospholipase C to cause the activation of genes containing AP1 elements in their promoter. A Pathdetect™ AP-1 cis-Reporting System (Stratagene, Catalogue # 219073) can be utilized following the protocol set forth above with respect to the

CREB reporter assay, except that the components of the calcium phosphate precipitate were 410 ng pAP1-Luc, 80 ng pCMV-receptor expression plasmid, and 20 ng CMV-SEAP.

c. SRF-LUC Reporter Assay (Gq- associated receptors)

One method to detect Gq stimulation depends on the known property of Gq-dependent phospholipase C to cause the activation of genes containing serum response factors in their promoter. A Pathdetect™ SRF-Luc-Reporting System (Stratagene) can be utilized to assay for Gq coupled activity in, *e.g.*, COS7 cells. Cells are transfected with the plasmid components of the system and the indicated expression plasmid encoding endogenous or non-endogenous GPCR using a Mammalian Transfection™ Kit (Stratagene, Catalogue #200285) according to the manufacturer's instructions. Briefly, 410 ng SRF-Luc, 80 ng pCMV-receptor expression plasmid and 20 ng CMV-SEAP (secreted alkaline phosphatase expression plasmid; alkaline phosphatase activity is measured in the media of transfected cells to control for variations in transfection efficiency between samples) are combined in a calcium phosphate precipitate as per the manufacturer's instructions. Half of the precipitate is equally distributed over 3 wells in a 96-well plate, kept on the cells in a serum free media for 24 hours. The last 5 hours the cells are incubated with 1 μM Angiotensin, where indicated. Cells are then lysed and assayed for luciferase activity using a Lucite™ Kit (Packard, Cat. # 6016911) and "Trilux 1450 Microbeta" liquid scintillation and luminescence counter (Wallac) as per the manufacturer's instructions. The data can be analyzed using GraphPad Prism™ 2.0a (GraphPad Software Inc.).

d. Intracellular IP₃ Accumulation Assay (Gq-associated receptors)

On day 1, cells comprising the receptors (endogenous and/or non-endogenous)
5 can be plated onto 24 well plates, usually 1×10^5 cells/well (although this number can be optimized. On day 2 cells can be transfected by firstly mixing 0.25 μ g DNA in 50 μ l serum free DMEM/well and 2 μ l lipofectamine in 50 μ l serumfree DMEM/well. The solutions are gently mixed and incubated for 15-30 min at room temperature. Cells are washed with 0.5 ml PBS and 400 μ l of serum free media is mixed with the transfection
10 media and added to the cells. The cells are then incubated for 3-4 hrs at 37°C/5%CO₂ and then the transfection media is removed and replaced with 1ml/well of regular growth media. On day 3 the cells are labeled with ³H-myo-inositol. Briefly, the media is removed and the cells are washed with 0.5 ml PBS. Then 0.5 ml inositol-free/serum free media (GIBCO BRL) is added/well with 0.25 μ Ci of ³H-myo-inositol/ well and the cells
15 are incubated for 16-18 hrs o/n at 37°C/5%CO₂. On Day 4 the cells are washed with 0.5 ml PBS and 0.45 ml of assay medium is added containing inositol-free/serum free media 10 μ M pargyline 10 mM lithium chloride or 0.4 ml of assay medium and 50 μ l of 10x ketanserin (ket) to final concentration of 10 μ M. The cells are then incubated for 30 min at 37°C. The cells are then washed with 0.5 ml PBS and 200 μ l of fresh/ice cold stop
20 solution (1M KOH; 18 mM Na-borate; 3.8 mM EDTA) is added/well. The solution is kept on ice for 5-10 min or until cells were lysed and then neutralized by 200 μ l of fresh/ice cold neutralization sol. (7.5 % HCL). The lysate is then transferred into 1.5 ml eppendorf tubes and 1 ml of chloroform/methanol (1:2) is added/tube. The solution is vortexed for 15 sec and the upper phase is applied to a Biorad AG1-X8™ anion
25 exchange resin (100-200 mesh). Firstly, the resin is washed with water at 1:1.25 W/V and 0.9 ml of upper phase is loaded onto the column. The column is washed with 10 mls of 5 mM myo-inositol and 10 ml of 5 mM Na-borate/60mM Na-formate. The inositol

tris phosphates are eluted into scintillation vials containing 10 ml of scintillation cocktail with 2 ml of 0.1 M formic acid/ 1 M ammonium formate. The columns are regenerated by washing with 10 ml of 0.1 M formic acid/3M ammonium formate and rinsed twice with dd H₂O and stored at 4°C in water.

5 Exemplary results are presented below in Table G:

TABLE G

Receptor	Mutation	Assay Utilized (Figure No.)	Signal Generated: CMV	Signal Generated: Endogenous Version (Relative Light Units)	Signal Generated: Non-Endogenous Version (Relative Light Units)	Difference (⇐) Between CMV v. Wild-type v. Mutant
hRUP12	N/A	IP ₃ (Figure 1)	317.03 cpm/mg protein	3463.29 cpm/mg protein	--	1. 11 Fold ⇐
hRUP13	N/A	cAMP (Figure 2)	8.06 pmol/cAMP/mg protein	19.10 pmol/cAMP/mg protein	--	1. 2.4 Fold ⇐
	A268K	8XCRE-LUC (Figure 3)	3665.43 LCPS	83280.17 LCPS	61713.6 LCPS	1. 23 Fold ⇐ 2. 26 % <
hRUP14	L246K	8XCRE-LUC (Figure 5)	86.07 LCPS	1962.87 LCPS	789.73 LCPS	1. 23 Fold ⇐ 2. 60% <
hRUP15	A398K	8XCRE-LUC (Figure 6)	86.07 LCPS	18286.77 LCPS	17034.83 LCPS	1. 212 Fold ⇐ 2. 1% <
	A398K	cAMP (Figure 7)	15.00 pmol/cAMP/mg protein	164.4 pmol/cAMP/mg protein	117.5 pmol/cAMP/mg protein	1. 11 Fold ⇐ 2. 29% <
hRUP17	N/A	IP ₃ (Figure 9)	317.03 cpm/mg protein	741.07 cpm/mg protein	--	1. 2.3 Fold ⇐
hRUP21	N/A	IP ₃ (Figure 10)	730.5 cpm/mg protein	1421.9 cpm/mg protein	--	1. 2 Fold ⇐
hRUP23	W275K	8XCRE-LUC (Figure 11)	311.73 pmol/cAMP/mg protein	13756.00 pmol/cAMP/mg protein	9756.87 pmol/cAMP/mg protein	1. 44 Fold ⇐ 2. 30% <

N/A = not applied

Exemplary results of GTP γ S assay for detecting constitutive activation, as disclosed in Example 4(1) above, was accomplished utilizing Gs:Fusion Protein Constructs on human RUP13 and RUP15. Table H below lists the signals generated from this assay and the difference in signals as indicated:

5

TABLE H

Receptor: Gs Fusion Protein	Assay Utilized	Signal Generated: CMV (cpm bound GTP)	Signal Generated: Fusion Protein (cpm bound GTP)	Signal Generated: CMV+ 10 μ M GDP (cpm bound GTP)	Signal Generated: Fusion Protein + 10 μ M GDP (cpm bound GTP)	Difference Between: 1. CMV v. Fusion Protein 2. CMV+GDP vs. Fusion+GDP 3. Fusion vs. Fusion+GDP (cpm bound GTP)
hRUP13-Gs	GTP γ S (Figure 4)	32494.0	49351.30	11148.30	28834.67	1. 1.5 Fold \Leftarrow 2. 2.6 Fold \Leftarrow 3. 42% \langle
hRUP15-Gs	GTP γ S (Figure 8)	30131.67	32493.67	7697.00	14157.33	1. 1.1 Fold \Leftarrow 2. 1.8 Fold \Leftarrow 3. 56% \langle

Example 5

FUSION PROTEIN PREPARATION

10 a. GPCR:Gs Fusion Construct

The design of the constitutively activated GPCR-G protein fusion construct was accomplished as follows: both the 5' and 3' ends of the rat G protein Gs α (long form; Itoh, H. et al., 83 *PNAS* 3776 (1986)) were engineered to include a HindIII (5'-
15 AAGCTT-3') sequence thereon. Following confirmation of the correct sequence (including the flanking HindIII sequences), the entire sequence was shuttled into pcDNA3.1(-) (Invitrogen, cat. no. V795-20) by subcloning using the HindIII restriction site of that vector. The correct orientation for the Gs α sequence was determined after

subcloning into pcDNA3.1(-). The modified pcDNA3.1(-) containing the rat Gs α gene at HindIII sequence was then verified; this vector was now available as a “universal” Gs α protein vector. The pcDNA3.1(-) vector contains a variety of well-known restriction sites upstream of the HindIII site, thus beneficially providing the ability to insert, upstream of the Gs protein, the coding sequence of an endogenous, constitutively active GPCR. This same approach can be utilized to create other “universal” G protein vectors, and, of course, other commercially available or proprietary vectors known to the artisan can be utilized – the important criteria is that the sequence for the GPCR be upstream and in-frame with that of the G protein.

RUP13 couples via Gs. For the following exemplary GPCR Fusion Proteins, fusion to Gs α was accomplished.

A RUP13-Gs α Fusion Protein construct was made as follows: primers were designed as follows:

5'-gatc[TCTAGAAT]GGAGTCCTCACCCATCCCCCAG-3' (SEQ.ID.NO.:97; sense)

5'-gatc[GATATC]CGTGACTCCAGCCGGGGTGAGGCGGC-3' (SEQ.ID.NO.:98; antisense).

Nucleotides in lower caps are included as spacers in the restriction sites (designated in brackets) between the G protein and RUP13. The sense and anti-sense primers included the restriction sites for XbaI and EcoRV, respectively, such that spacers (attributed to the restriction sites) exists between the G protein and RUP15.

PCR was then utilized to secure the respective receptor sequences for fusion within the Gs α universal vector disclosed above, using the following protocol for each: 100ng cDNA for RUP15 was added to separate tubes containing 2 μ l of each primer (sense and anti-sense), 3 μ L of 10mM dNTPs, 10 μ L of 10XTaqPlus™ Precision buffer, 1 μ L of TaqPlus™ Precision polymerase (Stratagene: #600211), and 80 μ L of water.

Reaction temperatures and cycle times for RUP15 were as follows with cycle steps 2

through 4 were repeated 35 times: 94°C for 1 min; 94°C for 30 seconds; 62°C for 20 sec; 72°C 1 min 40sec; and 72° C 5 min . PCR product for was run on a 1% agarose gel and then purified (data not shown). The purified product was digested with XbaI and EcoRV and the desired inserts purified and ligated into the Gs universal vector at the
5 respective restriction site. The positive clones was isolated following transformation and determined by restriction enzyme digest; expression using 293 cells was accomplished following the protocol set forth *infra*. Each positive clone for RUP15-Gs Fusion Protein was sequenced to verify correctness. (See, SEQ.ID.NO.:99 for nucleic acid sequence and SEQ.ID.NO.:100 for amino acid sequence).

10 RUP15 couples via Gs. For the following exemplary GPCR Fusion Proteins, fusion to Gs α was accomplished.

A RUP15-Gs α Fusion Protein construct was made as follows: primers were designed as follows:

5'-TCTAGAATGACGTCCACCTGCACCAACAGC-3' (SEQ.ID.NO.:101; sense)

15 5'-gatatcGCAGGAAAAGTAGCAGAATCGTAGGAAG-3' (SEQ.ID.NO.:102; antisense).

Nucleotides in lower caps are included as spacers in the restriction sites between the G protein and RUP15. The sense and anti-sense primers included the restriction sites for EcoRV and XbaI, respectively, such that spacers (attributed to the restriction sites) exists between the G protein and RUP15.

20 PCR was then utilized to secure the respective receptor sequences for fusion within the Gs α universal vector disclosed above, using the following protocol for each: 100ng cDNA for RUP15 was added to separate tubes containing 2 μ l of each primer (sense and anti-sense), 3 μ L of 10mM dNTPs, 10 μ L of 10XTaqPlus™ Precision buffer, 1 μ L of TaqPlus™ Precision polymerase (Stratagene: #600211), and 80 μ L of water.
25 Reaction temperatures and cycle times for RUP15 were as follows with cycle steps 2

through 4 were repeated 35 times: 94°C for 1 min; 94°C for 30 seconds; 62°C for 20 sec; 72°C 1 min 40sec; and 72° C 5 min . PCR product for was run on a 1% agarose gel and then purified (data not shown). The purified product was digested). The purified product was digested with EcoRV and XbaI and the desired inserts purified and
5 ligated into the Gs universal vector at the respective restriction site. The positive clones was isolated following transformation and determined by restriction enzyme digest; expression using 293 cells was accomplished following the protocol set forth *infra*. Each positive clone for RUP15-Gs Fusion Protein was sequenced to verify correctness. (See, SEQ.ID.NO.:103 for nucleic acid sequence and SEQ.ID.NO.:104 for amino acid
10 sequence).

b. Gq(6 amino acid deletion)/Gi Fusion Construct

The design of a Gq (del)/Gi fusion construct can be accomplished as follows:
the N-terminal six (6) amino acids (amino acids 2 through 7, having the sequence of TLESIM (SEQ.ID.NO.: 129) Gαq-subunit will be deleted and the C-terminal five (5)
15 amino acids, having the sequence EYNLV (SEQ.ID.NO.:130) will be replace with the corresponding amino acids of the Gαi Protein, having the sequence DCGLF (SEQ.ID.NO.:131). This fusion construct will be obtained by PCR using the following primers:

20 5'-gatcaagcttcCATGGCGTGCTGCCTGAGCGAGGAG-3' (SEQ.ID.NO.:132) and

5'-gatcggatccTTAGAACAGGCCGAGTCCTTCAGGTTTCAGCTGCAGGATGGTG-3' (SEQ.ID.NO.:133)

and Plasmid 63313 which contains the mouse Gαq-wild type version with a
25 hemagglutinin tag as template. Nucleotides in lower caps are included as spacers.

TaqPlus Precision DNA polymerase (Stratagene) will be utilized for the amplification by the following cycles, with steps 2 through 4 repeated 35 times: 95°C

for 2 min; 95°C for 20 sec; 56°C for 20 sec; 72°C for 2 min; and 72°C for 7 min. The PCR product will be cloned into a pCRII-TOPO vector (Invitrogen) and sequenced using the ABI Big Dye Terminator kit (P.E. Biosystem). Inserts from a TOPO clone containing the sequence of the fusion construct will be shuttled into the expression
5 vector pcDNA3.1(+) at the HindIII/BamHI site by a 2 step cloning process.

Example 6

TISSUE DISTRIBUTION OF THE DISCLOSED HUMAN GPCRs: RT-PCR

RT-PCR was applied to confirm the expression and to determine the tissue
10 distribution of several novel human GPCRs. Oligonucleotides utilized were GPCR-specific and the human multiple tissue cDNA panels (MTC, Clontech) as templates. Taq DNA polymerase (Stratagene) were utilized for the amplification in a 40µl reaction according to the manufacturer's instructions. 20µl of the reaction will be loaded on a 1.5% agarose gel to analyze the RT-PCR products. Table J below lists the
15 receptors, the cycle conditions and the primers utilized.

TABLE J

Receptor Identifier	Cycle Conditions Min ('), Sec (") Cycles 2-4 repeated 30 times	5' Primer (SEQ.ID.NO.)	3' Primer (SEQ.ID.NO.)	DNA Fragment	Tissue Expression
hRUP10	94° for 30" 94° for 10" 62°C for 20" 72° for 1' 72° for 7' *cycles 2-4 repeated 35 times	CATGTATGC CAGCGTCCT GCTCC (105)	GCTATGCCTG AAGCCAGTC TTGTG (106)	730bp	Kidney, leukocyte, liver, placenta and spleen
hRUP11	94° for 2' 94° for 15" 67°C for 15" 72° for 45" 72° for 5'	GCACCTGCT CCTGAGCAC CTTCTCC (107)	CACAGCGCT GCAGCCCTG CAGCTGGC (108)	630bp	Liver, kidney, pancreas, colon, small intestinal, spleen and prostate

hRUP12	94° for 2' 94° for 15" 66°C for 15" 72° for 45" 72° for 5'	CCAGTGATG ACTCTGTCC AGCCTG (109)	CAGACACTT GGCAGGGAC GAGGTG (110)	490bp	Brain, colon, heart, kidney, leukocyte, pancreas, prostate, small intestinal, spleen, testis, and thymus
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hRUP13	94° for 1' 94° for 15" 68°C for 20" 72° for 1' 45" 72° for 5'	CTTGTGGTCT ACTGCAGCA TGTTCCG (111)	CATATCCCTC CGAGTGTC AGCGGC (112)	700bp	Placenta and lung
hRUP14	94° for 1' 94° for 15" 68°C for 20" 72° for 1' 45" 72° for 5'	ATGGATCCT TATCATGGC TTCCTC (113)	CAAGAACAG GTCTCATCTA AGAGCTCC (114)	700bp	Not yet determined
hRUP16	94° for 30" 94° for 5" 69°C for 15" 72° for 30" 72° for 5'	CTCTGATGC CATCTGCTG GATTCTG (115)	GTAGTCCACT GAAAGTCCA GTGATCC (116)	370bp	Fetal brain, fetal kidney and fetal skeletal muscle
hRUP18	94° for 2' 94° for 15" 60°C for 20" 72° for 1' 72° for 5'	TGGTGGCGA TGGCCAACA GCGCTC (117)	GTTGCGCCTT AGCGACAGA TGACC (118)	330bp	Pancreas
hRUP21	94° for 1' 94° for 15" 56°C for 20" 72° for 40" *cycles 2-3 repeated 30 times	TCAACCTGT ATAGCAGCA TCCTC (119)	AAGGAGTAG CAGAATGGT TAGCC (120)		Kidney, lung and testis
hRUP22	94° for 30" 94° for 15" 69°C for 20" 72° for 40" *cycles 2-3 repeated 30 times	GACACCTGT CAGCGGTCG TGTGTG (121)	CTGATGGAA GTAGAGGCT GTCCATCTC (122)		Testis, thymus and spleen
hRUP23	94° for 2' 94° for 15" 60°C for 20" 72° for 1' 72° for 5'	GCGCTGAGC GCAGACCAG TGGCTG (123)	CACGGTGAC GAAGGGCAC GAGCTC (124)	520bp	Placenta
hRUP26	94° for 2' 94° for 15" 65°C for 20" 72° for 1' 72° for 5'	AGCCATCCC TGCCAGGAA GCATGG (125)	CCAGGTAGG TGTGCAGCA CAATGGC (126)	470bp	Pancreas
hRUP27	94° for 30" 94° for 10" 55°C for 20" 72° for 1' 72° for 3' *cycles 2-4 repeated 35 times	CTGTTCAAC AGGGCTGGT TGGCAAC (127)	ATCATGTCTA GACTCATGGT GATCC (128)	890bp	Brain

Example 7

Protocol: Direct Identification of Inverse Agonists and Agonists

A. [³⁵S]GTPγS Assay

5 Although we have utilized endogenous, constitutively active GPCRs for the direct identification of candidate compounds as, *e.g.*, inverse agonists, for reasons that are not altogether understood, intra-assay variation can become exacerbated. Preferably, then, a GPCR Fusion Protein, as disclosed above, is also utilized with a non-endogenous, constitutively activated GPCR. We have determined that when such a
10 protein is used, intra-assay variation appears to be substantially stabilized, whereby an effective signal-to-noise ratio is obtained. This has the beneficial result of allowing for a more robust identification of candidate compounds. Thus, it is preferred that for direct identification, a GPCR Fusion Protein be used and that when utilized, the following assay protocols be utilized.

15 1. Membrane Preparation

Membranes comprising the constitutively active orphan GPCR Fusion Protein of interest and for use in the direct identification of candidate compounds as inverse agonists, agonists or partial agonists are preferably prepared as follows:

a. Materials

20 “Membrane Scrape Buffer” is comprised of 20mM HEPES and 10mM EDTA, pH 7.4; “Membrane Wash Buffer” is comprised of 20 mM HEPES and 0.1 mM EDTA, pH 7.4; “Binding Buffer” is comprised of 20mM HEPES, 100 mM NaCl, and 10 mM MgCl₂, pH 7.4

b. Procedure

25 All materials will be kept on ice throughout the procedure. Firstly, the media will be aspirated from a confluent monolayer of cells, followed by rinse with 10ml cold

PBS, followed by aspiration. Thereafter, 5ml of Membrane Scrape Buffer will be added to scrape cells; this will be followed by transfer of cellular extract into 50ml centrifuge tubes (centrifuged at 20,000 rpm for 17 minutes at 4°C). Thereafter, the supernatant will be aspirated and the pellet will be resuspended in 30ml Membrane Wash Buffer
5 followed by centrifuge at 20,000 rpm for 17 minutes at 4°C. The supernatant will then be aspirated and the pellet resuspended in Binding Buffer. This will then be homogenized using a Brinkman polytron™ homogenizer (15-20 second bursts until the all material is in suspension). This is referred to herein as “Membrane Protein”.

2. Bradford Protein Assay

10 Following the homogenization, protein concentration of the membranes will be determined using the Bradford Protein Assay (protein can be diluted to about 1.5mg/ml, aliquoted and frozen (-80°C) for later use; when frozen, protocol for use will be as follows: on the day of the assay, frozen Membrane Protein is thawed at room temperature, followed by vortex and then homogenized with a polytron at about
15 12 x 1,000 rpm for about 5-10 seconds; it was noted that for multiple preparations, the homogenizer should be thoroughly cleaned between homogenization of different preparations).

a. Materials

Binding Buffer (as per above); Bradford Dye Reagent; Bradford Protein
20 Standard will be utilized, following manufacturer instructions (Biorad, cat. no. 500-0006).

b. Procedure

Duplicate tubes will be prepared, one including the membrane, and one as a control “blank”. Each contained 800ul Binding Buffer. Thereafter, 10ul of Bradford
25 Protein Standard (1mg/ml) will be added to each tube, and 10ul of membrane Protein

will then be added to just one tube (not the blank). Thereafter, 200ul of Bradford Dye Reagent will be added to each tube, followed by vortex of each. After five (5) minutes, the tubes will be re-vortexed and the material therein will be transferred to cuvettes. The cuvettes will then be read using a CECIL 3041 spectrophotometer, at
5 wavelength 595.

3. Direct Identification Assay

a. Materials

GDP Buffer consisted of 37.5 ml Binding Buffer and 2mg GDP (Sigma, cat. no. G-7127), followed by a series of dilutions in Binding Buffer to obtain 0.2 μ M GDP
10 (final concentration of GDP in each well was 0.1 μ M GDP); each well comprising a candidate compound, has a final volume of 200ul-consisting of 100 μ l GDP Buffer (final concentration, 0.1 μ M GDP), 50ul Membrane Protein in Binding Buffer, and 50 μ l [³⁵S]GTP γ S (0.6 nM) in Binding Buffer (2.5 μ l [³⁵S]GTP γ S per 10ml Binding Buffer).

b. Procedure

15 Candidate compounds will be preferably screened using a 96-well plate format (these can be frozen at -80°C). Membrane Protein (or membranes with expression vector excluding the GPCR Fusion Protein, as control), will be homogenized briefly until in suspension. Protein concentration will then be determined using the Bradford Protein Assay set forth above. Membrane Protein (and control) will then be diluted to
20 0.25mg/ml in Binding Buffer (final assay concentration, 12.5 μ g/well). Thereafter, 100 μ l GDP Buffer was added to each well of a Wallac Scintistrip™ (Wallac). A 5ul pin-tool will then be used to transfer 5 μ l of a candidate compound into such well (*i.e.*, 5 μ l in total assay volume of 200 μ l is a 1:40 ratio such that the final screening concentration of the candidate compound is 10 μ M). Again, to avoid contamination, after each transfer
25 step the pin tool should be rinsed in three reservoirs comprising water (1X), ethanol (1X)

and water (2X) – excess liquid should be shaken from the tool after each rinse and dried with paper and kimwipes. Thereafter, 50 µl of Membrane Protein will be added to each well (a control well comprising membranes without the GPCR Fusion Protein was also utilized), and pre-incubated for 5-10 minutes at room temperature. Thereafter, 50µl of
5 [35S]GTPγS (0.6 nM) in Binding Buffer will be added to each well, followed by incubation on a shaker for 60 minutes at room temperature (again, in this example, plates were covered with foil). The assay will then be stopped by spinning of the plates at 4000 RPM for 15 minutes at 22°C. The plates will then be aspirated with an 8 channel manifold and sealed with plate covers. The plates will then be read on a Wallac 1450
10 using setting “Prot. #37” (as per manufacturer instructions).

B. Cyclic AMP Assay

Another assay approach to directly identified candidate compound was accomplished by utilizing a cyclase-based assay. In addition to direct identification, this
15 assay approach can be utilized as an independent approach to provide confirmation of the results from the [35S]GTPγS approach as set forth above.

A modified Flash Plate™ Adenylyl Cyclase kit (New England Nuclear; Cat. No. SMP004A) was preferably utilized for direct identification of candidate compounds as inverse agonists and agonists to constitutively activated orphan GPCRs in accordance
20 with the following protocol.

Transfected cells were harvested approximately three days after transfection. Membranes were prepared by homogenization of suspended cells in buffer containing 20mM HEPES, pH 7.4 and 10mM MgCl₂. Homogenization was performed on ice using a Brinkman Polytron™ for approximately 10 seconds. The resulting homogenate is
25 centrifuged at 49,000 X g for 15 minutes at 4°C. The resulting pellet was then resuspended in buffer containing 20mM HEPES, pH 7.4 and 0.1 mM EDTA,

homogenized for 10 seconds, followed by centrifugation at 49,000 X g for 15 minutes at 4°C. The resulting pellet was then stored at -80°C until utilized. On the day of direct identification screening, the membrane pellet was slowly thawed at room temperature, resuspended in buffer containing 20mM HEPES, pH 7.4 and 10mM MgCl₂, to yield a
5 final protein concentration of 0.60mg/ml (the resuspended membranes are placed on ice until use).

cAMP standards and Detection Buffer (comprising 2 µCi of tracer [¹²⁵I] cAMP (100 µl) to 11 ml Detection Buffer) were prepared and maintained in accordance with the manufacturer's instructions. Assay Buffer was prepared fresh for screening and
10 contained 20mM HEPES, pH 7.4, 10mM MgCl₂, 20mM phosphocreatine (Sigma), 0.1 units/ml creatine phosphokinase (Sigma), 50 µM GTP (Sigma), and 0.2 mM ATP (Sigma); Assay Buffer was then stored on ice until utilized.

Candidate compounds identified as per above (if frozen, thawed at room temperature) were added, preferably, to 96-well plate wells (3µl/well; 12µM final assay
15 concentration), together with 40 µl Membrane Protein (30µg/well) and 50µl of Assay Buffer. This admixture was then incubated for 30 minutes at room temperature, with gentle shaking.

Following the incubation, 100µl of Detection Buffer was added to each well, followed by incubation for 2-24 hours. Plates were then counted in a Wallac
20 MicroBeta™ plate reader using "Prot. #31" (as per manufacturer instructions).

A representative screening assay plate (96 well format) result is presented in Figure 12. Each bar represents the results for a different compound in each well, plus RUP13-Gsα Fusion Protein construct, as prepared in Example 5(a) above. The representative results presented in Figure 12 also provide standard deviations based upon
25 the mean results of each plate ("m") and the mean plus two arbitrary preference for

selection of inverse agonists as "leads" from the primary screen involves selection of candidate compounds that reduce the per cent response by at least the mean plate response, minus two standard deviations. Conversely, an arbitrary preference for selection of an agonists as "leads" from the primary screen involves selection of candidate compounds that increase the per cent response by at least the mean plate response, plus the two standard deviations. Based upon these selection processes, the candidate compounds in the following wells were directly identified as putative inverse agonist (Compound A) and agonist (Compound B) to RUP13 in wells A2 and G9, respectively. *See*, Figure 12. It is noted for clarity: these compounds have been directly identified without any knowledge of the endogenous ligand for this GPCR. By focusing on assay techniques that are based upon receptor function, and not compound binding affinity, we are able to ascertain compounds that are able to reduce the functional activity of this receptor (Compound A) as well as increase the functional activity of the receptor (Compound B). Based upon the location of these receptor in lung tissue (see, for example, hRUP13 and hRUP21 in Example 6), pharmaceutical agents can be developed for potential therapeutic treatment of lung cancer.

References cited throughout this patent document, including co-pending and related patent applications, unless otherwise indicated, are fully incorporated herein by reference. Modifications and extension of the disclosed inventions that are within the purview of the skilled artisan are encompassed within the above disclosure and the claims that follow.

Although a variety of expression vectors are available to those in the art, for purposes of utilization for both the endogenous and non-endogenous human GPCRs, it is most preferred that the vector utilized be pCMV. This vector was deposited with the American Type Culture Collection (ATCC) on October 13, 1998 (10801 University

Blvd., Manassas, VA 20110-2209 USA) under the provisions of the Budapest Treaty for the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure. The DNA was tested by the ATCC and determined to be viable. The ATCC has assigned the following deposit number to pCMV: ATCC #203351.

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CLAIMS

What is claimed is:

1. A G protein-coupled receptor encoded by an amino acid sequence of
5 SEQ.ID.NO.:2.
2. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 1.
3. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:1.
4. A host cell comprising the plasmid of claim 3.
- 10 5. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:4.
6. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 5.
7. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:3.
- 15 8. A host cell comprising the plasmid of claim 7.
9. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:6.
10. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 9.
- 20 11. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:5.
12. A host cell comprising the plasmid of claim 11.
13. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:8.
- 25 14. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 13.

15. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:7.
16. A host cell comprising the plasmid of claim 15.
17. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:10.
- 5 18. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 17.
19. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:9.
20. A host cell comprising the plasmid of claim 19.
21. A G protein-coupled receptor encoded by an amino acid sequence of
10 SEQ.ID.NO.:12.
22. A non-endogenous, constitutively activated version of the G-protein-coupled
receptor of claim 21 comprising an amino acid sequence of SEQ.ID.NO.84.
23. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:11.
24. A host cell comprising the plasmid of claim 23.
- 15 25. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:14.
26. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 25 comprising an amino acid sequence of SEQ.ID.NO.88.
27. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:13.
- 20 28. A host cell comprising the plasmid of claim 27.
29. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:16.
30. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 29 comprising an amino acid sequence of SEQ.ID.NO.:92.
- 25 31. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:15.

32. A host cell comprising the plasmid of claim 31.
33. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:18.
34. A non-endogenous, constitutively activated version of the G protein-coupled
5 receptor of claim 33.
35. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:17.
36. A host cell comprising the plasmid of claim 35.
37. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:20.
- 10 38. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 37.
39. A plasmid comprising a vector and the cDNA of SE.ID.NO.:19.
40. A host cell comprising the plasmid of claim 39.
41. A G protein-coupled receptor encoded by an amino acid sequence of
15 SEQ.ID.NO.:22.
42. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 41.
43. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:21.
44. A host cell comprising the plasmid of claim 43.
- 20 45. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:24.
46. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 45.
47. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:23.
- 25 48. A host cell comprising the plasmid of claim 47.

49. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:26.
50. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 49.
- 5 51. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:25.
52. A host cell comprising the plasmid of claim 51.
53. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:28.
54. A non-endogenous, constitutively activated version of the G protein-coupled
10 receptor of claim 53.
55. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:27.
56. A host cell comprising the plasmid of claim 55.
57. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:30.
- 15 58. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 57.
59. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:29.
60. A host cell comprising the plasmid of claim 59.
61. A G protein-coupled receptor encoded by an amino acid sequence of
20 SEQ.ID.NO.:32.
62. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 61 comprising an amino acid sequence of SEQ.ID.NO.:96.
63. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:95.
64. A host cell comprising the plasmid of claim 63.

65. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:34.

66. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 65.

5 67. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:33.

68. A host cell comprising the plasmid of claim 67.

69. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:36.

70. A non-endogenous, constitutively activated version of the G protein-coupled
10 receptor of claim 69.

71. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:35.

72. A host cell comprising the plasmid of claim 71.

73. A G protein-coupled receptor encoded by an amino acid sequence of
SEQ.ID.NO.:38.

15 74. A non-endogenous, constitutively activated version of the G protein-coupled
receptor of claim 73.

75. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:37.

76. A host cell comprising the plasmid of claim 75.

77. A G protein-coupled receptor encoded by an amino acid sequence of
20 SEQ.ID.NO.:40.

78. A non-endogenous, constitutively activated version of the G protein-coupled
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79. A plasmid comprising a vector and the cDNA of SEQ.ID.NO.:39.

80. A host cell comprising the plasmid of claim 79.

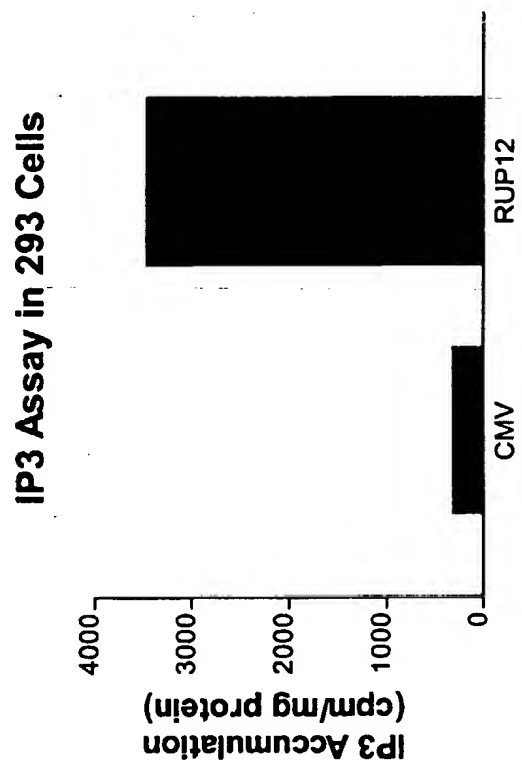


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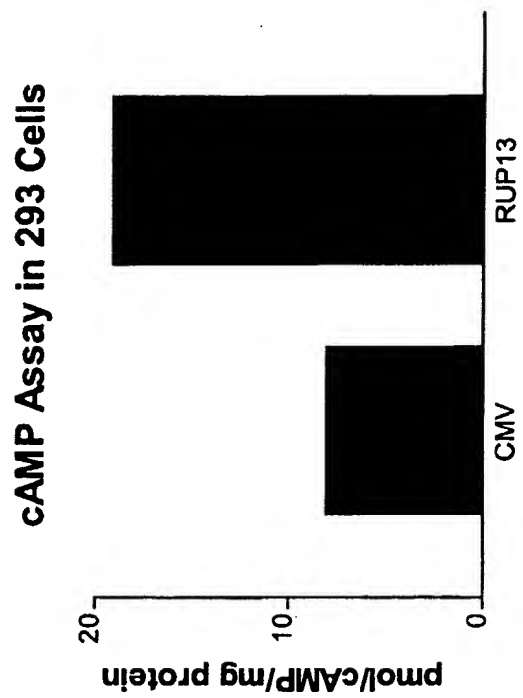


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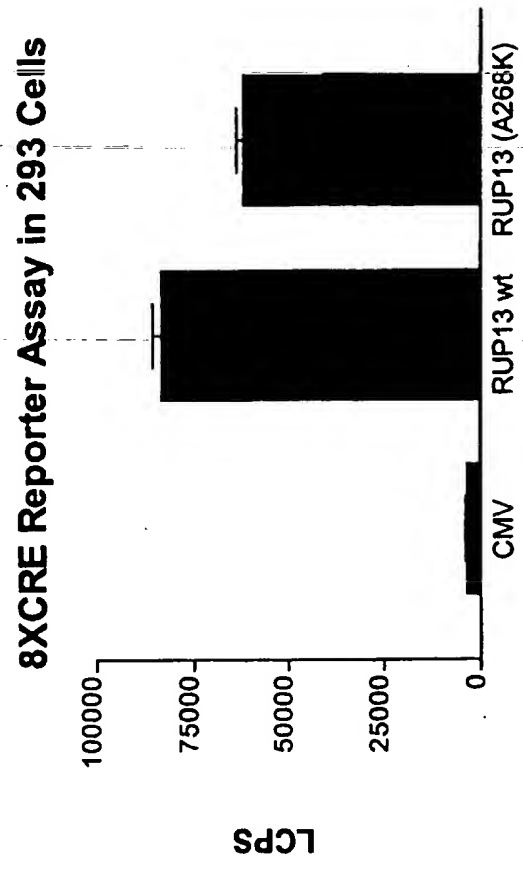


Figure 3

GTP Binding Assay
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membrane)

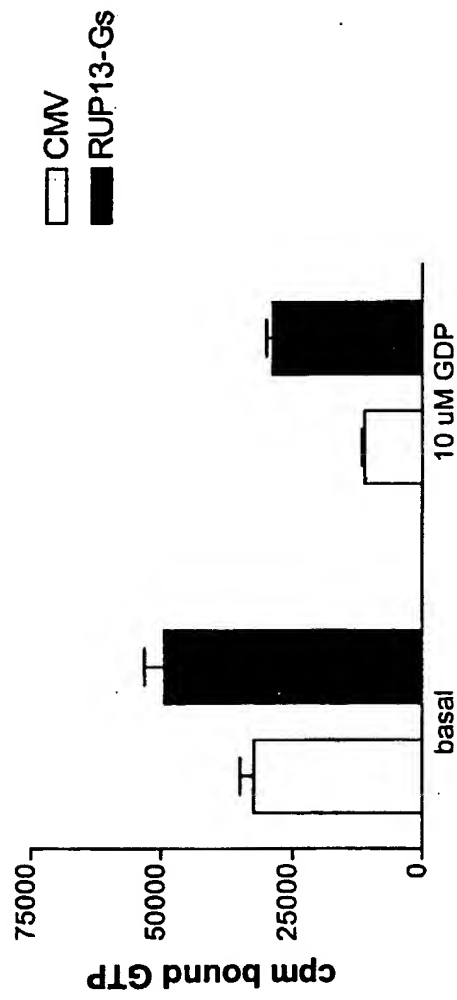


Figure 4

8XCRE Reporter Assay in 293 Cells

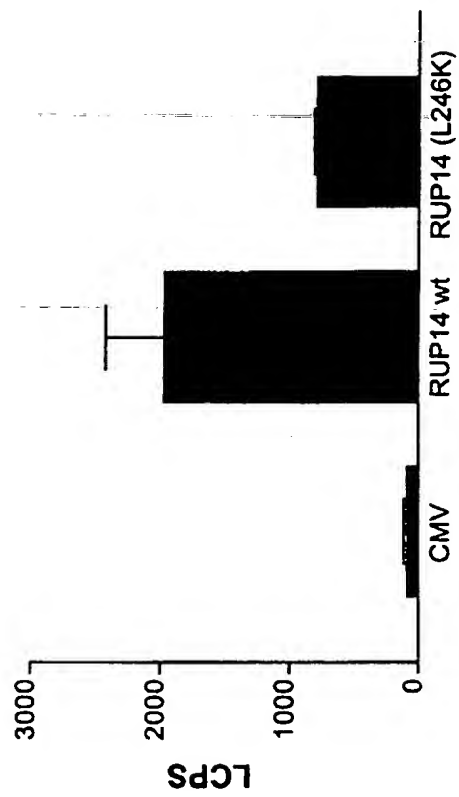


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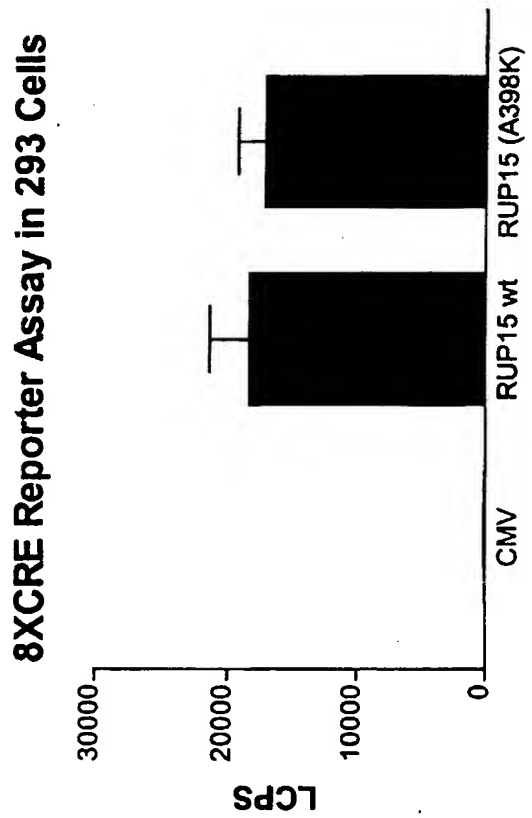


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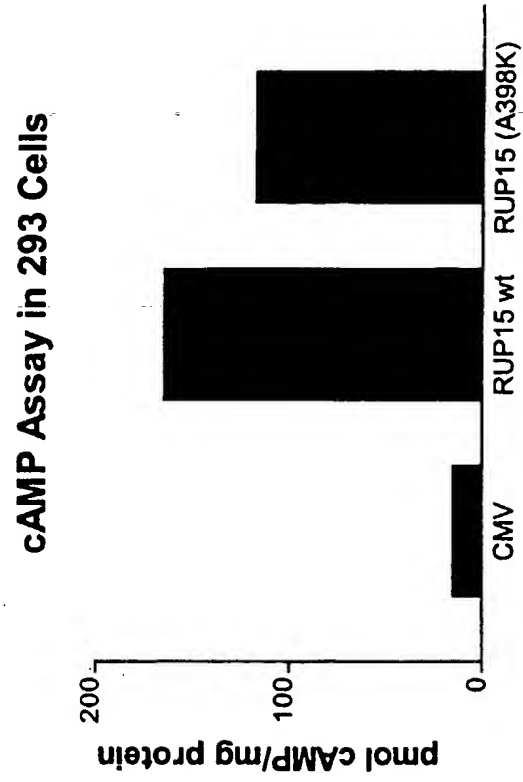


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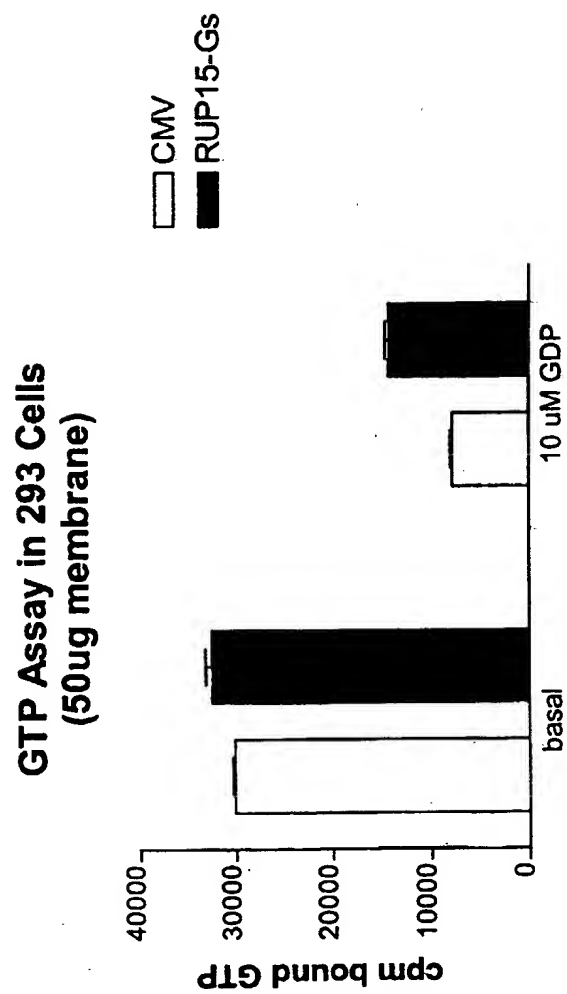


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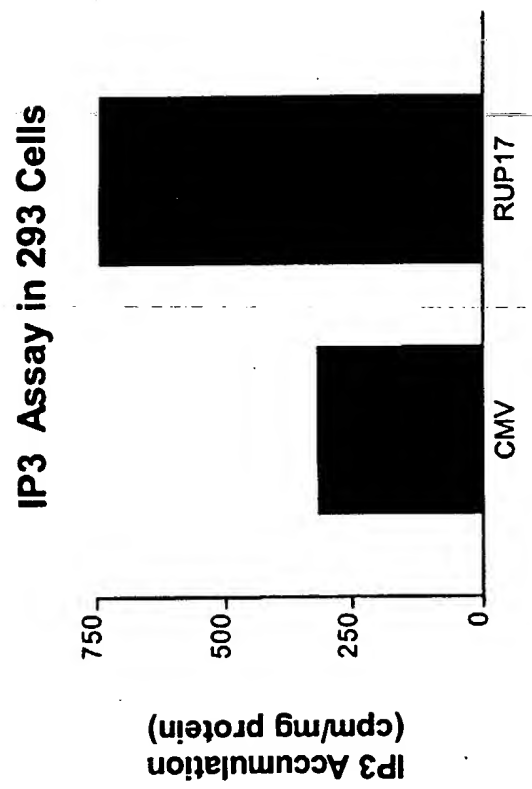


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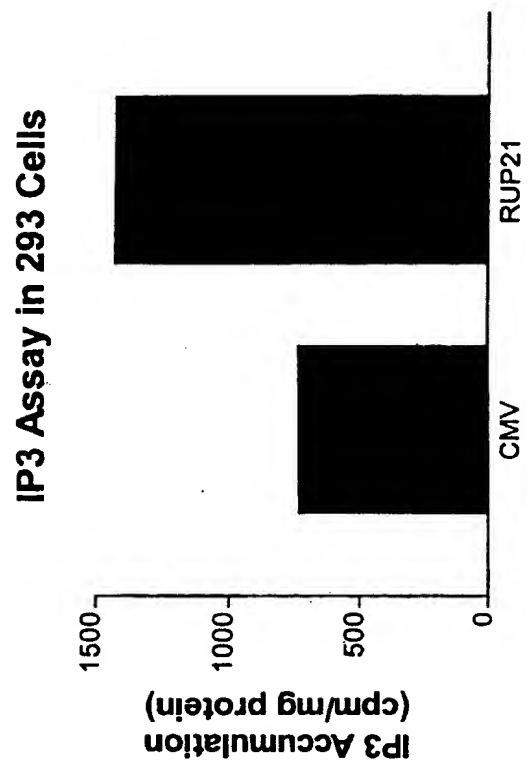


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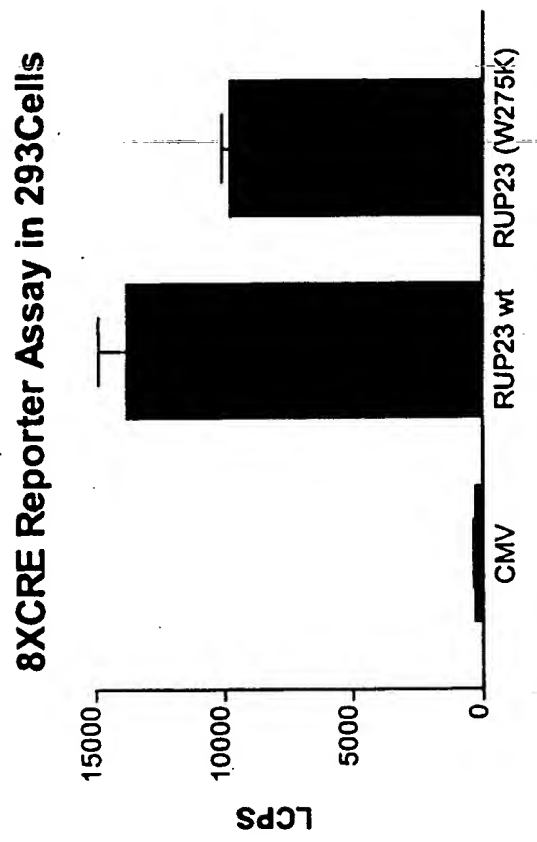


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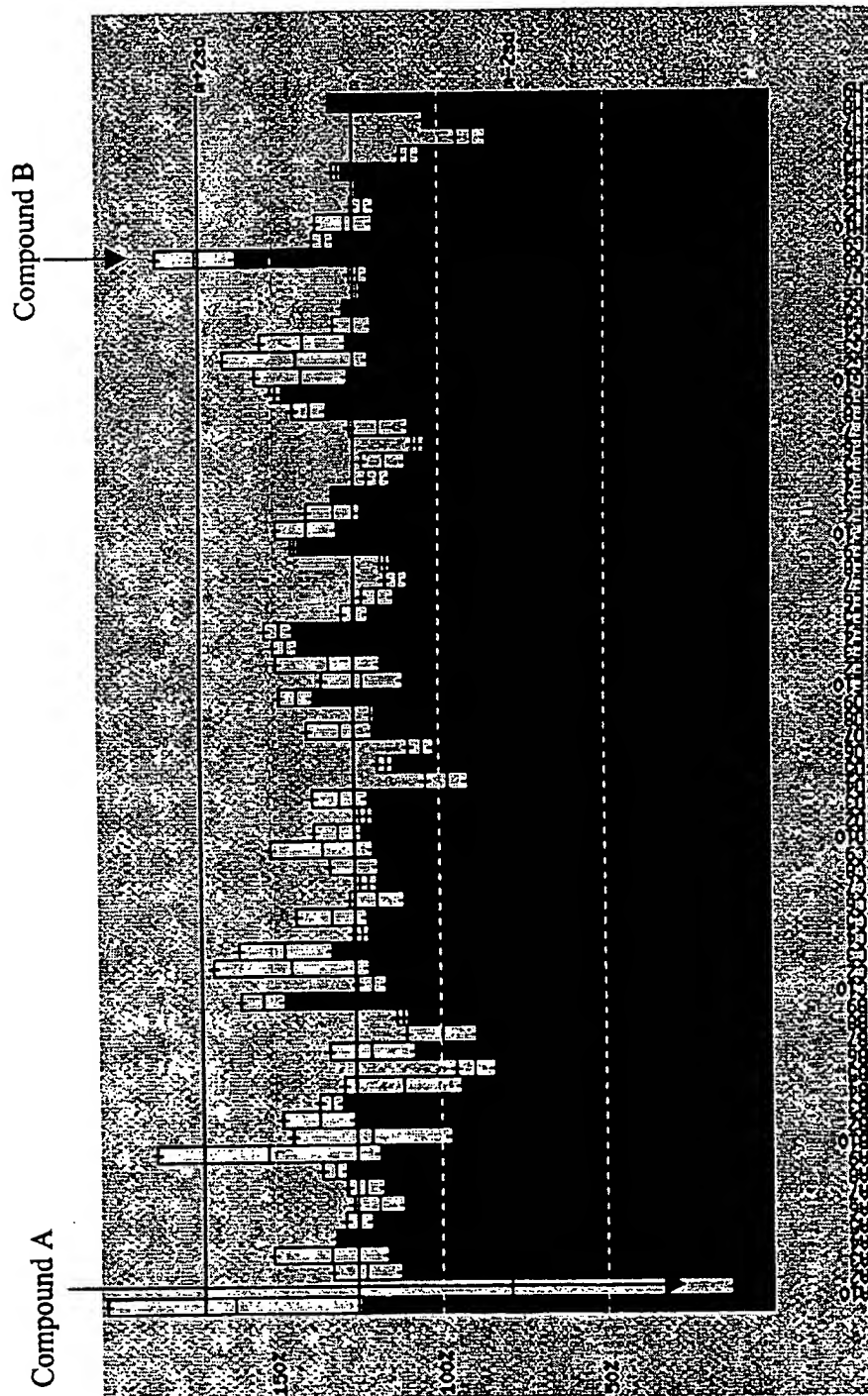


Figure 12

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Chen, Rupong
Dang, Huong T.
Lowitz, Kevin P.

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130					135					140					
Leu	Pro	Leu	Arg	Val	Asp	Tyr	Tyr	Leu	Leu	His	Glu	Thr	Trp	Arg	Phe
145					150					155			160		
Gly	Ala	Ala	Ala	Cys	Lys	Val	Asn	Leu	Phe	Met	Leu	Ser	Thr	Asn	Arg
165							170					175			
Thr	Ala	Ser	Val	Val	Phe	Leu	Thr	Ala	Ile	Ala	Leu	Asn	Arg	Tyr	Leu
180							185					190			
Lys	Val	Val	Gln	Pro	His	His	Val	Leu	Ser	Arg	Ala	Ser	Val	Gly	Ala
195							200					205			
Ala	Ala	Arg	Val	Ala	Gly	Gly	Leu	Trp	Val	Gly	Ile	Leu	Leu	Leu	Asn
210					215					220					
Gly	His	Leu	Leu	Leu	Ser	Thr	Phe	Ser	Gly	Pro	Ser	Cys	Leu	Ser	Tyr
225					230					235			240		
Arg	Val	Gly	Thr	Lys	Pro	Ser	Ala	Ser	Leu	Arg	Trp	His	Gln	Ala	Leu
245							250					255			
Tyr	Leu	Leu	Glu	Phe	Phe	Leu	Pro	Leu	Ala	Leu	Ile	Leu	Phe	Ala	Ile
260							265					270			
Val	Ser	Ile	Gly	Leu	Thr	Ile	Arg	Asn	Arg	Gly	Leu	Gly	Gly	Gln	Ala
275							280				285				
Gly	Pro	Gln	Arg	Ala	Met	Arg	Val	Leu	Ala	Met	Val	Val	Ala	Val	Tyr
290					295					300					
Thr	Ile	Cys	Phe	Leu	Pro	Ser	Ile	Ile	Phe	Gly	Met	Ala	Ser	Met	Val
305					310					315			320		
Ala	Phe	Trp	Leu	Ser	Ala	Cys	Arg	Ser	Leu	Asp	Leu	Cys	Thr	Gln	Leu
325							330					335			
Phe	His	Gly	Ser	Leu	Ala	Phe	Thr	Tyr	Leu	Asn	Ser	Val	Leu	Asp	Pro
340							345				350				
Val	Leu	Tyr	Cys	Phe	Ser	Ser	Pro	Asn	Phe	Leu	His	Gln	Ser	Arg	Ala
355							360				365				

Leu Leu Gly Leu Thr Arg Gly Arg Gln Gly Pro Val Ser Asp Glu Ser
 370 375 380
 Ser Tyr Gln Pro Ser Arg Gln Trp Arg Tyr Arg Glu Ala Ser Arg Lys
 385 390 395 400
 Ala Glu Ala Ile Gly Lys Leu Lys Val Gln Gly Glu Val Ser Leu Glu
 405 410 415
 Lys Glu Gly Ser Ser Gln Gly
 420

<210> 9
 <211> 966
 <212> DNA
 <213> Homo sapiens

<400> 9
 atgaaccaga ctttgaatag cagtgggacc gtggagtcag ccctaaacta ttccagaggg 60
 agcacagtgc acacggccta cctggtgctg agctccctgg ccatgttcac ctgcctgtgc 120
 gggatggcag gcaacagcat ggtgatctgg ctgctgggct ttcgaatgca caggaacccc 180
 ttctgcatct atatcctcaa cctggcggca gccgacctcc tcttcctctt cagcatggct 240
 tccacgctca gcctggaaac ccagcccctg gtcaatacca ctgacaaggc ccacgagctg 300
 atgaagagac tgatgtactt tgctacaca gtgggcctga gcctgctgac ggccatcagc 360
 acccagcgct gtctctctgt cctcttcctt atctggttca agtgtcaccg gcccaggcac 420
 ctgtcagcct ggggtgtgtg cctgctgtgg acactctgtc tcctgatgaa cgggttgacc 480
 tcttccttct gcagcaagtt cttgaaattc aatgaagatc ggtgcttcag ggtggacatg 540
 gtccaggccg cctcatcat gggggcttta accccagtga tgactctgtc cagcctgacc 600
 ctctttgtct ggggtgaggag gagctcccag cagtggcggc ggagcccac acggctgttc 660
 gtggtggtcc tggcctctgt cctggtgttc ctcatctgtt ccctgcctct gagcatctac 720
 tggtttgtgc tctactggtt gagcctgccg cccgagatgc aggtcctgtg cttcagcttg 780
 tcacgcctct cctcgtccgt aagcagcagc gccaaacccc tcacttactt cctggtgggc 840
 agccggagga gccacaggct gccaccagg tccctgggga ctgtgctcca acaggcgctt 900
 cgcgaggagc ccgagctgga aggtggggag acgcccaccg tgggcaccaa tgagatgggg 960
 gcttga 966

<210> 10
 <211> 321
 <212> PRT
 <213> Homo sapiens

<400> 10

Met Asn Gln Thr Leu Asn Ser Ser Gly Thr Val Glu Ser Ala Leu Asn
 1 5 10 15
 Tyr Ser Arg Gly Ser Thr Val His Thr Ala Tyr Leu Val Leu Ser Ser
 20 25 30

Leu Ala Met Phe Thr Cys Leu Cys Gly Met Ala Gly Asn Ser Met Val
 35 40 45
 Ile Trp Leu Leu Gly Phe Arg Met His Arg Asn Pro Phe Cys Ile Tyr
 50 55 60
 Ile Leu Asn Leu Ala Ala Asp Leu Leu Phe Leu Phe Ser Met Ala
 65 70 75 80
 Ser Thr Leu Ser Leu Glu Thr Gln Pro Leu Val Asn Thr Thr Asp Lys
 85 90 95
 Val His Glu Leu Met Lys Arg Leu Met Tyr Phe Ala Tyr Thr Val Gly
 100 105 110
 Leu Ser Leu Leu Thr Ala Ile Ser Thr Gln Arg Cys Leu Ser Val Leu
 115 120 125
 Phe Pro Ile Trp Phe Lys Cys His Arg Pro Arg His Leu Ser Ala Trp
 130 135 140
 Val Cys Gly Leu Leu Trp Thr Leu Cys Leu Leu Met Asn Gly Leu Thr
 145 150 155 160
 Ser Ser Phe Cys Ser Lys Phe Leu Lys Phe Asn Glu Asp Arg Cys Phe
 165 170 175
 Arg Val Asp Met Val Gln Ala Ala Leu Ile Met Gly Val Leu Thr Pro
 180 185 190
 Val Met Thr Leu Ser Ser Leu Thr Leu Phe Val Trp Val Arg Arg Ser
 195 200 205
 Ser Gln Gln Trp Arg Arg Gln Pro Thr Arg Leu Phe Val Val Val Leu
 210 215 220
 Ala Ser Val Leu Val Phe Leu Ile Cys Ser Leu Pro Leu Ser Ile Tyr
 225 230 235 240
 Trp Phe Val Leu Tyr Trp Leu Ser Leu Pro Pro Glu Met Gln Val Leu
 245 250 255
 Cys Phe Ser Leu Ser Arg Leu Ser Ser Ser Val Ser Ser Ser Ala Asn
 260 265 270
 Pro Val Ile Tyr Phe Leu Val Gly Ser Arg Arg Ser His Arg Leu Pro
 275 280 285
 Thr Arg Ser Leu Gly Thr Val Leu Gln Gln Ala Leu Arg Glu Glu Pro
 290 295 300
 Glu Leu Glu Gly Gly Glu Thr Pro Thr Val Gly Thr Asn Glu Met Gly
 305 310 315 320

Ala

<210> 11
 <211> 1356
 <212> DNA
 <213> Homo sapiens

<400> 11
 atggagtcct caccatccc ccagtcacga gggaactctt ccactttggg gagggtcctt

60

caaaccacag gtccctctac tgccagtggg gtcccggagg tggggctacg ggatgttgct 120
 tcggaatctg tggccctctt cttcatgctc ctgctggact tgactgctgt ggctggcaat 180
 gccgctgtga tggccgtgat cgccaagacg cctgccctcc gaaaatttgt cttcgtcttc 240
 cacctctgcc tgggtggacct gctggctgcc ctgacctca tgcccctggc catgctctcc 300
 agctctgccc tctttgacca cgccctcttt ggggaggtgg cctgccgcct ctacttgttt 360
 ctgagcgtgt gctttgtcag cctggccatc ctctcggtgt cagccatcaa tgtggagcgc 420
 tactattacg tagtccaccc catgcgctac gaggtgcgca tgacgctggg gctggtggcc 480
 tctgtgctgg tgggtgtgtg ggtgaaggcc ttggccatgg cttctgtgcc agtgttggga 540
 aggttctcct gggaggaagg agctcccagt gtccccccag gctgttact ccagtggagc 600
 cacagtgcct actgccagct ttttgtggtg gtctttgctg tctttactt tctgttggcc 660
 ctgctcctca tacttgtggt ctactgcagc atgttccgag tggcccgct ggctgccatg 720
 cagcacgggc cgctgccac gtggatggag acaccccggc aacgctccga atctctcagc 780
 agccgctcca cgatggtcac cagctcgggg gcccccaga ccacccaca ccggacgttt 840
 gggggaggga aagcagcagt ggttctcctg gctgtggggg gacagttcct gctctgttg 900
 ttgccctact tctctttcca cctctatgtt gccctgagt ctcagcccat ttcaactggg 960
 caggtggaga gtgtggtcac ctggattggc tacttttget tcaattccaa cctttcttc 1020
 tatggatgtc tcaaccggca gatccggggg gagctcagca agcagtttgt ctgcttcttc 1080
 aagccagctc cagaggagga gctgaggctg cctagccggg agggctccat tgaggagaac 1140
 ttctgcagt tcttcaggg gactggctgt cctctgagt cctgggtttc ccgacccta 1200
 cccagcccca agcaggagcc acctgctgtt gactttcgaa tcccaggcca gatagctgag 1260
 gagacctctg agttcctgga gcagcaactc accagcgaca tcatcatgtc agacagctac 1320
 ctccgtcctg ccgcctcacc ccggctggag tcatga 1356

<210> 12
 <211> 451
 <212> PRT
 <213> Homo sapiens

<400> 12

Met Glu Ser Ser Pro Ile Pro Gln Ser Ser Gly Asn Ser Ser Thr Leu
1 5 10 15

Gly Arg Val Pro Gln Thr Pro Gly Pro Ser Thr Ala Ser Gly Val Pro
20 25 30

Glu Val Gly Leu Arg Asp Val Ala Ser Glu Ser Val Ala Leu Phe Phe
35 40 45

Met Leu Leu Leu Asp Leu Thr Ala Val Ala Gly Asn Ala Ala Val Met
50 55 60

Ala Val Ile Ala Lys Thr Pro Ala Leu Arg Lys Phe Val Phe Val Phe
 65 70 75 80
 His Leu Cys Leu Val Asp Leu Leu Ala Ala Leu Thr Leu Met Pro Leu
 85 90 95
 Ala Met Leu Ser Ser Ser Ala Leu Phe Asp His Ala Leu Phe Gly Glu
 100 105 110
 Val Ala Cys Arg Leu Tyr Leu Phe Leu Ser Val Cys Phe Val Ser Leu
 115 120 125
 Ala Ile Leu Ser Val Ser Ala Ile Asn Val Glu Arg Tyr Tyr Tyr Val
 130 135 140
 Val His Pro Met Arg Tyr Glu Val Arg Met Thr Leu Gly Leu Val Ala
 145 150 155 160
 Ser Val Leu Val Gly Val Trp Val Lys Ala Leu Ala Met Ala Ser Val
 165 170 175
 Pro Val Leu Gly Arg Val Ser Trp Glu Glu Gly Ala Pro Ser Val Pro
 180 185 190
 Pro Gly Cys Ser Leu Gln Trp Ser His Ser Ala Tyr Cys Gln Leu Phe
 195 200 205
 Val Val Val Phe Ala Val Leu Tyr Phe Leu Leu Pro Leu Leu Leu Ile
 210 215 220
 Leu Val Val Tyr Cys Ser Met Phe Arg Val Ala Arg Val Ala Ala Met
 225 230 235 240
 Gln His Gly Pro Leu Pro Thr Trp Met Glu Thr Pro Arg Gln Arg Ser
 245 250 255
 Glu Ser Leu Ser Ser Arg Ser Thr Met Val Thr Ser Ser Gly Ala Pro
 260 265 270
 Gln Thr Thr Pro His Arg Thr Phe Gly Gly Gly Lys Ala Ala Val Val
 275 280 285
 Leu Leu Ala Val Gly Gly Gln Phe Leu Leu Cys Trp Leu Pro Tyr Phe
 290 295 300
 Ser Phe His Leu Tyr Val Ala Leu Ser Ala Gln Pro Ile Ser Thr Gly
 305 310 315 320
 Gln Val Glu Ser Val Val Thr Trp Ile Gly Tyr Phe Cys Phe Thr Ser
 325 330 335
 Asn Pro Phe Phe Tyr Gly Cys Leu Asn Arg Gln Ile Arg Gly Glu Leu
 340 345 350
 Ser Lys Gln Phe Val Cys Phe Phe Lys Pro Ala Pro Glu Glu Glu Leu
 355 360 365
 Arg Leu Pro Ser Arg Glu Gly Ser Ile Glu Glu Asn Phe Leu Gln Phe
 370 375 380
 Leu Gln Gly Thr Gly Cys Pro Ser Glu Ser Trp Val Ser Arg Pro Leu
 385 390 395 400
 Pro Ser Pro Lys Gln Glu Pro Pro Ala Val Asp Phe Arg Ile Pro Gly
 405 410 415

Gln Ile Ala Glu Glu Thr Ser Glu Phe Leu Glu Gln Gln Leu Thr Ser
 420 425 430

Asp Ile Ile Met Ser Asp Ser Tyr Leu Arg Pro Ala Ala Ser Pro Arg
 435 440 445

Leu Glu Ser
 450

<210> 13
 <211> 1041
 <212> DNA
 <213> Homo sapiens

<400> 13
 atggagagaa aatttatgtc cttgcaacca tccatctccg tatcagaaat ggaaccaaatt 60
 ggcaccttca gcaataacaa cagcaggaac tgcacaattg aaaacttcaa gagagaattt 120
 ttccaattg tataatctgat aatatttttc tggggagtct tgggaaatgg gttgtccata 180
 tatgttttcc tgcagcctta taagaagtcc acatctgtga acgttttcat gctaaatctg 240
 gccatttcag atctcctgtt cataagcacg cttcccttca gggctgacta ttatcttaga 300
 ggctccaatt ggatatttgg agacctggcc tgcaggatta tgtcttattc cttgtatgtc 360
 aacatgtaca gcagtattta tttcctgacc gtgctgagtg ttgtgcgttt cctggcaatg 420
 gttcacccct ttcggcttct gcatgtcacc agcatcagga gtgcctggat cctctgtggg 480
 atcatatgga tccttatcat ggcttcctca ataatgtccc tggacagtgg ctctgagcag 540
 aacggcagtg tcacatcatg cttagagctg aatctctata aaattgctaa gctgcagacc 600
 atgaactata ttgccttggg ggtgggctgc ctgctgccat ttttcacact cagcatctgt 660
 tatctgctga tcattcgggt tctgttaaaa gtggaggtcc cagaatcggg gctgcggggt 720
 tctcacagga aggcactgac caccatcatc atcaccttga tcattcttctt cttgtgtttc 780
 ctgccctatc acacactgag gaccgtccac ttgacgacat ggaaagtggg tttatgcaaa 840
 gacagactgc ataaagcttt ggttatcaca ctggccttgg cagcagccaa tgctgtcttc 900
 aatcctctgc tctattactt tgctggggag aattttaagg acagactaaa gtctgcactc 960
 agaaaaggcc atccacagaa ggcaaagaca aagtgtgttt tcctgttag tgtgtggttg 1020
 agaaaggaaa caagagtata a 1041

<210> 14
 <211> 346
 <212> PRT
 <213> Homo sapiens

<400> 14

Met Glu Arg Lys Phe Met Ser Leu Gln Pro Ser Ile Ser Val Ser Glu
 1 5 10 15

Met Glu Pro Asn Gly Thr Phe Ser Asn Asn Asn Ser Arg Asn Cys Thr
 20 25 30

Ile Glu Asn Phe Lys Arg Glu Phe Phe Pro Ile Val Tyr Leu Ile Ile
 35 40 45
 Phe Phe Trp Gly Val Leu Gly Asn Gly Leu Ser Ile Tyr Val Phe Leu
 50 55 60
 Gln Pro Tyr Lys Lys Ser Thr Ser Val Asn Val Phe Met Leu Asn Leu
 65 70 75 80
 Ala Ile Ser Asp Leu Leu Phe Ile Ser Thr Leu Pro Phe Arg Ala Asp
 85 90 95
 Tyr Tyr Leu Arg Gly Ser Asn Trp Ile Phe Gly Asp Leu Ala Cys Arg
 100 105 110
 Ile Met Ser Tyr Ser Leu Tyr Val Asn Met Tyr Ser Ser Ile Tyr Phe
 115 120 125
 Leu Thr Val Leu Ser Val Val Arg Phe Leu Ala Met Val His Pro Phe
 130 135 140
 Arg Leu Leu His Val Thr Ser Ile Arg Ser Ala Trp Ile Leu Cys Gly
 145 150 155 160
 Ile Ile Trp Ile Leu Ile Met Ala Ser Ser Ile Met Leu Leu Asp Ser
 165 170 175
 Gly Ser Glu Gln Asn Gly Ser Val Thr Ser Cys Leu Glu Leu Asn Leu
 180 185 190
 Tyr Lys Ile Ala Lys Leu Gln Thr Met Asn Tyr Ile Ala Leu Val Val
 195 200 205
 Gly Cys Leu Leu Pro Phe Phe Thr Leu Ser Ile Cys Tyr Leu Leu Ile
 210 215 220
 Ile Arg Val Leu Leu Lys Val Glu Val Pro Glu Ser Gly Leu Arg Val
 225 230 235 240
 Ser His Arg Lys Ala Leu Thr Thr Ile Ile Ile Thr Leu Ile Ile Phe
 245 250 255
 Phe Leu Cys Phe Leu Pro Tyr His Thr Leu Arg Thr Val His Leu Thr
 260 265 270
 Thr Trp Lys Val Gly Leu Cys Lys Asp Arg Leu His Lys Ala Leu Val
 275 280 285
 Ile Thr Leu Ala Leu Ala Ala Ala Asn Ala Cys Phe Asn Pro Leu Leu
 290 295 300
 Tyr Tyr Phe Ala Gly Glu Asn Phe Lys Asp Arg Leu Lys Ser Ala Leu
 305 310 315 320
 Arg Lys Gly His Pro Gln Lys Ala Lys Thr Lys Cys Val Phe Pro Val
 325 330 335
 Ser Val Trp Leu Arg Lys Glu Thr Arg Val
 340 345

<210> 15
 <211> 1527
 <212> DNA
 <213> Homo sapiens

<400> 15
atgacgtcca cctgcaccaa cagcacgcgc gagagtaaca gcagccacac gtgcatgccc 60
ctctccaaaa tgcccatcag cctggccccc ggcatcatcc gctcaaccgt gctgggtatc 120
ttcctcgccg cctctttcgt cggcaacata gtgctggcgc tagtggttga gcgcaagccg 180
cagctgctgc aggtgaccaa ccgttttatc tttaacctcc tcgtcaccga cctgctgcag 240
atttcgctcg tggcccccctg ggtgggtggc acctctgtgc ctctcttctg gccctcaac 300
agccacttct gcacggccct ggtagcctc acccactgt tcgccttcgc cagcgtcaac 360
accattgtcg tgggtgcagt ggatcgctac ttgtccatca tccacctct ctectaccg 420
tccaagatga ccagcgccg cggttacctg ctctctatg gcacctggat tgtggccatc 480
ctgcagagca ctctccact ctacggctgg ggccaggctg cctttgatga gcgcaatgct 540
ctctgctcca tgatctgggg ggccagcccc agctacacta ttctcagcgt ggtgtccttc 600
atcgtcattc cactgattgt catgattgcc tgctactccg tgggtgttctg tgcagcccgg 660
aggcagcatg ctctgctgta caatgtcaag agacacagct tggaagtgcg agtcaaggac 720
tgtgtggaga atgaggatga agaggagca gagaagaagg aggagttcca ggatgagagt 780
gagtttcgcc gccagcatga aggtgaggtc aaggccaagg agggcagaat ggaagccaag 840
gacggcagcc tgaaggccaa ggaaggaagc acggggacca gtgagagtag tgtagaggcc 900
aggggcagcg aggaggtcag agagagcagc acgggtggcca gcgacggcag catggagggt 960
aaggaaggca gcaccaaagt tgaggagaac agcatgaagg cagacaaggg tcgcacagag 1020
gtcaaccagt gcagcattga cttgggtgaa gatgacatgg agtttggtga agacgacatc 1080
aatttcagtg aggatgacgt cgaggcagtg aacatcccgg agagcctccc acccagtcgt 1140
cgtaacagca acagcaaccc tcctctgccc aggtgctacc agtgcaaagc tgctaaagtg 1200
atcttcatca tcattttctc ctatgtgcta tccttggggc cctactgctt tttagcagtc 1260
ctggccgtgt ggggtgatgt cgaaaccag gtaccccagt ggggtgatcac cataatcatc 1320
tggcttttct tcctgcagtg ctgcatccac ccctatgtct atggctacat gcacaagacc 1380
attaagaagg aaatccagga catgctgaag aagttcttct gcaaggaaaa gccccgaaa 1440
gaagatagcc acccagacct gcccggaaca gaggggtggga ctgaaggcaa gattgtccct 1500
tcctacgatt ctgctacttt tccttga 1527

<210> 16
<211> 508
<212> PRT
<213> Homo sapiens

<400> 16

Met Thr Ser Thr Cys Thr Asn Ser Thr Arg Glu Ser Asn Ser Ser His
1 5 10 15

Thr Cys Met Pro Leu Ser Lys Met Pro Ile Ser Leu Ala His Gly Ile
 20 25 30
 Ile Arg Ser Thr Val Leu Val Ile Phe Leu Ala Ala Ser Phe Val Gly
 35 40 45
 Asn Ile Val Leu Ala Leu Val Leu Gln Arg Lys Pro Gln Leu Leu Gln
 50 55 60
 Val Thr Asn Arg Phe Ile Phe Asn Leu Leu Val Thr Asp Leu Leu Gln
 65 70 75 80
 Ile Ser Leu Val Ala Pro Trp Val Val Ala Thr Ser Val Pro Leu Phe
 85 90 95
 Trp Pro Leu Asn Ser His Phe Cys Thr Ala Leu Val Ser Leu Thr His
 100 105 110
 Leu Phe Ala Phe Ala Ser Val Asn Thr Ile Val Val Val Ser Val Asp
 115 120 125
 Arg Tyr Leu Ser Ile Ile His Pro Leu Ser Tyr Pro Ser Lys Met Thr
 130 135 140
 Gln Arg Arg Gly Tyr Leu Leu Leu Tyr Gly Thr Trp Ile Val Ala Ile
 145 150 155 160
 Leu Gln Ser Thr Pro Pro Leu Tyr Gly Trp Gly Gln Ala Ala Phe Asp
 165 170 175
 Glu Arg Asn Ala Leu Cys Ser Met Ile Trp Gly Ala Ser Pro Ser Tyr
 180 185 190
 Thr Ile Leu Ser Val Val Ser Phe Ile Val Ile Pro Leu Ile Val Met
 195 200 205
 Ile Ala Cys Tyr Ser Val Val Phe Cys Ala Ala Arg Arg Gln His Ala
 210 215 220
 Leu Leu Tyr Asn Val Lys Arg His Ser Leu Glu Val Arg Val Lys Asp
 225 230 235 240
 Cys Val Glu Asn Glu Asp Glu Glu Gly Ala Glu Lys Lys Glu Glu Phe
 245 250 255
 Gln Asp Glu Ser Glu Phe Arg Arg Gln His Glu Gly Glu Val Lys Ala
 260 265 270
 Lys Glu Gly Arg Met Glu Ala Lys Asp Gly Ser Leu Lys Ala Lys Glu
 275 280 285
 Gly Ser Thr Gly Thr Ser Glu Ser Ser Val Glu Ala Arg Gly Ser Glu
 290 295 300
 Glu Val Arg Glu Ser Ser Thr Val Ala Ser Asp Gly Ser Met Glu Gly
 305 310 315 320
 Lys Glu Gly Ser Thr Lys Val Glu Glu Asn Ser Met Lys Ala Asp Lys
 325 330 335
 Gly Arg Thr Glu Val Asn Gln Cys Ser Ile Asp Leu Gly Glu Asp Asp
 340 345 350
 Met Glu Phe Gly Glu Asp Asp Ile Asn Phe Ser Glu Asp Asp Val Glu
 355 360 365

Ala Val Asn Ile Pro Glu Ser Leu Pro Pro Ser Arg Arg Asn Ser Asn
 370 375 380

Ser Asn Pro Pro Leu Pro Arg Cys Tyr Gln Cys Lys Ala Ala Lys Val
 385 390 395 400

Ile Phe Ile Ile Ile Phe Ser Tyr Val Leu Ser Leu Gly Pro Tyr Cys
 405 410 415

Phe Leu Ala Val Leu Ala Val Trp Val Asp Val Glu Thr Gln Val Pro
 420 425 430

Gln Trp Val Ile Thr Ile Ile Ile Trp Leu Phe Phe Leu Gln Cys Cys
 435 440 445

Ile His Pro Tyr Val Tyr Gly Tyr Met His Lys Thr Ile Lys Lys Glu
 450 455 460

Ile Gln Asp Met Leu Lys Lys Phe Phe Cys Lys Glu Lys Pro Pro Lys
 465 470 475 480

Glu Asp Ser His Pro Asp Leu Pro Gly Thr Glu Gly Gly Thr Glu Gly
 485 490 495

Lys Ile Val Pro Ser Tyr Asp Ser Ala Thr Phe Pro
 500 505

<210> 17
 <211> 1068
 <212> DNA
 <213> Homo sapiens

<400> 17
 atgcccttga cggacggcat ttcttcattt gaggacctct tggctaacaa tctcctcaga 60
 atatttgtct gggttatagc ttccattacc tgctttggaa-atctttttgt cattggcatg 120
 agatctttca ttaaagctga aaatacaact cagcgtatgt ccatcaaaat cctttgttgc 180
 gctgattgcc tgatgggtgt ttacttgttc tttgttgga ttttcgatat aaaataccga 240
 gggcagtatc agaagtatgc cttgctgtgg atggagagcg tgcagtgccg cctcatgggg 300
 ttcttgcca tgctgtccac cgaagtctct gttctgctac tgacctactt gactttggag 360
 aagttcctgg tcattgtctt ccccttcagt aacattcgac ctggaaaacg gcagacctca 420
 gtcacctca tttgcatctg gatggcgga tttttaatag ctgtaattcc attttggaa 480
 aaggattatt ttggaaactt ttatgggaaa aatggagtat gtttccact ttattatgac 540
 caaacagaag atattggaag caaagggtat tctcttgga ttttcttagg tgtgaacttg 600
 ctggcttttc tcatcattgt gttttcctat attactatgt tctgttccat tcaaaaaacc 660
 gccttgca cccagaagt aaggaattgt tttggaagag aggtggctgt tgcaaatcgt 720
 ttctttttta tagtggtctc tgatgccatc tgctggattc ctgtatttgt agttaaaatc 780
 ctttccctct tccgggtgga aataccagac acaatgactt cctggatagt gatttttttc 840
 cttccagtta acagtgttt gaatccaatc ctctatactc tcacaaccaa cttttttaag 900
 gacaagttga aacagtgtct gcacaaacat cagaggaaat caattttcaa aattaaaaaa 960

aaaagtttat ctacatccat tgtgtggata gaggactcct cttccctgaa acttggggtt 1020
 ttgaacaaaa taacacttgg agacagtata atgaaaccag tttcctag 1068

<210> 18
 <211> 355
 <212> PRT
 <213> Homo sapiens

<400> 18

Met Pro Leu Thr Asp Gly Ile Ser Ser Phe Glu Asp Leu Leu Ala Asn
 1 5 10 15
 Asn Ile Leu Arg Ile Phe Val Trp Val Ile Ala Phe Ile Thr Cys Phe
 20 25 30
 Gly Asn Leu Phe Val Ile Gly Met Arg Ser Phe Ile Lys Ala Glu Asn
 35 40 45
 Thr Thr His Ala Met Ser Ile Lys Ile Leu Cys Cys Ala Asp Cys Leu
 50 55 60
 Met Gly Val Tyr Leu Phe Phe Val Gly Ile Phe Asp Ile Lys Tyr Arg
 65 70 75 80
 Gly Gln Tyr Gln Lys Tyr Ala Leu Leu Trp Met Glu Ser Val Gln Cys
 85 90 95
 Arg Leu Met Gly Phe Leu Ala Met Leu Ser Thr Glu Val Ser Val Leu
 100 105 110
 Leu Leu Thr Tyr Leu Thr Leu Glu Lys Phe Leu Val Ile Val Phe Pro
 115 120 125
 Phe Ser Asn Ile Arg Pro Gly Lys Arg Gln Thr Ser Val Ile Leu Ile
 130 135 140
 Cys Ile Trp Met Ala Gly Phe Leu Ile Ala Val Ile Pro Phe Trp Asn
 145 150 155 160
 Lys Asp Tyr Phe Gly Asn Phe Tyr Gly Lys Asn Gly Val Cys Phe Pro
 165 170 175
 Leu Tyr Tyr Asp Gln Thr Glu Asp Ile Gly Ser Lys Gly Tyr Ser Leu
 180 185 190
 Gly Ile Phe Leu Gly Val Asn Leu Leu Ala Phe Leu Ile Ile Val Phe
 195 200 205
 Ser Tyr Ile Thr Met Phe Cys Ser Ile Gln Lys Thr Ala Leu Gln Thr
 210 215 220
 Thr Glu Val Arg Asn Cys Phe Gly Arg Glu Val Ala Val Ala Asn Arg
 225 230 235 240
 Phe Phe Phe Ile Val Phe Ser Asp Ala Ile Cys Trp Ile Pro Val Phe
 245 250 255
 Val Val Lys Ile Leu Ser Leu Phe Arg Val Glu Ile Pro Asp Thr Met
 260 265 270
 Thr Ser Trp Ile Val Ile Phe Phe Leu Pro Val Asn Ser Ala Leu Asn

275 280 285
 Pro Ile Leu Tyr Thr Leu Thr Thr Asn Phe Phe Lys Asp Lys Leu Lys
 290 295 300
 Gln Leu Leu His Lys His Gln Arg Lys Ser Ile Phe Lys Ile Lys Lys
 305 310 315 320
 Lys Ser Leu Ser Thr Ser Ile Val Trp Ile Glu Asp Ser Ser Ser Leu
 325 330 335
 Lys Leu Gly Val Leu Asn Lys Ile Thr Leu Gly Asp Ser Ile Met Lys
 340 345 350
 Pro Val Ser
 355

<210> 19
 <211> 969
 <212> DNA
 <213> Homo sapiens

<400> 19
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 gggctgacag gaaacgcagt tgtgctctgg ctctctgggt gccgcatgcg caggaacgcc 180
 ttctccatct acatctctcaa cttggcgcga gcagacttcc tcttctcag cggccgcctt 240
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 gtgatgatgt tttcctactt tgcaggcctg agctttctga gtgccgtgag caccgagcgc 360
 tgcctgtccg tctgtggcc catctggtac cgtgcccacc gcccacaca cctgtcagcg 420
 gtggtgtgtg tctgtctctg ggccctgtcc ctgctgcgga gcactctgga gtggatgtta 480
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 gtcgcgtggc tgattttttt atgtgtggtt ctctgtgggt ccagcctggt cctgctgac 600
 aggattctct gtggatcccg gaagataaccg ctgaccaggc tgtacgtgac catcctgctc 660
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 tccgctctta acagcagtgc caaccccatc atttacttct tcgtgggctc ctttaggcag 840
 cgtcaaaata ggcagaacct gaagctggtt ctccagaggg ctctgcagga cgcgtctgag 900
 gtggatgaag gtggagggca gcttcctgag gaaatcctgg agctgtcggg aagcagattg 960
 gagcagtga 969

<210> 20
 <211> 322
 <212> PRT
 <213> Homo sapiens

<400> 20

Met Asp Pro Thr Ile Ser Thr Leu Asp Thr Glu Leu Thr Pro Ile Asn
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 Gly Thr Glu Glu Thr Leu Cys Tyr Lys Gln Thr Leu Ser Leu Thr Val
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 Leu Thr Cys Ile Val Ser Leu Val Gly Leu Thr Gly Asn Ala Val Val
 35 40 45
 Leu Trp Leu Leu Gly Cys Arg Met Arg Arg Asn Ala Phe Ser Ile Tyr
 50 55 60
 Ile Leu Asn Leu Ala Ala Ala Asp Phe Leu Phe Leu Ser Gly Arg Leu
 65 70 75 80
 Ile Tyr Ser Leu Leu Ser Phe Ile Ser Ile Pro His Thr Ile Ser Lys
 85 90 95
 Ile Leu Tyr Pro Val Met Met Phe Ser Tyr Phe Ala Gly Leu Ser Phe
 100 105 110
 Leu Ser Ala Val Ser Thr Glu Arg Cys Leu Ser Val Leu Trp Pro Ile
 115 120 125
 Trp Tyr Arg Cys His Arg Pro Thr His Leu Ser Ala Val Val Cys Val
 130 135 140
 Leu Leu Trp Ala Leu Ser Leu Leu Arg Ser Ile Leu Glu Trp Met Leu
 145 150 155 160
 Cys Gly Phe Leu Phe Ser Gly Ala Asp Ser Ala Trp Cys Gln Thr Ser
 165 170 175
 Asp Phe Ile Thr Val Ala Trp Leu Ile Phe Leu Cys Val Val Leu Cys
 180 185 190
 Gly Ser Ser Leu Val Leu Leu Ile Arg Ile Leu Cys Gly Ser Arg Lys
 195 200 205
 Ile Pro Leu Thr Arg Leu Tyr Val Thr Ile Leu Leu Thr Val Leu Val
 210 215 220
 Phe Leu Leu Cys Gly Leu Pro Phe Gly Ile Gln Phe Phe Leu Phe Leu
 225 230 235 240
 Trp Ile His Val Asp Arg Glu Val Leu Phe Cys His Val His Leu Val
 245 250 255
 Ser Ile Phe Leu Ser Ala Leu Asn Ser Ser Ala Asn Pro Ile Ile Tyr
 260 265 270
 Phe Phe Val Gly Ser Phe Arg Gln Arg Gln Asn Arg Gln Asn Leu Lys
 275 280 285
 Leu Val Leu Gln Arg Ala Leu Gln Asp Ala Ser Glu Val Asp Glu Gly
 290 295 300
 Gly Gly Gln Leu Pro Glu Glu Ile Leu Glu Leu Ser Gly Ser Arg Leu
 305 310 315 320
 Glu Gln

<210> 21
 <211> 1305

<212> DNA
<213> Homo sapiens

<400> 21
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agccgccgcg tccgcctggt gttcctgggg gtcatccttg tggtagcggt ggcaggcaac 180
accacagtgc tgtgccgcct gtgcggcgcc ggcggggccct gggcgggccc caagcgtcgc 240
aagatggact tcctgctggt gcagctggcc ctggcgagcc tgtacgcgtg cgggggcacg 300
gcgctgtcac agctggcctg ggaactgctg ggcgagcccc gcgcggccac gggggacctg 360
gcgtgccgct tcctgcagct gctgcaggca tccggggcgg gcgcctcggc ccacctcgtg 420
gtgctcatcg ccctcgagcg ccggcgcgcg gtgcgtcttc cgcacggccg gccgctgccc 480
gcgcgtgccc tcgccgccct gggctggctg ctggcactgc tgctggcgct gccccggccc 540
ttcgtggtgc gcggggactc cccctcgccg ctgccgccgc cgcgcgcccc aacgtccctg 600
cagccaggcg cgcgcccgcc cgcgcgcccc tggccggggg agcgtcgtg ccacgggatc 660
ttcgcgcccc tgccgcgtg gcacctgcag gtctacgctg tctacgagc cgtcgcgggc 720
ttcgtcgcgc ctgttacggt cctgggcgtc gcttgcgcc acctactctc cgtctggtgg 780
cggcaccggc cgcaggcccc cgcggctgca gcgcctggt cggcgagccc aggtcgagcc 840
cctgcgcccc gcgcgctgcc ccgcgccaag gtgcagagcc tgaagatgag cctgctgctg 900
gcgctgctgt tcgtgggctg cgagctgccc tactttgccg cccggctggc ggccgcgtgg 960
tcgtccgggc ccgcgggaga ctgggaggga gagggcctgt cggcggcgct gcgcgtggtg 1020
gcgatggcca acagcgctct caatcccttc gtctacctct tcttcaggc gggcgactgc 1080
cggctccggc gacagctgcg gaagcggctg ggctctctgt gctgcgcgcc gcagggaggc 1140
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cctcattatc accatgctcg gcgggaaccg ctggacgagg gcggcttgcg cccaccccct 1260
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<210> 22
<211> 434
<212> PRT
<213> Homo sapiens

<400> 22
Met Glu Asp Leu Phe Ser Pro Ser Ile Leu Pro Pro Ala Pro Asn Ile
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Ser Val Pro Ile Leu Leu Gly Trp Gly Leu Asn Leu Thr Leu Gly Gln
20 25 30
Gly Ala Pro Ala Ser Gly Pro Pro Ser Arg Arg Val Arg Leu Val Phe
35 40 45

Leu Gly Val Ile Leu Val Val Ala Val Ala Gly Asn Thr Thr Val Leu
 50 55 60
 Cys Arg Leu Cys Gly Gly Gly Gly Pro Trp Ala Gly Pro Lys Arg Arg
 65 70 75 80
 Lys Met Asp Phe Leu Leu Val Gln Leu Ala Leu Ala Asp Leu Tyr Ala
 85 90 95
 Cys Gly Gly Thr Ala Leu Ser Gln Leu Ala Trp Glu Leu Leu Gly Glu
 100 105 110
 Pro Arg Ala Ala Thr Gly Asp Leu Ala Cys Arg Phe Leu Gln Leu Leu
 115 120 125
 Gln Ala Ser Gly Arg Gly Ala Ser Ala His Leu Val Val Leu Ile Ala
 130 135 140
 Leu Glu Arg Arg Arg Ala Val Arg Leu Pro His Gly Arg Pro Leu Pro
 145 150 155 160
 Ala Arg Ala Leu Ala Ala Leu Gly Trp Leu Leu Ala Leu Leu Leu Ala
 165 170 175
 Leu Pro Pro Ala Phe Val Val Arg Gly Asp Ser Pro Ser Pro Leu Pro
 180 185 190
 Pro Pro Pro Pro Pro Thr Ser Leu Gln Pro Gly Ala Pro Pro Ala Ala
 195 200 205
 Arg Ala Trp Pro Gly Glu Arg Arg Cys His Gly Ile Phe Ala Pro Leu
 210 215 220
 Pro Arg Trp His Leu Gln Val Tyr Ala Phe Tyr Glu Ala Val Ala Gly
 225 230 235 240
 Phe Val Ala Pro Val Thr Val Leu Gly Val Ala Cys Gly His Leu Leu
 245 250 255
 Ser Val Trp Trp Arg His Arg Pro Gln Ala Pro Ala Ala Ala Ala Pro
 260 265 270
 Trp Ser Ala Ser Pro Gly Arg Ala Pro Ala Pro Ser Ala Leu Pro Arg
 275 280 285
 Ala Lys Val Gln Ser Leu Lys Met Ser Leu Leu Leu Ala Leu Leu Phe
 290 295 300
 Val Gly Cys Glu Leu Pro Tyr Phe Ala Ala Arg Leu Ala Ala Ala Trp
 305 310 315 320
 Ser Ser Gly Pro Ala Gly Asp Trp Glu Gly Glu Gly Leu Ser Ala Ala
 325 330 335
 Leu Arg Val Val Ala Met Ala Asn Ser Ala Leu Asn Pro Phe Val Tyr
 340 345 350
 Leu Phe Phe Gln Ala Gly Asp Cys Arg Leu Arg Arg Gln Leu Arg Lys
 355 360 365
 Arg Leu Gly Ser Leu Cys Cys Ala Pro Gln Gly Gly Ala Glu Asp Glu
 370 375 380
 Glu Gly Pro Arg Gly His Gln Ala Leu Tyr Arg Gln Arg Trp Pro His
 385 390 395 400

Pro His Tyr His His Ala Arg Arg Glu Pro Leu Asp Glu Gly Gly Leu
 405 410 415

Arg Pro Pro Pro Pro Arg Pro Arg Pro Leu Pro Cys Ser Cys Glu Ser
 420 425 430

Ala Phe

<210> 23
 <211> 1041
 <212> DNA
 <213> Homo sapiens

<400> 23
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 tgcttcacaca tgaagacctg gaagcccagc actgtttacc ttttcaattt ggccgtggct 180
 gatttcctcc ttatgatctg cctgcctttt cggacagact attacctcag acgtagacac 240
 tgggctttttg gggacattcc ctgccgagtg gggctcttca cgttggccat gaacagggcc 300
 gggagcatcg tgctccttac ggtggtggct gcggacaggt atttcaaagt ggtccacccc 360
 caccacgcgg tgaacactat ctccaccggg gtggcggtcg gcatcgctcg caccctgtgg 420
 gccctggtea tcctgggaac agtgtatctt ttgtgggaga accatctctg cgtgcaagag 480
 acggccgtct cctgtgagag cttcatcatg gagtcggcca atggctggca tgacatcatg 540
 ttccagctgg agttctttat gccctcggc atcatcttat tttgctcctt caagattggt 600
 tggagcctga ggcggaggca gcagctggcc agacaggctc ggatgaagaa ggcgaccgg 660
 ttcacatcatg tgggtggcaat tgtgttcac acatgctacc tgcccagcgt gtctgctaga 720
 ctctatttcc tctggacggt gccctcgagt gcctgcgac cctctgtcca tggggccctg 780
 cacataaccc tcagcttcac ctacatgaac agcatgctgg atcccctggt gtattatattt 840
 tcaagcccct cttttcccaa attctacaac aagctcaaaa tctgcagtct gaaacccaag 900
 cagccaggac actcaaaaac acaaaggccg gaagagatgc caatttcgaa cctcggtcgc 960
 aggagttagc tcagtgtggc aaatagtttc caaagccagt ctgatgggca atgggatccc 1020
 cacattgttg agtggcactg a 1041

<210> 24
 <211> 346
 <212> PRT
 <213> Homo sapiens

<400> 24

Met Tyr Asn Gly Ser Cys Cys Arg Ile Glu Gly Asp Thr Ile Ser Gln
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Val Met Pro Pro Leu Leu Ile Val Ala Phe Val Leu Gly Ala Leu Gly
 20 25 30

Asn Gly Val Ala Leu Cys Gly Phe Cys Phe His Met Lys Thr Trp Lys
 35 40 45
 Pro Ser Thr Val Tyr Leu Phe Asn Leu Ala Val Ala Asp Phe Leu Leu
 50 55 60
 Met Ile Cys Leu Pro Phe Arg Thr Asp Tyr Tyr Leu Arg Arg Arg His
 65 70 75 80
 Trp Ala Phe Gly Asp Ile Pro Cys Arg Val Gly Leu Phe Thr Leu Ala
 85 90 95
 Met Asn Arg Ala Gly Ser Ile Val Phe Leu Thr Val Val Ala Ala Asp
 100 105 110
 Arg Tyr Phe Lys Val Val His Pro His His Ala Val Asn Thr Ile Ser
 115 120 125
 Thr Arg Val Ala Ala Gly Ile Val Cys Thr Leu Trp Ala Leu Val Ile
 130 135 140
 Leu Gly Thr Val Tyr Leu Leu Leu Glu Asn His Leu Cys Val Gln Glu
 145 150 155 160
 Thr Ala Val Ser Cys Glu Ser Phe Ile Met Glu Ser Ala Asn Gly Trp
 165 170 175
 His Asp Ile Met Phe Gln Leu Glu Phe Phe Met Pro Leu Gly Ile Ile
 180 185 190
 Leu Phe Cys Ser Phe Lys Ile Val Trp Ser Leu Arg Arg Arg Gln Gln
 195 200 205
 Leu Ala Arg Gln Ala Arg Met Lys Lys Ala Thr Arg Phe Ile Met Val
 210 215 220
 Val Ala Ile Val Phe Ile Thr Cys Tyr Leu Pro Ser Val Ser Ala Arg
 225 230 235 240
 Leu Tyr Phe Leu Trp Thr Val Pro Ser Ser Ala Cys Asp Pro Ser Val
 245 250 255
 His Gly Ala Leu His Ile Thr Leu Ser Phe Thr Tyr Met Asn Ser Met
 260 265 270
 Leu Asp Pro Leu Val Tyr Tyr Phe Ser Ser Pro Ser Phe Pro Lys Phe
 275 280 285
 Tyr Asn Lys Leu Lys Ile Cys Ser Leu Lys Pro Lys Gln Pro Gly His
 290 295 300
 Ser Lys Thr Gln Arg Pro Glu Glu Met Pro Ile Ser Asn Leu Gly Arg
 305 310 315 320
 Arg Ser Cys Ile Ser Val Ala Asn Ser Phe Gln Ser Gln Ser Asp Gly
 325 330 335
 Gln Trp Asp Pro His Ile Val Glu Trp His
 340 345

<210> 25
 <211> 1011
 <212> DNA
 <213> Homo sapiens

<400> 25
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atttaccatcc tcctttgtat tgttggtgtt tttggaaaca ctctctctca atggatatct 120
ttaacaaaaa taggtaaaaa aacatcaacg cacatctacc tgtcacacct tgtgactgca 180
aacttacttg tgtgcagtgc catgcctttc atgagtatct atttcctgaa aggtttccaa 240
tggaataatc aatctgctca atgcagagtg gtcaattttc tgggaactct atccatgcat 300
gcaagtatgt ttgtcagtct cttaatttta agttggattg ccataagccg ctatgctacc 360
ttaatgcaaa aggattcctc gcaagagact acttcagtct atgagaaaat attttatggc 420
catttactga aaaaatttcg ccagcccaac tttgctagaa aactatgcat ttacatatgg 480
ggagtgttac tgggcataat cattccagtt accgtatact actcagtcac agaggctaca 540
gaaggagaag agagcctatg ctacaatcgg cagatggaac taggagccat gatctctcag 600
attgcaggtc tcattggaac cacatttatt ggattttcct ttttagtagt actaacatca 660
tactactctt ttgtaagcca tctgagaaaa ataagaacct gtacgtccat tatggagaaa 720
gatttgactt acagttctgt gaaaagacat cttttgggtc tccagattct actaatagtt 780
tgcttccttc cttatagtat ttttaaacc cttttttatg ttctacacca aagagataac 840
tgtcagcaat tgaattattt aatagaaaca aaaaacattc tcacctgtct tgcttcggcc 900
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tataatctct ttacaaagtc taattcagca catatgcaat catatggttg a 1011

<210> 26
<211> 336
<212> PRT
<213> Homo sapiens

<400> 26

Met	Asn	Asn	Asn	Thr	Thr	Cys	Ile	Gln	Pro	Ser	Met	Ile	Ser	Ser	Met
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Ala	Leu	Pro	Ile	Ile	Tyr	Ile	Leu	Leu	Cys	Ile	Val	Gly	Val	Phe	Gly
			20					25					30		
Asn	Thr	Leu	Ser	Gln	Trp	Ile	Phe	Leu	Thr	Lys	Ile	Gly	Lys	Lys	Thr
		35				40						45			
Ser	Thr	His	Ile	Tyr	Leu	Ser	His	Leu	Val	Thr	Ala	Asn	Leu	Leu	Val
	50					55					60				
Cys	Ser	Ala	Met	Pro	Phe	Met	Ser	Ile	Tyr	Phe	Leu	Lys	Gly	Phe	Gln
65					70					75					80
Trp	Glu	Tyr	Gln	Ser	Ala	Gln	Cys	Arg	Val	Val	Asn	Phe	Leu	Gly	Thr
			85					90						95	
Leu	Ser	Met	His	Ala	Ser	Met	Phe	Val	Ser	Leu	Leu	Ile	Leu	Ser	Trp
			100					105						110	

Ile Ala Ile Ser Arg Tyr Ala Thr Leu Met Gln Lys Asp Ser Ser Gln
 115 120 125
 Glu Thr Thr Ser Cys Tyr Glu Lys Ile Phe Tyr Gly His Leu Leu Lys
 130 135 140
 Lys Phe Arg Gln Pro Asn Phe Ala Arg Lys Leu Cys Ile Tyr Ile Trp
 145 150 155 160
 Gly Val Val Leu Gly Ile Ile Ile Pro Val Thr Val Tyr Tyr Ser Val
 165 170 175
 Ile Glu Ala Thr Glu Gly Glu Glu Ser Leu Cys Tyr Asn Arg Gln Met
 180 185 190
 Glu Leu Gly Ala Met Ile Ser Gln Ile Ala Gly Leu Ile Gly Thr Thr
 195 200 205
 Phe Ile Gly Phe Ser Phe Leu Val Val Leu Thr Ser Tyr Tyr Ser Phe
 210 215 220
 Val Ser His Leu Arg Lys Ile Arg Thr Cys Thr Ser Ile Met Glu Lys
 225 230 235 240
 Asp Leu Thr Tyr Ser Ser Val Lys Arg His Leu Leu Val Ile Gln Ile
 245 250 255
 Leu Leu Ile Val Cys Phe Leu Pro Tyr Ser Ile Phe Lys Pro Ile Phe
 260 265 270
 Tyr Val Leu His Gln Arg Asp Asn Cys Gln Gln Leu Asn Tyr Leu Ile
 275 280 285
 Glu Thr Lys Asn Ile Leu Thr Cys Leu Ala Ser Ala Arg Ser Ser Thr
 290 295 300
 Asp Pro Ile Ile Phe Leu Leu Asp Lys Thr Phe Lys Lys Thr Leu
 305 310 315 320
 Tyr Asn Leu Phe Thr Lys Ser Asn Ser Ala His Met Gln Ser Tyr Gly
 325 330 335

<210> 27
 <211> 1014
 <212> DNA
 <213> Homo sapiens

<400> 27
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 ggcattatct tctcgtggg atttccaggc aatgcagtag tgatatccac ttacattttc 180
 aaaatgagac cttggaagag cagcaccatc attatgctga acctggcctg cacagatctg 240
 ctgtatctga ccagcctccc ctctctgatt cactactatg ccagtggcga aaactggatc 300
 tttggagatt tcatgtgtaa gtttatccgc ttcagcttcc atttcaacct gtatagcagc 360
 atcctcttcc tcaactgttt cagcatcttc cgctactgtg tgatcattca ccaatgagc 420
 tgcttttcca ttcacaaaac tcgatgtgca gttgtagcct gtgctgtggt gtggatcatt 480
 tcactggtag ctgtcattcc gatgacctc ttgatcacat caaccaacag gaccaacaga 540

tcagcctgtc tcgacctcac cagttcggat gaactcaata ctattaagtg gtacaacctg 600
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 attatccaca ctctgaccca tggactgcaa actgacagct gccttaagca gaaagcacga 720
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 agggtcattc ggatcgaatc tcgcctgctt tcaatcagtt gttccattga gaatcagatc 840
 catgaagctt acatcgtht tagaccatta gctgctctga acacctttgg taacctgtta 900
 ctatatgtgg tggtcagcga caactttcag caggctgtct gctcaacagt gagatgcaaa 960
 gtaagcggga accttgagca agcaaagaaa attagttact caaacaaccc ttga 1014

<210> 28
 <211> 337
 <212> PRT
 <213> Homo sapiens

<400> 28

Met Asn Glu Pro Leu Asp Tyr Leu Ala Asn Ala Ser Asp Phe Pro Asp
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 Tyr Ala Ala Ala Phe Gly Asn Cys Thr Asp Glu Asn Ile Pro Leu Lys
 20 25 30
 Met His Tyr Leu Pro Val Ile Tyr Gly Ile Ile Phe Leu Val Gly Phe
 35 40 45
 Pro Gly Asn Ala Val Val Ile Ser Thr Tyr Ile Phe Lys Met Arg Pro
 50 55 60
 Trp Lys Ser Ser Thr Ile Ile Met Leu Asn Leu Ala Cys Thr Asp Leu
 65 70 75 80
 Leu Tyr Leu Thr Ser Leu Pro Phe Leu Ile His Tyr Tyr Ala Ser Gly
 85 90 95
 Glu Asn Trp Ile Phe Gly Asp Phe Met Cys Lys Phe Ile Arg Phe Ser
 100 105 110
 Phe His Phe Asn Leu Tyr Ser Ser Ile Leu Phe Leu Thr Cys Phe Ser
 115 120 125
 Ile Phe Arg Tyr Cys Val Ile Ile His Pro Met Ser Cys Phe Ser Ile
 130 135 140
 His Lys Thr Arg Cys Ala Val Val Ala Cys Ala Val Val Trp Ile Ile
 145 150 155 160
 Ser Leu Val Ala Val Ile Pro Met Thr Phe Leu Ile Thr Ser Thr Asn
 165 170 175
 Arg Thr Asn Arg Ser Ala Cys Leu Asp Leu Thr Ser Ser Asp Glu Leu
 180 185 190
 Asn Thr Ile Lys Trp Tyr Asn Leu Ile Leu Thr Ala Thr Thr Phe Cys
 195 200 205
 Leu Pro Leu Val Ile Val Thr Leu Cys Tyr Thr Thr Ile Ile His Thr

210

215

220

Leu Thr His Gly Leu Gln Thr Asp Ser Cys Leu Lys Gln Lys Ala Arg
 225 230 235 240

Arg Leu Thr Ile Leu Leu Leu Leu Ala Phe Tyr Val Cys Phe Leu Pro
 245 250 255

Phe His Ile Leu Arg Val Ile Arg Ile Glu Ser Arg Leu Leu Ser Ile
 260 265 270

Ser Cys Ser Ile Glu Asn Gln Ile His Glu Ala Tyr Ile Val Ser Arg
 275 280 285

Pro Leu Ala Ala Leu Asn Thr Phe Gly Asn Leu Leu Leu Tyr Val Val
 290 295 300

Val Ser Asp Asn Phe Gln Gln Ala Val Cys Ser Thr Val Arg Cys Lys
 305 310 315 320

Val Ser Gly Asn Leu Glu Gln Ala Lys Lys Ile Ser Tyr Ser Asn Asn
 325 330 335

Pro

<210> 29
 <211> 993
 <212> DNA
 <213> Homo sapiens

<400> 29
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 gccctggctg ggctggtagg aaacgggttt gtgctctggc tctgggctt ccgcatgcgc 180
 aggaacgcct tctctgtcta cgctctcagc ctggccgggg ccgacttctt cttctctgc 240
 ttccagatta taaattgcct ggtgtacctc agtaacttct tctgttccat ctccatcaat 300
 ttccctagct tcttcaccac tgtgatgacc tgtgcctacc ttgcaggcct gagcatgctg 360
 agcaccgtca gcaccgagcg ctgcctgtcc gtctgtggc ccactctgga tcgctgccgc 420
 cgccccagac acctgtcagc ggtcgtgtgt gtctgtctt gggccctgtc cctactgctg 480
 agcatcttgg aagggaagtt ctgtggcttc ttatttagtg atggtgactc tggttggtgt 540
 cagacatttg atttcatcac tgcagcgtgg ctgatttttt tattcatggt tctctgtggg 600
 tccagtctgg cctgtctggt caggatcctc tgtggctcca ggggtctgcc actgaccagg 660
 ctgtacctga ccactctgct cacagtgtgt gtgttctctc tctgcggcct gccctttggc 720
 attcagtgtt tctaatatt atggatctgg aaggattctg atgtcttatt ttgtcatatt 780
 catccagttt cagttgtcct gtcactctct aacagcagtg ccaaccccat catttacttc 840
 ttctgtgggt ctttttaggaa gcagtggcgg ctgcagcagc cgatcctcaa gctggctctc 900
 cagagggctc tgcaggacat tgctgaggtg gatcacagtg aaggatgctt ccgtcagggc 960
 accccggaga tgctgagaag cagtctggtg tag 993

<210> 30
 <211> 330
 <212> PRT
 <213> Homo sapiens

<400> 30

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Met Asp Pro Thr Thr Pro Ala Trp Gly Thr Glu Ser Thr Thr Val Asn
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Gly Asn Asp Gln Ala Leu Leu Leu Leu Cys Gly Lys Glu Thr Leu Ile
20      25      30
Pro Val Phe Leu Ile Leu Phe Ile Ala Leu Val Gly Leu Val Gly Asn
35      40      45
Gly Phe Val Leu Trp Leu Leu Gly Phe Arg Met Arg Arg Asn Ala Phe
50      55      60
Ser Val Tyr Val Leu Ser Leu Ala Gly Ala Asp Phe Leu Phe Leu Cys
65      70      75      80
Phe Gln Ile Ile Asn Cys Leu Val Tyr Leu Ser Asn Phe Phe Cys Ser
85      90      95
Ile Ser Ile Asn Phe Pro Ser Phe Phe Thr Thr Val Met Thr Cys Ala
100     105     110
Tyr Leu Ala Gly Leu Ser Met Leu Ser Thr Val Ser Thr Glu Arg Cys
115     120     125
Leu Ser Val Leu Trp Pro Ile Trp Tyr Arg Cys Arg Arg Pro Arg His
130     135     140
Leu Ser Ala Val Val Cys Val Leu Leu Trp Ala Leu Ser Leu Leu Leu
145     150     155     160
Ser Ile Leu Glu Gly Lys Phe Cys Gly Phe Leu Phe Ser Asp Gly Asp
165     170     175
Ser Gly Trp Cys Gln Thr Phe Asp Phe Ile Thr Ala Ala Trp Leu Ile
180     185     190
Phe Leu Phe Met Val Leu Cys Gly Ser Ser Leu Ala Leu Leu Val Arg
195     200     205
Ile Leu Cys Gly Ser Arg Gly Leu Pro Leu Thr Arg Leu Tyr Leu Thr
210     215     220
Ile Leu Leu Thr Val Leu Val Phe Leu Leu Cys Gly Leu Pro Phe Gly
225     230     235     240
Ile Gln Trp Phe Leu Ile Leu Trp Ile Trp Lys Asp Ser Asp Val Leu
245     250     255
Phe Cys His Ile His Pro Val Ser Val Val Leu Ser Ser Leu Asn Ser
260     265     270
Ser Ala Asn Pro Ile Ile Tyr Phe Phe Val Gly Ser Phe Arg Lys Gln
275     280     285
Trp Arg Leu Gln Gln Pro Ile Leu Lys Leu Ala Leu Gln Arg Ala Leu
290     295     300

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Gln Asp Ile Ala Glu Val Asp His Ser Glu Gly Cys Phe Arg Gln Gly
 305 310 315 320

Thr Pro Glu Met Ser Arg Ser Ser Leu Val
 325 330

<210> 31
 <211> 1092
 <212> DNA
 <213> Homo sapiens

<400> 31
 atgggccccg gcgaggcgct gctggcgggt ctctgtgtga tggtagctggc cgtggcgctg 60
 ctatccaacg cactggtgct gctttgttgc gcctacagcg ctgagctccg cactcgagcc 120
 tcaggcgctcc tcctggtgaa tctgtcgtcg ggccacctgc tgctggcggc gctggacatg 180
 cccttcacgc tgctcggtgt gatgcgcggg cggacaccgt cggcgcccgg cgcattgcaa 240
 gtcattggct tcctggacac ctctctggcg tccaacgcgg cgctgagcgt ggcggcgctg 300
 agcgagagacc agtggctggc agtgggcttc cactgagcgt acgcccagcg cctgcgaccg 360
 cgctatgccg gcctgctgct gggctgtgcc tggggacagt cgctggcctt ctcaggcgct 420
 gcacttggct gctcgtggct tggctacagc agcgcttcg cgtcctgttc gctgcgcctg 480
 ccgcccagac ctgagcgctc gcgcttcgca gccttcaccg ccacgctcca tgccgtgggc 540
 ttcgtgctgc cgctggcggg gctctgcctc acctcgctcc aggtgcaccg ggtggcacgc 600
 agccactgcc agcgcatgga caccgtcacc atgaaggcgc tcgcgctgct cgccgacctg 660
 caccacagtg tgccgagcg ctgcctcctc cagcagaagc ggcgccgcca ccgcccacc 720
 aggaagattg gcattgctat tgcgaccttc ctcatctgct ttgccccgta tgtcatgacc 780
 aggtggtggc agctcgtgcc ctctgcacc gtgaacgccc agtggggcat cctcagcaag 840
 tgctgacct acagcaaggc ggtggccgac ccgttcacgt actctctgct ccgcccggcg 900
 ttccgccaag tcctggccgg catggtgcac cggtgctga agagaacccc gcgcccagca 960
 tccacccatg acagctctct ggatgtggcc ggcatggtgc accagctgct gaagagaacc 1020
 ccgcccagc cgtccacca caacggctct gtggacacag agaatgattc ctgcctgcag 1080
 cagacacact ga 1092

<210> 32
 <211> 363
 <212> PRT
 <213> Homo sapiens

<400> 32

Met Gly Pro Gly Glu Ala Leu Leu Ala Gly Leu Leu Val Met Val Leu
 1 5 10 15

Ala Val Ala Leu Leu Ser Asn Ala Leu Val Leu Leu Cys Cys Ala Tyr
 20 25 30

Ser Ala Glu Leu Arg Thr Arg Ala Ser Gly Val Leu Leu Val Asn Leu
 35 40 45
 Ser Leu Gly His Leu Leu Leu Ala Ala Leu Asp Met Pro Phe Thr Leu
 50 55 60
 Leu Gly Val Met Arg Gly Arg Thr Pro Ser Ala Pro Gly Ala Cys Gln
 65 70 75 80
 Val Ile Gly Phe Leu Asp Thr Phe Leu Ala Ser Asn Ala Ala Leu Ser
 85 90 95
 Val Ala Ala Leu Ser Ala Asp Gln Trp Leu Ala Val Gly Phe Pro Leu
 100 105 110
 Arg Tyr Ala Gly Arg Leu Arg Pro Arg Tyr Ala Gly Leu Leu Leu Gly
 115 120 125
 Cys Ala Trp Gly Gln Ser Leu Ala Phe Ser Gly Ala Ala Leu Gly Cys
 130 135 140
 Ser Trp Leu Gly Tyr Ser Ser Ala Phe Ala Ser Cys Ser Leu Arg Leu
 145 150 155 160
 Pro Pro Glu Pro Glu Arg Pro Arg Phe Ala Ala Phe Thr Ala Thr Leu
 165 170 175
 His Ala Val Gly Phe Val Leu Pro Leu Ala Val Leu Cys Leu Thr Ser
 180 185 190
 Leu Gln Val His Arg Val Ala Arg Ser His Cys Gln Arg Met Asp Thr
 195 200 205
 Val Thr Met Lys Ala Leu Ala Leu Leu Ala Asp Leu His Pro Ser Val
 210 215 220
 Arg Gln Arg Cys Leu Ile Gln Gln Lys Arg Arg Arg His Arg Ala Thr
 225 230 235 240
 Arg Lys Ile Gly Ile Ala Ile Ala Thr Phe Leu Ile Cys Phe Ala Pro
 245 250 255
 Tyr Val Met Thr Arg Leu Ala Glu Leu Val Pro Phe Val Thr Val Asn
 260 265 270
 Ala Gln Trp Gly Ile Leu Ser Lys Cys Leu Thr Tyr Ser Lys Ala Val
 275 280 285
 Ala Asp Pro Phe Thr Tyr Ser Leu Leu Arg Arg Pro Phe Arg Gln Val
 290 295 300
 Leu Ala Gly Met Val His Arg Leu Leu Lys Arg Thr Pro Arg Pro Ala
 305 310 315 320
 Ser Thr His Asp Ser Ser Leu Asp Val Ala Gly Met Val His Gln Leu
 325 330 335
 Leu Lys Arg Thr Pro Arg Pro Ala Ser Thr His Asn Gly Ser Val Asp
 340 345 350
 Thr Glu Asn Asp Ser Cys Leu Gln Gln Thr His
 355 360

<210> 33
 <211> 1125

<212> DNA
<213> Homo sapiens

<400> 33
atgccacac tcaatacttc tgcctctcca cccacattct tctgggcca tgcctccgga 60
ggcagtgtgc tgagtgtga tgatgtccg atgcctgtca aattcctagc cctgaggctc 120
atggttgccc tggcctatgg gcttgtggg gccattggct tgctgggaaa tttggcggtg 180
ctgtgggtac tgagtaactg tgcccggaga gcccttgcc caccttcaga caccttcgtc 240
ttcaacctgg ctctggcgga cctgggactg gcactcactc tccccttttg ggcagccgag 300
tcggcactgg actttcactg gcccttcgga ggtgccctct gcaagatggt tctgacggcc 360
actgtcctca acgtctatgc cagcatcttc ctcatcacag cgctgagcgt tgctcgtac 420
tgggtggtgg ccatggctgc ggggccaggc acccacctct cactcttctg ggcccgaata 480
gccaccctgg cagtgtgggc ggcggctgcc ctggtgacgg tgcccacagc tgtcttcggg 540
gtggagggtg aggtgtgtgg tgtgcgcctt tgctgtctgc gtttccccag caggtactgg 600
ctgggggcct accagctgca gaggggtggtg ctggctttca tggtgccctt gggcgtcac 660
accaccagct acctgtgtgt gctggccttc ctgcagcggc ggcaacggcg gcggcaggac 720
agcagggtcg tggcccgcgc tgccgcacac ctggtggcctt ccttcttcct ctgctggttt 780
cccaaccatg tggtcactct ctgggggtgc ctggtgaagt ttgacctggt gccctggaac 840
agtactttct atactatcca gacgtatgtc ttccctgtca ctacttgctt ggcacacagc 900
aatagctgcc tcaaccctgt gctgtactgt ctctgaggc gggagccccg gcaggctctg 960
gcaggcacct tcagggatct gcggtcgagg ctgtggcccc agggcgagg ctgggtgcaa 1020
caggtggccc taaagcaggt aggcaggcgg tgggtcgcaa gcaacccccg ggagagccgc 1080
ccttctaccc tgctcaccaa cctggacaga gggacaccgc ggtga 1125

<210> 34
<211> 374
<212> PRT
<213> Homo sapiens

<400> 34
Met Pro Thr Leu Asn Thr Ser Ala Ser Pro Pro Thr Phe Phe Trp Ala
1 5 10 15
Asn Ala Ser Gly Gly Ser Val Leu Ser Ala Asp Asp Ala Pro Met Pro
20 25 30
Val Lys Phe Leu Ala Leu Arg Leu Met Val Ala Leu Ala Tyr Gly Leu
35 40 45
Val Gly Ala Ile Gly Leu Leu Gly Asn Leu Ala Val Leu Trp Val Leu
50 55 60
Ser Asn Cys Ala Arg Arg Ala Pro Gly Pro Pro Ser Asp Thr Phe Val
65 70 75 80

Phe Asn Leu Ala Leu Ala Asp Leu Gly Leu Ala Leu Thr Leu Pro Phe
 85 90 95
 Trp Ala Ala Glu Ser Ala Leu Asp Phe His Trp Pro Phe Gly Gly Ala
 100 105 110
 Leu Cys Lys Met Val Leu Thr Ala Thr Val Leu Asn Val Tyr Ala Ser
 115 120 125
 Ile Phe Leu Ile Thr Ala Leu Ser Val Ala Arg Tyr Trp Val Val Ala
 130 135 140
 Met Ala Ala Gly Pro Gly Thr His Leu Ser Leu Phe Trp Ala Arg Ile
 145 150 155 160
 Ala Thr Leu Ala Val Trp Ala Ala Ala Ala Leu Val Thr Val Pro Thr
 165 170 175
 Ala Val Phe Gly Val Glu Gly Glu Val Cys Gly Val Arg Leu Cys Leu
 180 185 190
 Leu Arg Phe Pro Ser Arg Tyr Trp Leu Gly Ala Tyr Gln Leu Gln Arg
 195 200 205
 Val Val Leu Ala Phe Met Val Pro Leu Gly Val Ile Thr Thr Ser Tyr
 210 215 220
 Leu Leu Leu Leu Ala Phe Leu Gln Arg Arg Gln Arg Arg Arg Gln Asp
 225 230 235 240
 Ser Arg Val Val Ala Arg Ser Val Arg Ile Leu Val Ala Ser Phe Phe
 245 250 255
 Leu Cys Trp Phe Pro Asn His Val Val Thr Leu Trp Gly Val Leu Val
 260 265 270
 Lys Phe Asp Leu Val Pro Trp Asn Ser Thr Phe Tyr Thr Ile Gln Thr
 275 280 285
 Tyr Val Phe Pro Val Thr Thr Cys Leu Ala His Ser Asn Ser Cys Leu
 290 295 300
 Asn Pro Val Leu Tyr Cys Leu Leu Arg Arg Glu Pro Arg Gln Ala Leu
 305 310 315 320
 Ala Gly Thr Phe Arg Asp Leu Arg Ser Arg Leu Trp Pro Gln Gly Gly
 325 330 335
 Gly Trp Val Gln Gln Val Ala Leu Lys Gln Val Gly Arg Arg Trp Val
 340 345 350
 Ala Ser Asn Pro Arg Glu Ser Arg Pro Ser Thr Leu Leu Thr Asn Leu
 355 360 365
 Asp Arg Gly Thr Pro Gly
 370

<210> 35
 <211> 1092
 <212> DNA
 <213> Homo sapiens

<400> 35.
 atgaatcggc accatctgca ggatcacttt ctggaaatag acaagaagaa ctgctgtgtg

60

ttccgagatg acttcattgt caaggtgttg ccgccggtgt tggggctgga gtttatcttc 120
 gggcttcttg gcaatggcct tgccctgttg attttctgtt tccacctcaa gtccctggaaa 180
 tccagccgga ttttctgtt caacctggca gtggctgact ttctactgat catctgcctg 240
 cccttctga tggacaacta tgtgaggcgt tgggactgga agtttgggga catcccttgc 300
 cggctgatgc tcttcatgtt ggctatgaac cgccagggca gcatcatctt cctcacggtg 360
 gtggcggtag acaggatattt ccgggtgttc catccccacc acgccctgaa caagatctcc 420
 aatcggacag cagccatcat ctcttgctt ctgtggggca tcaactattg cctgacagtc 480
 cacctcctga agaagaagat gccgatccag aatggcggtg caaatttgtg cagcagcttc 540
 agcatctgcc ataccttcca gtggcacgaa gccatgttcc tcttgagatt ctctctgccc 600
 ctgggcatca tctgttctg ctgagccaga attatctgga gcctgcggca gagacaaatg 660
 gaccggcatg ccaagatcaa gagagccatc accttcatca tgggtgtggc catcgtcttt 720
 gtcactgtct tccttcccag cgtggttgtg cggatccgca tcttctggct cctgcacact 780
 tcgggcacgc agaattgtga agtgtaccgc tcgggtggacc tggcgttctt tactactctc 840
 agcttcacct acatgaacag catgctggac cccgtggtgt actacttctc cagcccatcc 900
 tttcccaact tcttctccac tttgatcaac cgctgcctcc agaggaagat gacaggtgag 960
 ccagataata accgcagcac gagcgtcgag ctacagggg accccaacaa aaccagaggc 1020
 gctccagagg cgttaatggc caactccggt gagccatgga gccctctta tctgggcccc 1080
 acctctctt aa 1092

<210> 36
 <211> 363
 <212> PRT
 <213> Homo sapiens

<400> 36

Met	Asn	Arg	His	His	Leu	Gln	Asp	His	Phe	Leu	Glu	Ile	Asp	Lys	Lys
1				5					10					15	
Asn	Cys	Cys	Val	Phe	Arg	Asp	Asp	Phe	Ile	Val	Lys	Val	Leu	Pro	Pro
			20					25					30		
Val	Leu	Gly	Leu	Glu	Phe	Ile	Phe	Gly	Leu	Leu	Gly	Asn	Gly	Leu	Ala
		35					40					45			
Leu	Trp	Ile	Phe	Cys	Phe	His	Leu	Lys	Ser	Trp	Lys	Ser	Ser	Arg	Ile
	50					55					60				
Phe	Leu	Phe	Asn	Leu	Ala	Val	Ala	Asp	Phe	Leu	Leu	Ile	Ile	Cys	Leu
65					70				75					80	
Pro	Phe	Leu	Met	Asp	Asn	Tyr	Val	Arg	Arg	Trp	Asp	Trp	Lys	Phe	Gly
				85				90						95	
Asp	Ile	Pro	Cys	Arg	Leu	Met	Leu	Phe	Met	Leu	Ala	Met	Asn	Arg	Gln
			100				105						110		

Gly Ser Ile Ile Phe Leu Thr Val Val Ala Val Asp Arg Tyr Phe Arg
 115 120 125
 Val Val His Pro His His Ala Leu Asn Lys Ile Ser Asn Arg Thr Ala
 130 135 140
 Ala Ile Ile Ser Cys Leu Leu Trp Gly Ile Thr Ile Gly Leu Thr Val
 145 150 155 160
 His Leu Leu Lys Lys Lys Met Pro Ile Gln Asn Gly Gly Ala Asn Leu
 165 170 175
 Cys Ser Ser Phe Ser Ile Cys His Thr Phe Gln Trp His Glu Ala Met
 180 185 190
 Phe Leu Leu Glu Phe Phe Leu Pro Leu Gly Ile Ile Leu Phe Cys Ser
 195 200 205
 Ala Arg Ile Ile Trp Ser Leu Arg Gln Arg Gln Met Asp Arg His Ala
 210 215 220
 Lys Ile Lys Arg Ala Ile Thr Phe Ile Met Val Val Ala Ile Val Phe
 225 230 235 240
 Val Ile Cys Phe Leu Pro Ser Val Val Val Arg Ile Arg Ile Phe Trp
 245 250 255
 Leu Leu His Thr Ser Gly Thr Gln Asn Cys Glu Val Tyr Arg Ser Val
 260 265 270
 Asp Leu Ala Phe Phe Ile Thr Leu Ser Phe Thr Tyr Met Asn Ser Met
 275 280 285
 Leu Asp Pro Val Val Tyr Tyr Phe Ser Ser Pro Ser Phe Pro Asn Phe
 290 295 300
 Phe Ser Thr Leu Ile Asn Arg Cys Leu Gln Arg Lys Met Thr Gly Glu
 305 310 315 320
 Pro Asp Asn Asn Arg Ser Thr Ser Val Glu Leu Thr Gly Asp Pro Asn
 325 330 335
 Lys Thr Arg Gly Ala Pro Glu Ala Leu Met Ala Asn Ser Gly Glu Pro
 340 345 350
 Trp Ser Pro Ser Tyr Leu Gly Pro Thr Ser Pro
 355 360

<210> 37
 <211> 1044
 <212> DNA
 <213> Homo sapiens

<400> 37
 atgggggatg agctggcacc ttgccctgtg ggcactacag cttggccggc cctgatccag 60
 ctcatcagca agacaccctg catgccccaa gcagccagca acatttcctt gggcctgggg 120
 gacctcaggg tgcccagctc catgctgtac tggcttttcc ttccctcaag cctgctggct 180
 gcagccacac tggctgtcag cccctgtctg ctggtgacca tcctgcggaa ccaacggctg 240
 cgacaggagc cccactacct gctccgggct aacatcctgc tctcagacct ggctacatt 300
 ctctccaca tgctcatctc ctccagcagc ctgggtggct gggagctggg ccgcatggcc 360

tgtggcattc tcaactgatgc tgtcttcgcc gcctgcacca gcaccatcct gtccttcacc 420
 gccattgtgc tgcacaccta cctggcagtc atccatccac tgcgctacct ctccttcattg 480
 tcccatgggg ctgcctggaa ggcagtggcc ctcatctggc tggtgacctg ctgcttcccc 540
 acattcctta tttggctcag caagtggcag gatgcccagc tggaggagca aggagcttca 600
 tacatcctac caccaagcat gggcaccacg ccgggatgtg gcctcctggt cattgttacc 660
 tacacctcca ttctgtgctg tctgttcctc tgcacagctc tcattgcaa ctgtttctgg 720
 aggatctatg cagaggccaa gacttcaggc atctgggggc agggctattc ccgggccagg 780
 ggcaccctgc tgatccactc agtgctgac acattgtacg tgagcacagg ggtgggtgttc 840
 tccctggaca tgggtctgac caggtaccac cacattgact ctgggactca cacatggctc 900
 ctggcagcta acagtgaggt actcatgatg ctccccctg ccatgctccc atacctgtac 960
 ctgctccgct accggcagct gttgggcatg gtccggggcc acctcccatc caggaggcac 1020
 caggccatct ttaccatttc ctag 1044

<210> 38
 <211> 347
 <212> PRT
 <213> Homo sapiens

<400> 38

Met	Gly	Asp	Glu	Leu	Ala	Pro	Cys	Pro	Val	Gly	Thr	Thr	Ala	Trp	Pro
1			5						10					15	
Ala	Leu	Ile	Gln	Leu	Ile	Ser	Lys	Thr	Pro	Cys	Met	Pro	Gln	Ala	Ala
			20					25					30		
Ser	Asn	Thr	Ser	Leu	Gly	Leu	Gly	Asp	Leu	Arg	Val	Pro	Ser	Ser	Met
		35					40					45			
Leu	Tyr	Trp	Leu	Phe	Leu	Pro	Ser	Ser	Leu	Leu	Ala	Ala	Ala	Thr	Leu
	50					55					60				
Ala	Val	Ser	Pro	Leu	Leu	Leu	Val	Thr	Ile	Leu	Arg	Asn	Gln	Arg	Leu
65					70					75					80
Arg	Gln	Glu	Pro	His	Tyr	Leu	Leu	Pro	Ala	Asn	Ile	Leu	Leu	Ser	Asp
				85					90					95	
Leu	Ala	Tyr	Ile	Leu	Leu	His	Met	Leu	Ile	Ser	Ser	Ser	Ser	Leu	Gly
			100					105						110	
Gly	Trp	Glu	Leu	Gly	Arg	Met	Ala	Cys	Gly	Ile	Leu	Thr	Asp	Ala	Val
		115					120					125			
Phe	Ala	Ala	Cys	Thr	Ser	Thr	Ile	Leu	Ser	Phe	Thr	Ala	Ile	Val	Leu
	130						135					140			
His	Thr	Tyr	Leu	Ala	Val	Ile	His	Pro	Leu	Arg	Tyr	Leu	Ser	Phe	Met
145					150					155					160
Ser	His	Gly	Ala	Ala	Trp	Lys	Ala	Val	Ala	Leu	Ile	Trp	Leu	Val	Ala
			165						170					175	

Cys Cys Phe Pro Thr Phe Leu Ile Trp Leu Ser Lys Trp Gln Asp Ala
 180 185 190
 Gln Leu Glu Glu Gln Gly Ala Ser Tyr Ile Leu Pro Pro Ser Met Gly
 195 200 205
 Thr Gln Pro Gly Cys Gly Leu Leu Val Ile Val Thr Tyr Thr Ser Ile
 210 215 220
 Leu Cys Val Leu Phe Leu Cys Thr Ala Leu Ile Ala Asn Cys Phe Trp
 225 230 235 240
 Arg Ile Tyr Ala Glu Ala Lys Thr Ser Gly Ile Trp Gly Gln Gly Tyr
 245 250 255
 Ser Arg Ala Arg Gly Thr Leu Leu Ile His Ser Val Leu Ile Thr Leu
 260 265 270
 Tyr Val Ser Thr Gly Val Val Phe Ser Leu Asp Met Val Leu Thr Arg
 275 280 285
 Tyr His His Ile Asp Ser Gly Thr His Thr Trp Leu Leu Ala Ala Asn
 290 295 300
 Ser Glu Val Leu Met Met Leu Pro Arg Ala Met Leu Pro Tyr Leu Tyr
 305 310 315 320
 Leu Leu Arg Tyr Arg Gln Leu Leu Gly Met Val Arg Gly His Leu Pro
 325 330 335
 Ser Arg Arg His Gln Ala Ile Phe Thr Ile Ser
 340 345

<210> 39
 <211> 1023
 <212> DNA
 <213> Homo sapiens

<400> 39
 atgaatccat ttcatgcatc ttgttggaac acctctgccg aacttttaaa caaatcctgg 60
 aataaagagt ttgcttatca aactgccagt gtggtagata cagtcacacct cccttccatg 120
 attgggatta tctgttcaac agggctgggt ggcaacatcc tcattgtatt cactataata 180
 agatccagga aaaaaacagt ccctgacatc tatatctgca acctggctgt ggctgatttg 240
 gtccacatag ttggaatgcc ttttcttatt caccaatggg ccgaggggg agagtgggtg 300
 tttggggggc ctctctgcac catcatcaca tccctggata cttgtaacca atttgcctgt 360
 agtgccatca tgactgtaat gagtgtggac aggtactttg cctcgtcca accatttcga 420
 ctgacacggt ggagaacaag gtacaagacc atccgatca atttgggcct ttgggcagct 480
 tcctttatcc tggcattgcc tgtctgggtc tactcgaagg tcatcaaatt taaagacggt 540
 gttgagagtt gtgcttttga ttgacatcc cctgacgatg tactctggta tacactttat 600
 ttgacgataa caactttttt tttccctcta cccttgattt tgggtgtgcta tattttaatt 660
 ttatgctata cttgggagat gtatcaacag aataaggatg ccagatgctg caatcccagt 720
 gtacaaaac agagagtgat gaagttgaca aagatggtgc tgggtgctggt ggtagtcttt 780

atcctgagtg ctgcccctta tcatgtgata caactggtga acttacagat ggaacagccc 840
 aactggcct tctatgtggg ttattacctc tccatctgtc tcagctatgc cagcagcagc 900
 attaaccctt ttctctacat cctgctgagt ggaaatttcc agaaacgtct gcctcaaate 960
 caaagaagag cgactgagaa ggaaatcaac aatatgggaa acactctgaa atcacacttt 1020
 tag 1023

<210> 40
 <211> 340
 <212> PRT
 <213> Homo sapiens

<400> 40

Met Asn Pro Phe His Ala Ser Cys Trp Asn Thr Ser Ala Glu Leu Leu
 1 5 10 15
 Asn Lys Ser Trp Asn Lys Glu Phe Ala Tyr Gln Thr Ala Ser Val Val
 20 25 30
 Asp Thr Val Ile Leu Pro Ser Met Ile Gly Ile Ile Cys Ser Thr Gly
 35 40 45
 Leu Val Gly Asn Ile Leu Ile Val Phe Thr Ile Ile Arg Ser Arg Lys
 50 55 60
 Lys Thr Val Pro Asp Ile Tyr Ile Cys Asn Leu Ala Val Ala Asp Leu
 65 70 75 80
 Val His Ile Val Gly Met Pro Phe Leu Ile His Gln Trp Ala Arg Gly
 85 90 95
 Gly Glu Trp Val Phe Gly Gly Pro Leu Cys Thr Ile Ile Thr Ser Leu
 100 105 110
 Asp Thr Cys Asn Gln Phe Ala Cys Ser Ala Ile Met Thr Val Met Ser
 115 120 125
 Val Asp Arg Tyr Phe Ala Leu Val Gln Pro Phe Arg Leu Thr Arg Trp
 130 135 140
 Arg Thr Arg Tyr Lys Thr Ile Arg Ile Asn Leu Gly Leu Trp Ala Ala
 145 150 155 160
 Ser Phe Ile Leu Ala Leu Pro Val Trp Val Tyr Ser Lys Val Ile Lys
 165 170 175
 Phe Lys Asp Gly Val Glu Ser Cys Ala Phe Asp Leu Thr Ser Pro Asp
 180 185 190
 Asp Val Leu Trp Tyr Thr Leu Tyr Leu Thr Ile Thr Thr Phe Phe Phe
 195 200 205
 Pro Leu Pro Leu Ile Leu Val Cys Tyr Ile Leu Ile Leu Cys Tyr Thr
 210 215 220
 Trp Glu Met Tyr Gln Gln Asn Lys Asp Ala Arg Cys Cys Asn Pro Ser
 225 230 235 240
 Val Pro Lys Gln Arg Val Met Lys Leu Thr Lys Met Val Leu Val Leu

245 250 255
 Val Val Val Phe Ile Leu Ser Ala Ala Pro Tyr His Val Ile Gln Leu
 260 265 270
 Val Asn Leu Gln Met Glu Gln Pro Thr Leu Ala Phe Tyr Val Gly Tyr
 275 280 285
 Tyr Leu Ser Ile Cys Leu Ser Tyr Ala Ser Ser Ser Ile Asn Pro Phe
 290 295 300
 Leu Tyr Ile Leu Leu Ser Gly Asn Phe Gln Lys Arg Leu Pro Gln Ile
 305 310 315 320
 Gln Arg Arg Ala Thr Glu Lys Glu Ile Asn Asn Met Gly Asn Thr Leu
 325 330 335
 Lys Ser His Phe
 340

<210> 41
 <211> 24
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc_feature
 <223> Novel Sequence

<400> 41
 cttgcagaca tcaccatggc agcc

24

<210> 42
 <211> 24
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc_feature
 <223> Novel Sequence

<400> 42
 gtgatgctct gagtactgga ctgg

24

<210> 43
 <211> 20
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc_feature
 <223> Novel Sequence

<400> 43
 gaagctgtga agagtgatgc

20

<210> 44
 <211> 24
 <212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

<223> Novel Sequence

<400> 44

gtcagcaata ttgataagca gcag

24

<210> 45

<211> 27

<212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

<223> Novel Sequence

<400> 45

ccatggggaa cgattctgtc agctacg

27

<210> 46

<211> 24

<212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

<223> Novel Sequence

<400> 46

gctatgcctg aagccagtct tgtg

24

<210> 47

<211> 26

<212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

<223> Novel Sequence

<400> 47

ccaggatgtt gtgtcaccgt ggtggc

26

<210> 48

<211> 26

<212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

<223> Novel Sequence

<400> 48

cacagcgctg cagccctgca gctggc

26

<210> 49
<211> 26
<212> DNA
<213> Artificial Sequence

<220>
<221> misc_feature
<223> Novel Sequence

<400> 49
cttcctctcg tagggatgaa ccagac

26

<210> 50
<211> 26
<212> DNA
<213> Artificial Sequence

<220>
<221> misc_feature
<223> Novel Sequence

<400> 50
ctcgcacagg tgggaagcac ctgtgg

26

<210> 51
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
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<400> 51
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23

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<400> 52
catatccctc cgagtgtcca gcggc

25

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<223> Novel Sequence

<400> 53
gcatggagag aaaatttatg tccttgcaac c 31

<210> 54
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<220>
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<223> Novel Sequence

<400> 54
caagaacagg tctcatctaa gagctcc 27

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<400> 55
gctgttgcca tgacgtccac ctgcac 26

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<400> 56
ggacagttca aggtttgcct tagaac 26

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<400> 57
ctttcgatac tgctcctatg etc 23

<210> 58
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<400> 58
gtagtccact gaaagtccag tgatcc

26

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<400> 59
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<400> 60
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<220>
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<400> 61
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<400> 62

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28

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<400> 63
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22

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<400> 64
gtctgcttgt tcagtgccac tcaac

25

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<210> 66
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<400> 66
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<210> 68
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<400> 68
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<210> 69
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<400> 69
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<210> 70
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<400> 70
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<210> 71
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<220>
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<400> 71
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<210> 72

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<400> 72
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<223> Novel Sequence

<400> 73
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27

<210> 74
<211> 27
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<400> 74
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27

<210> 75
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<400> 75
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22

<210> 76
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<400> 76
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24

<210> 77

<211> 24
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<400> 77
agccatccct gccaggaagc atgg

24

<210> 78
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<400> 78
ccagactgtg gactcaagaa ctctagg

27

<210> 79
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<220>
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<223> Novel Sequence

<400> 79
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28

<210> 80
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<212> DNA
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<221> misc_feature
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<400> 80
atcatgtcta gactcatggt gatcc

25

<210> 81
<211> 30
<212> DNA
<213> Artificial Sequence

<220>
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<400> 81
ggggagggaa agcaaaggtg gtcctcctgg

30

<210> 82
<211> 30
<212> DNA
<213> Artificial Sequence

<220>
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<223> Novel Sequence

<400> 82
ccaggagaac cacctttgct ttccctcccc

30

<210> 83
<211> 1356
<212> DNA
<213> Homo sapiens

<400> 83
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tcggaatctg tgccctctt cttcatgctc ctgctggact tgactgctgt ggctggcaat 180
gccgctgtga tggccgtgat cgccaagacg cctgccctcc gaaaatttgt cttcgtcttc 240
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tactattacg tagtccaccc catgcgtac gaggtgcgca tgacgctggg gctggtgccc 480
tctgtgctgg tgggtgtgtg ggtgaaggcc ttggccatgg cttctgtgcc agtgttggga 540
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ctcgcgtcctg ccgcctcacc ccggctggag tcatga 1356

<210> 84
<211> 451
<212> PRT
<213> Homo sapiens

<400> 84

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Gly Arg Val Pro Gln Thr Pro Gly Pro Ser Thr Ala Ser Gly Val Pro
20 25 30
Glu Val Gly Leu Arg Asp Val Ala Ser Glu Ser Val Ala Leu Phe Phe
35 40 45
Met Leu Leu Leu Asp Leu Thr Ala Val Ala Gly Asn Ala Ala Val Met
50 55 60
Ala Val Ile Ala Lys Thr Pro Ala Leu Arg Lys Phe Val Phe Val Phe
65 70 75 80
His Leu Cys Leu Val Asp Leu Leu Ala Ala Leu Thr Leu Met Pro Leu
85 90 95
Ala Met Leu Ser Ser Ser Ala Leu Phe Asp His Ala Leu Phe Gly Glu
100 105 110
Val Ala Cys Arg Leu Tyr Leu Phe Leu Ser Val Cys Phe Val Ser Leu
115 120 125
Ala Ile Leu Ser Val Ser Ala Ile Asn Val Glu Arg Tyr Tyr Tyr Val
130 135 140
Val His Pro Met Arg Tyr Glu Val Arg Met Thr Leu Gly Leu Val Ala
145 150 155 160
Ser Val Leu Val Gly Val Trp Val Lys Ala Leu Ala Met Ala Ser Val
165 170 175
Pro Val Leu Gly Arg Val Ser Trp Glu Glu Gly Ala Pro Ser Val Pro
180 185 190
Pro Gly Cys Ser Leu Gln Trp Ser His Ser Ala Tyr Cys Gln Leu Phe
195 200 205
Val Val Val Phe Ala Val Leu Tyr Phe Leu Leu Pro Leu Leu Leu Ile
210 215 220
Leu Val Val Tyr Cys Ser Met Phe Arg Val Ala Arg Val Ala Ala Met
225 230 235 240
Gln His Gly Pro Leu Pro Thr Trp Met Glu Thr Pro Arg Gln Arg Ser
245 250 255
Glu Ser Leu Ser Ser Arg Ser Thr Met Val Thr Ser Ser Gly Ala Pro
260 265 270
Gln Thr Thr Pro His Arg Thr Phe Gly Gly Gly Lys Ala Lys Val Val

275	280	285
Leu Leu Ala Val Gly Gly Gln Phe Leu Leu Cys Trp Leu Pro Tyr Phe 290 295 300		
Ser Phe His Leu Tyr Val Ala Leu Ser Ala Gln Pro Ile Ser Thr Gly 305 310 315 320		
Gln Val Glu Ser Val Val Thr Trp Ile Gly Tyr Phe Cys Phe Thr Ser 325 330 335		
Asn Pro Phe Phe Tyr Gly Cys Leu Asn Arg Gln Ile Arg Gly Glu Leu 340 345 350		
Ser Lys Gln Phe Val Cys Phe Phe Lys Pro Ala Pro Glu Glu Glu Leu 355 360 365		
Arg Leu Pro Ser Arg Glu Gly Ser Ile Glu Glu Asn Phe Leu Gln Phe 370 375 380		
Leu Gln Gly Thr Gly Cys Pro Ser Glu Ser Trp Val Ser Arg Pro Leu 385 390 395 400		
Pro Ser Pro Lys Gln Glu Pro Pro Ala Val Asp Phe Arg Ile Pro Gly 405 410 415		
Gln Ile Ala Glu Glu Thr Ser Glu Phe Leu Glu Gln Gln Leu Thr Ser 420 425 430		
Asp Ile Ile Met Ser Asp Ser Tyr Leu Arg Pro Ala Ala Ser Pro Arg 435 440 445		
Leu Glu Ser 450		

<210> 85
 <211> 28
 <212> DNA
 <213> Homo sapiens

<400> 85
 caggaaggca aagaccacca tcatcatc

28

<210> 86
 <211> 28
 <212> DNA
 <213> Homo sapiens

<400> 86
 gatgatgatg gtggtctttg ccttcctg

28

<210> 87
 <211> 1041
 <212> DNA
 <213> Homo sapiens

<400> 87
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 at
 ggcaccttca gcaataacaa cagcaggaac tgcacaattg aaaacttcaa gagagaattt
 ttcccaattg tatatctgat aatatttttc tggggagtct tgggaaatgg gttgtccata

60

120

180

tatgttttcc tgcagcctta taagaagtcc acatctgtga acgttttcat gctaaatctg	240
gccatttcag atctcctgtt cataagcacg cttcccttca gggctgacta ttatcttaga	300
ggctccaatt ggatatttgg agacctggcc tgcaggatta tgtcttattc cttgtatgtc	360
aacatgtaca gcagtattta tttcctgacc gtgctgagtg ttgtgcgttt cctggcaatg	420
gttcaccctt ttöggcttct gcatgtcacc agcatcagga gtgcctggat cctctgtggg	480
atcatatgga tccttatcat ggcttcctca ataatgtctc tggacagtgg ctctgagcag	540
aacggcagtg tcacatcatg cttagagctg aatctctata aaattgctaa gctgcagacc	600
atgaactata ttgccttggg ggtgggctgc ctgctgccat ttttcacact cagcatctgt	660
tatctgctga tcattcgggt tctgttaaaa gtggaggtcc cagaatcggg gctgcggggt	720
tctcacagga aggcaaagac caccatcatc atcaccttga tcattcttct cttgtgtttc	780
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gacagactgc ataaagcttt ggttatcaca ctggccttgg cagcagccaa tgcctgcttc	900
aatcctctgc tctattactt tgctggggag aattttaagg acagactaaa gtctgcactc	960
agaaaaggcc atccacagaa ggcaaagaca aagtgtgttt tccctgttag tgtgtggttg	1020
agaaaggaaa caagagtata a	1041

<210> 88
 <211> 346
 <212> PRT
 <213> Homo sapiens

<400> 88

Met	Glu	Arg	Lys	Phe	Met	Ser	Leu	Gln	Pro	Ser	Ile	Ser	Val	Ser	Glu	1	5	10	15
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Ile	Glu	Asn	Phe	Lys	Arg	Glu	Phe	Phe	Pro	Ile	Val	Tyr	Leu	Ile	Ile	35	40	45	
Phe	Phe	Trp	Gly	Val	Leu	Gly	Asn	Gly	Leu	Ser	Ile	Tyr	Val	Phe	Leu	50	55	60	
Gln	Pro	Tyr	Lys	Lys	Ser	Thr	Ser	Val	Asn	Val	Phe	Met	Leu	Asn	Leu	65	70	75	80
Ala	Ile	Ser	Asp	Leu	Leu	Phe	Ile	Ser	Thr	Leu	Pro	Phe	Arg	Ala	Asp	85	90	95	
Tyr	Tyr	Leu	Arg	Gly	Ser	Asn	Trp	Ile	Phe	Gly	Asp	Leu	Ala	Cys	Arg	100	105	110	
Ile	Met	Ser	Tyr	Ser	Leu	Tyr	Val	Asn	Met	Tyr	Ser	Ser	Ile	Tyr	Phe	115	120	125	
Leu	Thr	Val	Leu	Ser	Val	Val	Arg	Phe	Leu	Ala	Met	Val	His	Pro	Phe	130	135	140	

Arg Leu Leu His Val Thr Ser Ile Arg Ser Ala Trp Ile Leu Cys Gly
 145 150 155 160
 Ile Ile Trp Ile Leu Ile Met Ala Ser Ser Ile Met Leu Leu Asp Ser
 165 170 175
 Gly Ser Glu Gln Asn Gly Ser Val Thr Ser Cys Leu Glu Leu Asn Leu
 180 185 190
 Tyr Lys Ile Ala Lys Leu Gln Thr Met Asn Tyr Ile Ala Leu Val Val
 195 200 205
 Gly Cys Leu Leu Pro Phe Phe Thr Leu Ser Ile Cys Tyr Leu Leu Ile
 210 215 220
 Ile Arg Val Leu Leu Lys Val Glu Val Pro Glu Ser Gly Leu Arg Val
 225 230 235 240
 Ser His Arg Lys Ala Lys Thr Thr Ile Ile Ile Thr Leu Ile Ile Phe
 245 250 255
 Phe Leu Cys Phe Leu Pro Tyr His Thr Leu Arg Thr Val His Leu Thr
 260 265 270
 Thr Trp Lys Val Gly Leu Cys Lys Asp Arg Leu His Lys Ala Leu Val
 275 280 285
 Ile Thr Leu Ala Leu Ala Ala Ala Asn Ala Cys Phe Asn Pro Leu Leu
 290 295 300
 Tyr Tyr Phe Ala Gly Glu Asn Phe Lys Asp Arg Leu Lys Ser Ala Leu
 305 310 315 320
 Arg Lys Gly His Pro Gln Lys Ala Lys Thr Lys Cys Val Phe Pro Val
 325 330 335
 Ser Val Trp Leu Arg Lys Glu Thr Arg Val
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<210> 89
 <211> 28
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc_feature
 <223> Novel Sequence

<400> 89
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28

<210> 90
 <211> 28
 <212> DNA
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<220>
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 <223> Novel Sequence

<400> 90
 gaagatcact ttcttagctt tgactgg

28

<210> 91
 <211> 1527
 <212> DNA
 <213> Homo sapiens

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 ttctctgccc cctctttcgt cggcaacata gtgctggcgc tagtggtgca gcgcaagccg 180
 cagctgctgc aggtgaccaa ccgtttttatc tttaacctcc tcgtcaccga cctgctgcag 240
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 tccaagatga cccagcgccg cggttacctg ctctctatg gcacctggat tgtggccatc 480
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 tcctacgatt ctgctacttt tccttga 1527

<210> 92
 <211> 508
 <212> PRT
 <213> Homo sapiens

<400> 92

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Thr Cys Met Pro Leu Ser Lys Met Pro Ile Ser Leu Ala His Gly Ile
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Ile Arg Ser Thr Val Leu Val Ile Phe Leu Ala Ala Ser Phe Val Gly
35 40 45
Asn Ile Val Leu Ala Leu Val Leu Gln Arg Lys Pro Gln Leu Leu Gln
50 55 60
Val Thr Asn Arg Phe Ile Phe Asn Leu Leu Val Thr Asp Leu Leu Gln
65 70 75 80
Ile Ser Leu Val Ala Pro Trp Val Val Ala Thr Ser Val Pro Leu Phe
85 90 95
Trp Pro Leu Asn Ser His Phe Cys Thr Ala Leu Val Ser Leu Thr His
100 105 110
Leu Phe Ala Phe Ala Ser Val Asn Thr Ile Val Val Val Ser Val Asp
115 120 125
Arg Tyr Leu Ser Ile Ile His Pro Leu Ser Tyr Pro Ser Lys Met Thr
130 135 140
Gln Arg Arg Gly Tyr Leu Leu Leu Tyr Gly Thr Trp Ile Val Ala Ile
145 150 155 160
Leu Gln Ser Thr Pro Pro Leu Tyr Gly Trp Gly Gln Ala Ala Phe Asp
165 170 175
Glu Arg Asn Ala Leu Cys Ser Met Ile Trp Gly Ala Ser Pro Ser Tyr
180 185 190
Thr Ile Leu Ser Val Val Ser Phe Ile Val Ile Pro Leu Ile Val Met
195 200 205
Ile Ala Cys Tyr Ser Val Val Phe Cys Ala Ala Arg Arg Gln His Ala
210 215 220
Leu Leu Tyr Asn Val Lys Arg His Ser Leu Glu Val Arg Val Lys Asp
225 230 235 240
Cys Val Glu Asn Glu Asp Glu Glu Gly Ala Glu Lys Lys Glu Glu Phe
245 250 255
Gln Asp Glu Ser Glu Phe Arg Arg Gln His Glu Gly Glu Val Lys Ala
260 265 270
Lys Glu Gly Arg Met Glu Ala Lys Asp Gly Ser Leu Lys Ala Lys Glu
275 280 285
Gly Ser Thr Gly Thr Ser Glu Ser Ser Val Glu Ala Arg Gly Ser Glu
290 295 300
Glu Val Arg Glu Ser Ser Thr Val Ala Ser Asp Gly Ser Met Glu Gly
305 310 315 320
Lys Glu Gly Ser Thr Lys Val Glu Glu Asn Ser Met Lys Ala Asp Lys
325 330 335

Gly Arg Thr Glu Val Asn Gln Cys Ser Ile Asp Leu Gly Glu Asp Asp
 340 345 350
 Met Glu Phe Gly Glu Asp Asp Ile Asn Phe Ser Glu Asp Asp Val Glu
 355 360 365
 Ala Val Asn Ile Pro Glu Ser Leu Pro Pro Ser Arg Arg Asn Ser Asn
 370 375 380
 Ser Asn Pro Pro Leu Pro Arg Cys Tyr Gln Cys Lys Ala Lys Lys Val
 385 390 395 400
 Ile Phe Ile Ile Ile Phe Ser Tyr Val Leu Ser Leu Gly Pro Tyr Cys
 405 410 415
 Phe Leu Ala Val Leu Ala Val Trp Val Asp Val Glu Thr Gln Val Pro
 420 425 430
 Gln Trp Val Ile Thr Ile Ile Ile Trp Leu Phe Phe Leu Gln Cys Cys
 435 440 445
 Ile His Pro Tyr Val Tyr Gly Tyr Met His Lys Thr Ile Lys Lys Glu
 450 455 460
 Ile Gln Asp Met Leu Lys Lys Phe Phe Cys Lys Glu Lys Pro Pro Lys
 465 470 475 480
 Glu Asp Ser His Pro Asp Leu Pro Gly Thr Glu Gly Gly Thr Glu Gly
 485 490 495
 Lys Ile Val Pro Ser Tyr Asp Ser Ala Thr Phe Pro
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<210> 93
 <211> 29
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc feature
 <223> Novel Sequence

<400> 93
 gccgccaccg cgccaagagg aagattggc

29

<210> 94
 <211> 29
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc feature
 <223> Novel Sequence

<400> 94
 gccaatcttc ctcttggcgc ggtggcggc

29

<210> 95
 <211> 1092
 <212> DNA

<213> Homo sapiens

<400> 95

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tcaggcgctc tcctggtgaa tctgtcgtcg ggccacctgc tgcctggcggc gctggacatg      180
cccttcacgc tgctcggtgt gatgcgcggg cggacaccgt cggcgcccgg cgcattgcaa      240
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aggaagattg gcattgctat tgcgaccttc ctcatctgct ttgccccgta tgcattgacc      780
aggctggcgg agctcgtgcc ctctgtcacc gtgaacgcc agaaggcat cctcagcaag      840
tgcttgacct acagcaaggc ggtggccgac ccgttcacgt actctctgct ccgcccggcg      900
ttccgccaag tcctggccgg catggtgcac cggctgctga agagaacccc gcgccagca      960
tccacccatg acagctctct ggatgtggc ggcatggtgc accagctgct gaagagaacc     1020
ccgcgccag cgccaccaca caacggctct gtggacacag agaatgattc ctgcctgcag     1080
cagacacact ga                                                                1092
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<210> 96

<211> 363

<212> PRT

<213> Homo sapiens

<400> 96

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Ala Val Ala Leu Leu Ser Asn Ala Leu Val Leu Leu Cys Cys Ala Tyr
20          25          30
Ser Ala Glu Leu Arg Thr Arg Ala Ser Gly Val Leu Leu Val Asn Leu
35          40          45
Ser Leu Gly His Leu Leu Leu Ala Ala Leu Asp Met Pro Phe Thr Leu
50          55          60
Leu Gly Val Met Arg Gly Arg Thr Pro Ser Ala Pro Gly Ala Cys Gln
65          70          75          80
Val Ile Gly Phe Leu Asp Thr Phe Leu Ala Ser Asn Ala Ala Leu Ser
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85	90	95
Val Ala Ala Leu Ser Ala Asp Gln Trp Leu Ala Val Gly Phe Pro Leu 100 105 110		
Arg Tyr Ala Gly Arg Leu Arg Pro Arg Tyr Ala Gly Leu Leu Leu Gly 115 120 125		
Cys Ala Trp Gly Gln Ser Leu Ala Phe Ser Gly Ala Ala Leu Gly Cys 130 135 140		
Ser Trp Leu Gly Tyr Ser Ser Ala Phe Ala Ser Cys Ser Leu Arg Leu 145 150 155 160		
Pro Pro Glu Pro Glu Arg Pro Arg Phe Ala Ala Phe Thr Ala Thr Leu 165 170 175		
His Ala Val Gly Phe Val Leu Pro Leu Ala Val Leu Cys Leu Thr Ser 180 185 190		
Leu Gln Val His Arg Val Ala Arg Ser His Cys Gln Arg Met Asp Thr 195 200 205		
Val Thr Met Lys Ala Leu Ala Leu Ala Asp Leu His Pro Ser Val 210 215 220		
Arg Gln Arg Cys Leu Ile Gln Gln Lys Arg Arg Arg His Arg Ala Thr 225 230 235 240		
Arg Lys Ile Gly Ile Ala Ile Ala Thr Phe Leu Ile Cys Phe Ala Pro 245 250 255		
Tyr Val Met Thr Arg Leu Ala Glu Leu Val Pro Phe Val Thr Val Asn 260 265 270		
Ala Gln Lys Gly Ile Leu Ser Lys Cys Leu Thr Tyr Ser Lys Ala Val 275 280 285		
Ala Asp Pro Phe Thr Tyr Ser Leu Leu Arg Arg Pro Phe Arg Gln Val 290 295 300		
Leu Ala Gly Met Val His Arg Leu Leu Lys Arg Thr Pro Arg Pro Ala 305 310 315 320		
Ser Thr His Asp Ser Ser Leu Asp Val Ala Gly Met Val His Gln Leu 325 330 335		
Leu Lys Arg Thr Pro Arg Pro Ala Ser Thr His Asn Gly Ser Val Asp 340 345 350		
Thr Glu Asn Asp Ser Cys Leu Gln Gln Thr His 355 360		

<210> 97
 <211> 34
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc feature
 <223> Novel Sequence

<400> 97
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<210> 98
 <211> 36
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc_feature
 <223> Novel Sequence

<400> 98
 gatcgatata cgtgactcca gccggggtga ggcggc 36

<210> 99
 <211> 2610
 <212> DNA
 <213> Homo sapiens and Rat

<400> 99
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 tcggaatctg tggccctctt cttcatgctc ctgctggact tgactgctgt ggctggcaat 180
 gccgctgtga tggccgtgat cgccaagacg cctgccctcc gaaaatttgt ctctgtcttc 240
 cacctctgcc tgggtggact gctggctgcc ctgacctca tgccctggc catgctctcc 300
 agctctgcc tctttgacca cgccctcttt ggggaggtgg cctgccgcct ctacttgttt 360
 ctgagcgtgt gctttgtcag cctggccatc ctctcgtgt cagccatcaa tgtggagcgc 420
 tactattacg tagtccacc catgcgctac gaggtgcgca tgacgctggg gctggtggcc 480
 tctgtgctgg tgggtgtgtg ggtgaaggcc ttggccatgg cttctgtgcc agtgttggga 540
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 cacagtgcct actgccagct ttttgtgtg gtctttgtc tcttttactt tctgttgccc 660
 ctgctcctca tacttgttgt ctactgcagc atgttccgag tggcccgct ggctgccatg 720
 cagcacgggc cgctgccac gtggatggag acaccccggc aacgctccga atctctcagc 780
 agccgctcca cgatggtcac cagctcgggg gccccccaga ccacccaca ccggacgttt 840
 gggggaggga aagcagcagt ggttctcctg gctgtggggg gacagttcct gctctgttgg 900
 ttgcctact tctctttcca cctctatgtt gccctgagt ctcagcccat ttcaactggg 960
 cagggtggaga gtgtggtcac ctggattggc tacttttgc tcaactcaa ccctttcttc 1020
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ctccgtcctg ccgcctcacc ccggctggag tcagcgatat ctgcagaatt ccaccacact 1380
 ggactagtgg atccgagctc ggtaccaagc ttgggctgca ggtcgatggg ctgcctcggc 1440
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 atcgagaagc agctgcagaa ggacaagcag gtctaccggg ccacgcaccg cctgctgctg 1560
 ctgggtgctg gagagtctgg caaaagcacc attgtgaagc agatgaggat cctacatgtt 1620
 aatgggttta acggagaggg cggcgaagag gacccgcagg ctgcaaggag caacagcgat 1680
 ggtgagaagg ccaccaaagt gcaggacatc aaaaacaacc tgaaggaggc cattgaaacc 1740
 attgtggccg ccatgagcaa cctggtgccc cccgtggagc tggccaaccc tgagaaccag 1800
 ttcagagtgg actacattct gagcgtgatg aacgtgccaa actttgactt cccacctgaa 1860
 ttctatgagc atgccaaggc tctgtgggag gatgaggagg ttcgtgcctg ctacgagcgc 1920
 tccaacgagt accagctgat cgactgtgcc cagtacttcc tggacaagat tgatgtgatc 1980
 aagcaggccg actacgtgcc aagtgaccag gacctgcttc gctgccgct cctgacctct 2040
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 ggcagcgcg atgaacgccg caagtggatc cagtgttca atgatgtgac tgccatcatc 2160
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 ctgagaatca gcactgctag tggagatgga cgtcactact gctaccctca ctttacctgc 2520
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 atgcatcttc gccaatatga gctgctctaa 2610

<210> 100
 <211> 869
 <212> PRT
 <213> Homo sapiens and Rat

<400> 100

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Gly	Arg	Val	Pro	Gln	Thr	Pro	Gly	Pro	Ser	Thr	Ala	Ser	Gly	Val	Pro
			20					25					30		
Glu	Val	Gly	Leu	Arg	Asp	Val	Ala	Ser	Glu	Ser	Val	Ala	Leu	Phe	Phe
		35					40					45			
Met	Leu	Leu	Leu	Asp	Leu	Thr	Ala	Val	Ala	Gly	Asn	Ala	Ala	Val	Met
	50					55					60				

Ala Val Ile Ala Lys Thr Pro Ala Leu Arg Lys Phe Val Phe Val Phe
 65 70 75 80
 His Leu Cys Leu Val Asp Leu Leu Ala Ala Leu Thr Leu Met Pro Leu
 85 90 95
 Ala Met Leu Ser Ser Ser Ala Leu Phe Asp His Ala Leu Phe Gly Glu
 100 105 110
 Val Ala Cys Arg Leu Tyr Leu Phe Leu Ser Val Cys Phe Val Ser Leu
 115 120 125
 Ala Ile Leu Ser Val Ser Ala Ile Asn Val Glu Arg Tyr Tyr Tyr Val
 130 135 140
 Val His Pro Met Arg Tyr Glu Val Arg Met Thr Leu Gly Leu Val Ala
 145 150 155 160
 Ser Val Leu Val Gly Val Trp Val Lys Ala Leu Ala Met Ala Ser Val
 165 170 175
 Pro Val Leu Gly Arg Val Ser Trp Glu Glu Gly Ala Pro Ser Val Pro
 180 185 190
 Pro Gly Cys Ser Leu Gln Trp Ser His Ser Ala Tyr Cys Gln Leu Phe
 195 200 205
 Val Val Val Phe Ala Val Leu Tyr Phe Leu Leu Pro Leu Leu Leu Ile
 210 215 220
 Leu Val Val Tyr Cys Ser Met Phe Arg Val Ala Arg Val Ala Ala Met
 225 230 235 240
 Gln His Gly Pro Leu Pro Thr Trp Met Glu Thr Pro Arg Gln Arg Ser
 245 250 255
 Glu Ser Leu Ser Ser Arg Ser Thr Met Val Thr Ser Ser Gly Ala Pro
 260 265 270
 Gln Thr Thr Pro His Arg Thr Phe Gly Gly Gly Lys Ala Ala Val Val
 275 280 285
 Leu Leu Ala Val Gly Gly Gln Phe Leu Leu Cys Trp Leu Pro Tyr Phe
 290 295 300
 Ser Phe His Leu Tyr Val Ala Leu Ser Ala Gln Pro Ile Ser Thr Gly
 305 310 315 320
 Gln Val Glu Ser Val Val Thr Trp Ile Gly Tyr Phe Cys Phe Thr Ser
 325 330 335
 Asn Pro Phe Phe Tyr Gly Cys Leu Asn Arg Gln Ile Arg Gly Glu Leu
 340 345 350
 Ser Lys Gln Phe Val Cys Phe Phe Lys Pro Ala Pro Glu Glu Glu Leu
 355 360 365
 Arg Leu Pro Ser Arg Glu Gly Ser Ile Glu Glu Asn Phe Leu Gln Phe
 370 375 380
 Leu Gln Gly Thr Gly Cys Pro Ser Glu Ser Trp Val Ser Arg Pro Leu
 385 390 395 400
 Pro Ser Pro Lys Gln Glu Pro Pro Ala Val Asp Phe Arg Ile Pro Gly
 405 410 415

Gln Ile Ala Glu Glu Thr Ser Glu Phe Leu Glu Gln Gln Leu Thr Ser
 420 425 430
 Asp Ile Ile Met Ser Asp Ser Tyr Leu Arg Pro Ala Ala Ser Pro Arg
 435 440 445
 Leu Glu Ser Ala Ile Ser Ala Glu Phe His His Thr Gly Leu Val Asp
 450 455 460
 Pro Ser Ser Val Pro Ser Leu Gly Cys Arg Ser Met Gly Cys Leu Gly
 465 470 475 480
 Asn Ser Lys Thr Glu Asp Gln Arg Asn Glu Glu Lys Ala Gln Arg Glu
 485 490 495
 Ala Asn Lys Lys Ile Glu Lys Gln Leu Gln Lys Asp Lys Gln Val Tyr
 500 505 510
 Arg Ala Thr His Arg Leu Leu Leu Leu Gly Ala Gly Glu Ser Gly Lys
 515 520 525
 Ser Thr Ile Val Lys Gln Met Arg Ile Leu His Val Asn Gly Phe Asn
 530 535 540
 Gly Glu Gly Gly Glu Glu Asp Pro Gln Ala Ala Arg Ser Asn Ser Asp
 545 550 555 560
 Gly Glu Lys Ala Thr Lys Val Gln Asp Ile Lys Asn Asn Leu Lys Glu
 565 570 575
 Ala Ile Glu Thr Ile Val Ala Ala Met Ser Asn Leu Val Pro Pro Val
 580 585 590
 Glu Leu Ala Asn Pro Glu Asn Gln Phe Arg Val Asp Tyr Ile Leu Ser
 595 600 605
 Val Met Asn Val Pro Asn Phe Asp Phe Pro Pro Glu Phe Tyr Glu His
 610 615 620
 Ala Lys Ala Leu Trp Glu Asp Glu Gly Val Arg Ala Cys Tyr Glu Arg
 625 630 635 640
 Ser Asn Glu Tyr Gln Leu Ile Asp Cys Ala Gln Tyr Phe Leu Asp Lys
 645 650 655
 Ile Asp Val Ile Lys Gln Ala Asp Tyr Val Pro Ser Asp Gln Asp Leu
 660 665 670
 Leu Arg Cys Arg Val Leu Thr Ser Gly Ile Phe Glu Thr Lys Phe Gln
 675 680 685
 Val Asp Lys Val Asn Phe His Met Phe Asp Val Gly Gly Gln Arg Asp
 690 695 700
 Glu Arg Arg Lys Trp Ile Gln Cys Phe Asn Asp Val Thr Ala Ile Ile
 705 710 715 720
 Phe Val Val Ala Ser Ser Tyr Asn Met Val Ile Arg Glu Asp Asn
 725 730 735
 Gln Thr Asn Arg Leu Gln Glu Ala Leu Asn Leu Phe Lys Ser Ile Trp
 740 745 750
 Asn Asn Arg Trp Leu Arg Thr Ile Ser Val Ile Leu Phe Leu Asn Lys

755 760 765
 Gln Asp Leu Leu Ala Glu Lys Val Leu Ala Gly Lys Ser Lys Ile Glu
 770 775 780
 Asp Tyr Phe Pro Glu Phe Ala Arg Tyr Thr Thr Pro Glu Asp Ala Thr
 785 790 795 800
 Pro Glu Pro Gly Glu Asp Pro Arg Val Thr Arg Ala Lys Tyr Phe Ile
 805 810 815
 Arg Asp Glu Phe Leu Arg Ile Ser Thr Ala Ser Gly Asp Gly Arg His
 820 825 830
 Tyr Cys Tyr Pro His Phe Thr Cys Ala Val Asp Thr Glu Asn Ile Arg
 835 840 845
 Arg Val Phe Asn Asp Cys Arg Asp Ile Ile Gln Arg Met His Leu Arg
 850 855 860

Gln Tyr Glu Leu Leu
865

<210> 101
 <211> 30
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc feature
 <223> Novel Sequence

<400> 101
 tctagaatga cgtccacctg caccaacagc 30

<210> 102
 <211> 34
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc feature
 <223> Novel Sequence

<400> 102
 gatatcgcag gaaaagtagc agaatcgtag gaag 34

<210> 103
 <211> 2781
 <212> DNA
 <213> Homo Sapiens and Rat

<400> 103
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 ctctccaaaa tgcccatcag cctggccac ggcacatcc gctcaaccgt gctggttatc 120
 ttcctcgccg cctctttcgt cggcaacata gtgctggcgc tagtggtgca gcgcaagccg 180
 cagctgctgc aggtgaccaa ccgttttatc tttaacctcc tcgtcaccga cctgctgcag 240

atttcgctcg tggccccctg ggtggtggcc acctctgtgc ctctcttctg gcccctcaac	300
agccacttct gcacggccct ggttagcctc acccacctgt tcgccttcgc cagcgtcaac	360
accattgtcg tgggtgcagt ggatcgctac ttgtccatca tccacctct ctctacccg	420
tccaagatga cccagcgccg cggttacctg ctctctatg gcacctggat tgtggccatc	480
ctgcagagca ctctccact ctacggctgg ggccaggctg cctttgatga ggcgaatgct	540
ctctgtcca tgatctgggg ggccagcccc agctacacta ttctcagcgt ggtgtccttc	600
atcgtcattc cactgattgt catgattgcc tgctactccg tgggtttctg tgcagcccgg	660
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tgtgtggaga atgaggatga agagggagca gagaagaagg aggagtcca ggatgagagt	780
gagtttcgcc gccagcatga aggtgaggtc aaggccaagg agggcagaat ggaagccaag	840
gacggcagcc tgaaggccaa ggaagggaag acggggacca gtgagagtag tgtagaggcc	900
aggggcagcg aggaggtcag agagagcagc acgggtggcca gcgacggcag catggagggt	960
aaggaaggca gcaccaaagt tgaggagaac agcatgaagg cagacaaggg tcgcacagag	1020
gtcaaccagt gcagcattga cttgggtgaa gatgacatgg agtttggtga agacgacatc	1080
aatttcagtg aggatgacgt cgaggcagtg aacatcccg agagcctccc acccagtcgt	1140
cgtaacagca acagcaaccc tcctctgccc aggtgctacc agtgcaaagc tgctaaagtg	1200
atcttcatca tcattttctc ctatgtgcta tccttggggc cctactgctt tttagcagtc	1260
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tggttttct tcctgcagtg ctgcatccac ccctatgtct atggctacat gcacaagacc	1380
attaagaagg aaatccagga catgctgaag aagttcttct gcaaggaaa gccccgaaa	1440
gaagatagcc acccagacct gcccggaaca gaggtgaggc ctgaaggcaa gattgtccct	1500
tcctacgatt ctgctacttt tcctgcgata tctgcagaat tccaccacac tggactagt	1560
gatccgagct cggtagcaag cttgggctgc aggtcgatgg gctgcctcgg caacagtaag	1620
accgaggacc agcgcaacga ggagaaggcg cagcgcgagg ccaacaaaa gatcgagaag	1680
cagctgcaga aggacaagca ggtctaccgg gccacgcacc gcctgtgct gctgggtgct	1740
ggagagtctg gcaaaagcac cattgtgaag cagatgagga tcctacatgt taatgggttt	1800
aacggagagg gcggcgaaga ggacccgcag gctgcaagga gcaacagcga tggtgagaag	1860
gccaccaaag tgcaggacat caaaaacaac ctgaaggagg ccattgaaac cattgtggcc	1920
gccatgagca acctggtgcc ccccgaggag ctggccaacc ctgagaacca gttcagagt	1980
gactacattc tgagcgtgat gaacgtgcca aactttgact tccacctga attctatgag	2040
catgccaaag ctctgtggga ggatgaggga gttcgtgcct gctacgagcg ctccaacgag	2100
taccagctga tcgactgtgc ccagtacttc ctggacaaga ttgatgtgat caagcaggcc	2160
gactacgtgc caagtgacca ggacctgctt cgctgccgcg tcctgacctc tggaatcttt	2220

gagaccaagt tccaggtgga caaagtcaac ttccacatgt tccgatgtggg cggccagcgc 2280
gatgaacgcc gcaagtggat ccagtgttc aatgatgtga ctgccatcat cttcgtggtg 2340
gccagcagca gctacaacat ggtcatccgg gaggacaacc agaccaaccg tctgcaggag 2400
gctctgaacc tcttcaagag catctggaac aacagatggc tgcgtacat ctctgtgatc 2460
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cgccaatacg agctgctcta a 2781

<210> 104
<211> 926
<212> PRT
<213> Homo sapiens and Rat

<400> 104

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20 25 30
Ile Arg Ser Thr Val Leu Val Ile Phe Leu Ala Ala Ser Phe Val Gly
35 40 45
Asn Ile Val Leu Ala Leu Val Leu Gln Arg Lys Pro Gln Leu Leu Gln
50 55 60
Val Thr Asn Arg Phe Ile Phe Asn Leu Leu Val Thr Asp Leu Leu Gln
65 70 75 80
Ile Ser Leu Val Ala Pro Trp Val Val Ala Thr Ser Val Pro Leu Phe
85 90 95
Trp Pro Leu Asn Ser His Phe Cys Thr Ala Leu Val Ser Leu Thr His
100 105 110
Leu Phe Ala Phe Ala Ser Val Asn Thr Ile Val Val Val Ser Val Asp
115 120 125
Arg Tyr Leu Ser Ile Ile His Pro Leu Ser Tyr Pro Ser Lys Met Thr
130 135 140
Gln Arg Arg Gly Tyr Leu Leu Leu Tyr Gly Thr Trp Ile Val Ala Ile
145 150 155 160
Leu Gln Ser Thr Pro Pro Leu Tyr Gly Trp Gly Gln Ala Ala Phe Asp
165 170 175
Glu Arg Asn Ala Leu Cys Ser Met Ile Trp Gly Ala Ser Pro Ser Tyr
180 185 190

Thr Ile Leu Ser Val Val Ser Phe Ile Val Ile Pro Leu Ile Val Met
 195 200 205
 Ile Ala Cys Tyr Ser Val Val Phe Cys Ala Ala Arg Arg Gln His Ala
 210 215 220
 Leu Leu Tyr Asn Val Lys Arg His Ser Leu Glu Val Arg Val Lys Asp
 225 230 235 240
 Cys Val Glu Asn Glu Asp Glu Glu Gly Ala Glu Lys Lys Glu Glu Phe
 245 250 255
 Gln Asp Glu Ser Glu Phe Arg Arg Gln His Glu Gly Glu Val Lys Ala
 260 265 270
 Lys Glu Gly Arg Met Glu Ala Lys Asp Gly Ser Leu Lys Ala Lys Glu
 275 280 285
 Gly Ser Thr Gly Thr Ser Glu Ser Ser Val Glu Ala Arg Gly Ser Glu
 290 295 300
 Glu Val Arg Glu Ser Ser Thr Val Ala Ser Asp Gly Ser Met Glu Gly
 305 310 315 320
 Lys Glu Gly Ser Thr Lys Val Glu Glu Asn Ser Met Lys Ala Asp Lys
 325 330 335
 Gly Arg Thr Glu Val Asn Gln Cys Ser Ile Asp Leu Gly Glu Asp Asp
 340 345 350
 Met Glu Phe Gly Glu Asp Asp Ile Asn Phe Ser Glu Asp Asp Val Glu
 355 360 365
 Ala Val Asn Ile Pro Glu Ser Leu Pro Pro Ser Arg Arg Asn Ser Asn
 370 375 380
 Ser Asn Pro Pro Leu Pro Arg Cys Tyr Gln Cys Lys Ala Ala Lys Val
 385 390 395 400
 Ile Phe Ile Ile Ile Phe Ser Tyr Val Leu Ser Leu Gly Pro Tyr Cys
 405 410 415
 Phe Leu Ala Val Leu Ala Val Trp Val Asp Val Glu Thr Gln Val Pro
 420 425 430
 Gln Trp Val Ile Thr Ile Ile Ile Trp Leu Phe Phe Leu Gln Cys Cys
 435 440 445
 Ile His Pro Tyr Val Tyr Gly Tyr Met His Lys Thr Ile Lys Lys Glu
 450 455 460
 Ile Gln Asp Met Leu Lys Lys Phe Phe Cys Lys Glu Lys Pro Pro Lys
 465 470 475 480
 Glu Asp Ser His Pro Asp Leu Pro Gly Thr Glu Gly Gly Thr Glu Gly
 485 490 495
 Lys Ile Val Pro Ser Tyr Asp Ser Ala Thr Phe Pro Ala Ile Ser Ala
 500 505 510
 Glu Phe His His Thr Gly Leu Val Asp Pro Ser Ser Val Pro Ser Leu
 515 520 525
 Gly Cys Arg Ser Met Gly Cys Leu Gly Asn Ser Lys Thr Glu Asp Gln
 530 535 540

Arg Asn Glu Glu Lys Ala Gln Arg Glu Ala Asn Lys Lys Ile Glu Lys
 545 550 555 560
 Gln Leu Gln Lys Asp Lys Gln Val Tyr Arg Ala Thr His Arg Leu Leu
 565 570 575
 Leu Leu Gly Ala Gly Glu Ser Gly Lys Ser Thr Ile Val Lys Gln Met
 580 585 590
 Arg Ile Leu His Val Asn Gly Phe Asn Gly Glu Gly Gly Glu Glu Asp
 595 600 605
 Pro Gln Ala Ala Arg Ser Asn Ser Asp Gly Glu Lys Ala Thr Lys Val
 610 615 620
 Gln Asp Ile Lys Asn Asn Leu Lys Glu Ala Ile Glu Thr Ile Val Ala
 625 630 635 640
 Ala Met Ser Asn Leu Val Pro Pro Val Glu Leu Ala Asn Pro Glu Asn
 645 650 655
 Gln Phe Arg Val Asp Tyr Ile Leu Ser Val Met Asn Val Pro Asn Phe
 660 665 670
 Asp Phe Pro Pro Glu Phe Tyr Glu His Ala Lys Ala Leu Trp Glu Asp
 675 680 685
 Glu Gly Val Arg Ala Cys Tyr Glu Arg Ser Asn Glu Tyr Gln Leu Ile
 690 695 700
 Asp Cys Ala Gln Tyr Phe Leu Asp Lys Ile Asp Val Ile Lys Gln Ala
 705 710 715 720
 Asp Tyr Val Pro Ser Asp Gln Asp Leu Leu Arg Cys Arg Val Leu Thr
 725 730 735
 Ser Gly Ile Phe Glu Thr Lys Phe Gln Val Asp Lys Val Asn Phe His
 740 745 750
 Met Phe Asp Val Gly Gly Gln Arg Asp Glu Arg Arg Lys Trp Ile Gln
 755 760 765
 Cys Phe Asn Asp Val Thr Ala Ile Ile Phe Val Val Ala Ser Ser Ser
 770 775 780
 Tyr Asn Met Val Ile Arg Glu Asp Asn Gln Thr Asn Arg Leu Gln Glu
 785 790 795 800
 Ala Leu Asn Leu Phe Lys Ser Ile Trp Asn Asn Arg Trp Leu Arg Thr
 805 810 815
 Ile Ser Val Ile Leu Phe Leu Asn Lys Gln Asp Leu Leu Ala Glu Lys
 820 825 830
 Val Leu Ala Gly Lys Ser Lys Ile Glu Asp Tyr Phe Pro Glu Phe Ala
 835 840 845
 Arg Tyr Thr Thr Pro Glu Asp Ala Thr Pro Glu Pro Gly Glu Asp Pro
 850 855 860
 Arg Val Thr Arg Ala Lys Tyr Phe Ile Arg Asp Glu Phe Leu Arg Ile
 865 870 875 880
 Ser Thr Ala Ser Gly Asp Gly Arg His Tyr Cys Tyr Pro His Phe Thr

885

890

895

Cys Ala Val Asp Thr Glu Asn Ile Arg Arg Val Phe Asn Asp Cys Arg
 900 905 910

Asp Ile Ile Gln Arg Met His Leu Arg Gln Tyr Glu Leu Leu
 915 920 925

<210> 105
 <211> 23
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc_feature
 <223> Novel Sequence

<400> 105
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23

<210> 106
 <211> 24
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc_feature
 <223> Novel Sequence

<400> 106
 gctatgcctg aagccagtct tgtg

24

<210> 107
 <211> 25
 <212> DNA
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<220>
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<400> 107
 gcacctgctc ctgagcacct tctcc

25

<210> 108
 <211> 26
 <212> DNA
 <213> Artificial Sequence

<220>
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<400> 108
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26

<210> 109

<211> 24
<212> DNA
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<220>
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ccagtgatga ctctgtccag cctg

24

<210> 110
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<400> 114
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27

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ctctgatgcc atctgctgga ttcctg

26

<210> 116
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<400> 116
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26

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tggtggcgat ggccaacagc gctc

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<400> 119
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<210> 120
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<210> 121
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<400> 121
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<400> 122
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<400> 123
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24

<210> 124
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<400> 124
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24

<210> 125
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<400> 125
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24

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<400> 126
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<400> 127
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<210> 128
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<400> 128
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25

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<400> 129
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1 5

<210> 130
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<213> Artificial Sequence

<220>
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<400> 130
Glu Tyr Asn Leu Val
1 5

<210> 131
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<400> 131
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<210> 132

<211> 36
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<400> 132

Gly Ala Thr Cys Ala Ala Gly Cys Thr Thr Cys Cys Ala Thr Gly Gly
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Cys Gly Thr Gly Cys Thr Gly Cys Cys Thr Gly Ala Gly Cys Gly Ala
20 25 30

Gly Gly Ala Gly
35

<210> 133
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<212> PRT
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<400> 133

Gly Ala Thr Cys Gly Gly Ala Thr Cys Cys Thr Thr Ala Gly Ala Ala
1 5 10 15

Cys Ala Gly Gly Cys Cys Gly Cys Ala Gly Thr Cys Cys Thr Thr Cys
20 25 30

Ala Gly Gly Thr Thr Cys Ala Gly Cys Thr Gly Cys Ala Gly Gly Ala
35 40 45

Thr Gly Gly Thr Gly
50

